

Review Article

Erythropoietin in COVID-19-Induced Neuroinflammation; EPO Plus Losartan Might be PromisingReza Nejat^{1,2*}, Ahmad Shahr Sadr^{3,4*}, David Najafi^{5,6,7}¹Former Assistant Prof., Department of Anesthesiology and Critical Care Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran.²Former ICU Chief, Bazarganan Hospital, Tehran, Iran.³Bioinformatics Research Center, Cheragh Medical Institute and Hospital, Kabul, Afghanistan.⁴Department of Computer and Data Sciences, Faculty of Mathematical Sciences, Shahid Beheshti University, Tehran, Iran.⁵Former Assistant Clinical Professor, Department of Ophthalmology, UCSD, California, USA.⁶Vitreoretinal surgeon, Scripps and Sharp Memorial Hospitals, San Diego, California, USA.⁷Founder and President of Alliance Retinal Consultants, La Mesa, California, USA.

ARTICLE INFO

ABSTRACT

Received 14.05.2020
 Revised 01.06.2020
 Accepted 17.06.2020
 Published 20.06.2020

Key words:

COVID -19;
 SARS-CoV2;
 Calcium;
 Neuroinflammation;
 Erythropoietin;
 Ang II;
 HIF;
 papain-like protease;
 excitotoxicity

Introduction: Neuroinflammation is the inflammatory reaction in the central nervous system (CNS) provoked by diverse insults. This phenomenon results in a cascade of release of inflammatory mediators and intracellular messengers such as reactive oxygen species. The elicited responses are the cause of many neurological and neurodegenerative disorders. Erythropoietin (EPO) has been considered effective in attenuating this inflammatory process in the CNS, yet its administration in COVID-19 needs meticulously designed studies.

Discussion: Neuroinflammation in COVID-19 due to probable contribution of renin-angiotensin system dysregulation resulting in surplus of Ang II and owing to the synergistic interaction between this octapeptide and EPO needs special consideration. Both of these compounds increase intracellular Ca²⁺ which may induce release of cytokine and inflammatory mediators leading to aggravation of neuroinflammation. In addition, Ang II elevates HIF even in normoxia which by itself increases EPO. It is implicated that EPO and HIF may likely increase in patients with COVID-19 which makes administration of EPO to these patients hazardous. Furthermore, papain-like protease of SARS-CoV2 as a deubiquitinase may also increase HIF.

Conclusion: It is hypothesized that administration of EPO to patients with COVID-19-induced neuroinflammation may not be safe and in case EPO is needed for any reason in this disease adding of losartan may block AT1R-mediated post-receptor harmful effects of Ang II in synergism with EPO. Inhibition of papain-like protease might additionally decrease HIF in this disease. More in vitro, in vivo and clinical studies are needed to validate these hypotheses.

Abbreviations: ALS: amyotrophic lateral sclerosis; Ang II: angiotensin II; ARB: angiotensin receptor blocker; AT1R: angiotensin II type 1 receptor; AT2R: angiotensin II type 2 receptor; [Ca²⁺]: intracellular calcium concentration; CNS: central nervous system, DAMP: danger/damage-associated molecular patterns; EPO: erythropoietin; EPOR: erythropoietin receptor; HIF: Hypoxia-inducible factor; MAPK: mitogen-activated protein kinases; MMP: matrix metalloproteinase; MS: multiple sclerosis; NLR: NOD-like receptors; PAMP: pathogen associated molecular pattern; PRR: pattern-recognition receptors; RAGE: receptor for advanced glycation end products; ROS: reactive oxygen species; RNS: reactive nitrogen species; TBI: Traumatic brain injury; TIMP: tissue inhibitor of metalloproteinase; TLR: Toll-like receptors; TRPC: transient receptor potential canonical;

* Corresponding Authors' Email: Rezanejat@yahoo.com & a_shahirsadr@sbu.ac.ir

Introduction

Neuroinflammatory cascade mediated by cytokines, chemokines, and other inflammatory mediators including intracellular messengers like reactive oxygen species in the CNS (1) eventuates into devastatingly debilitating outcomes in a time- and severity-dependent fashion. As a repairing phenomenon in a short mild form, neuroinflammation plays a significant role in neurodevelopment, neuroprotection and neuroplasticity. However, prolongation of this process may result in lifelong disabilities (2, 3). Hypoglycemia (4), ischemic brain diseases (5, 6), intracerebral hemorrhage (7, 8), traumatic brain injury (TBI) (9, 10), Alzheimer's disease (11-13), Parkinson's disease (14, 15) and other neurodegenerative disorders such as amyotrophic lateral sclerosis (ALS) (16), and multiple sclerosis (MS) (17, 18) are accompanied by this pathology which might also be responsible for disorders such as epilepsy (19), depression (20, 21), obsessive-compulsive disorders (22) and schizophrenia (23). EPO, as an immunomodulator, has been shown to have attenuating effect on neuroinflammation irrespective to its origin (24-27). Although EPO's effect on the increase in blood pressure, platelets and probability of thrombophilia have been debated to be hazardous in patients, high doses (up to 400 units/kg) of this glycoprotein has been considered safe and effective in ameliorating neuroinflammatory responses such as seen in ischemic brain injuries in a few studies (27-33). Moreover, daily low dose of EPO (4000 units/day for a maximum of 2 weeks) was also reported to be effective in a patient with huge subdural hematoma who was at risk of brain death (34).

Pathophysiology of Neuroinflammation

Neuroinflammation triggered by a variety of stressors such as oxidative, traumatic or ischemic insults which result in activation of astrocytes as well as microglial cells initiate a cascade of inflammatory responses in the nervous system (35). Astrocytes while maintaining the homeostasis of neural tissue, simultaneously modulate neurotransmitter secretion and synaptic transmission. Astrocytic cells are morphologically and functionally of two lineage: radial glial-like and reactive astrocytes (36, 37). Clones of radial glial-like astrocytes exhibit characteristics of mitogenically active multipotent stem cells which generate neural cells and other astrocytes. (38, 39) Defending against pathogens or stressors is the responsibility of reactive astrocytes along with microglial and neural cells (40-42). In this process pattern-recognition receptors (PRRs) including Toll-like receptors (TLR), NOD-like receptors (NLRs), receptor for advanced glycation end products (RAGE), and scavenger, complement and mannose receptors play a crucial part (43-49).

Pathogen-associated molecular patterns (PAMPs) or host-derived danger/damage-associated molecular patterns (DAMPs) (heat shock proteins, ATP, S100B and HMGB) are recognized through intracellular PRRs (41, 50). Mitogen-activated protein kinases (MAPKs) and nuclear factor-kappa B (NF- κ B) signaling pathways activated by PAMPs and DAMPs induce expression of cascade of inflammatory proteins including ICAM-1, VCAM-1, E-selection, and iNOS. (51-54) The role of NF- κ B pathway as an inducing factor in neuroinflammation in Parkinson's Disease, Alzheimer's Disease and other neurodegenerative diseases as well as in traumatic and ischemic brain injuries has been described (55-61).

In a neuroinflammatory response, cytokines (IL-1 β , TNF- α , and IL-6), α -chemokines (MCP-1, MIP-1, and RANTES), and other inflammatory mediators such as cyclooxygenase-2 and matrix metalloproteinase 9 (MMP-9) are released following activation of astrocytes and microglia (62-64). MMPs are able to proteolyze the remains of extracellular matrix proteins following destruction of the brain tissue, open new routes to accommodate newly formed vessels in the injured area, regulate vascular endothelial growth factor (VEGF) release and modulate pro-inflammatory cytokines such as TNF- α (65-67). MMPs should be regulated by tissue inhibitor of metalloproteinase (TIMP), otherwise tissue destruction spreads from the inflamed core to the circumferential normal tissue, as well (68).

In neuroinflammation, blood brain barrier (BBB) is disrupted which allows the inflammatory mediators and cells to enter the brain parenchyma. Besides, as water channels, called aquaporins (AQPs) with constitutional pro-inflammatory characteristics, are dysregulated in inflammatory responses, water content of the brain parenchyma increases. This results in astrocytes swelling and migration, dysfunction of BBB, and cytokine release. (69, 70)

Amazingly, high metabolic rate and oxygen demand on one hand and the presence of vascular injury and shortage of oxygen delivery to the inflamed tissue in any type of brain injuries on the other hand, eventuates into tissue relative or absolute hypoxia in early phases (6-12 hours post-injury period) of the insult which induces hypoxia inducible factor-1 α (HIF-1 α). This factor promotes pro-apoptotic genes (BNIP3, NIX and NOXA) and upregulates caspase 3⁽⁷¹⁾.

Furthermore, free radicals (ROS, RNS) generated in the mitochondria in hypoxic milieu of the injured brain stabilize HIF-1 α (72, 73). In normoxia, insulin-like growth factor-1 (IGF-1), thyroid hormone (T₃), cytokines (IL- β , IL-6, TGF- β , TNF- α), NF- κ B, free radicals (ROS, RNS), thrombin, PAMPs and DAMPs upregulate HIF-1 α , as well (74-80). Initially, HIF-1 α release is associated with pro-apoptotic effects but after 48 hours it induces pro-survival proteomes like EPO, VEGF, glucose transporter-1, aldolase A, lactic dehydrogenase A and phosphofructokinase protein(81, 82). It is implied that in the acute phase of neuroinflammatory states accompanied by pro-apoptotic effects an urgent intervention may be of great benefit.

EPO and Neuroinflammation

Anti-inflammatory/anti-apoptotic properties of EPO have long attracted the attention of experts (83-85). In adults, this glycoprotein exerts some trophic protective effects in the brain(86, 87). Secreted or systemically administered EPO can cross the BBB slowly through extracellular pathways or transported to the brain by binding to its receptor, EPO-R (fig.1), presenting on the luminal surface of cerebral capillary endothelium (88, 89). In neuroinflammation the disrupted BBB lets this glycoprotein reach the brain parenchyma easily (89). EPO, along with EPO-R, can also be produced de novo especially in astrocytes and in the case of EPO-R, in a wide range of cells in the CNS including endothelial and neural progenitor cells (90, 91).

Proliferative effect of EPO on neural progenitor cell (NPC) in culture media has been investigated in animal studies which showed its capacity to promote differentiation of NPCs to mature neurons and oligodendrocyte in the hippocampus^(92, 93). In animal studies it was shown that

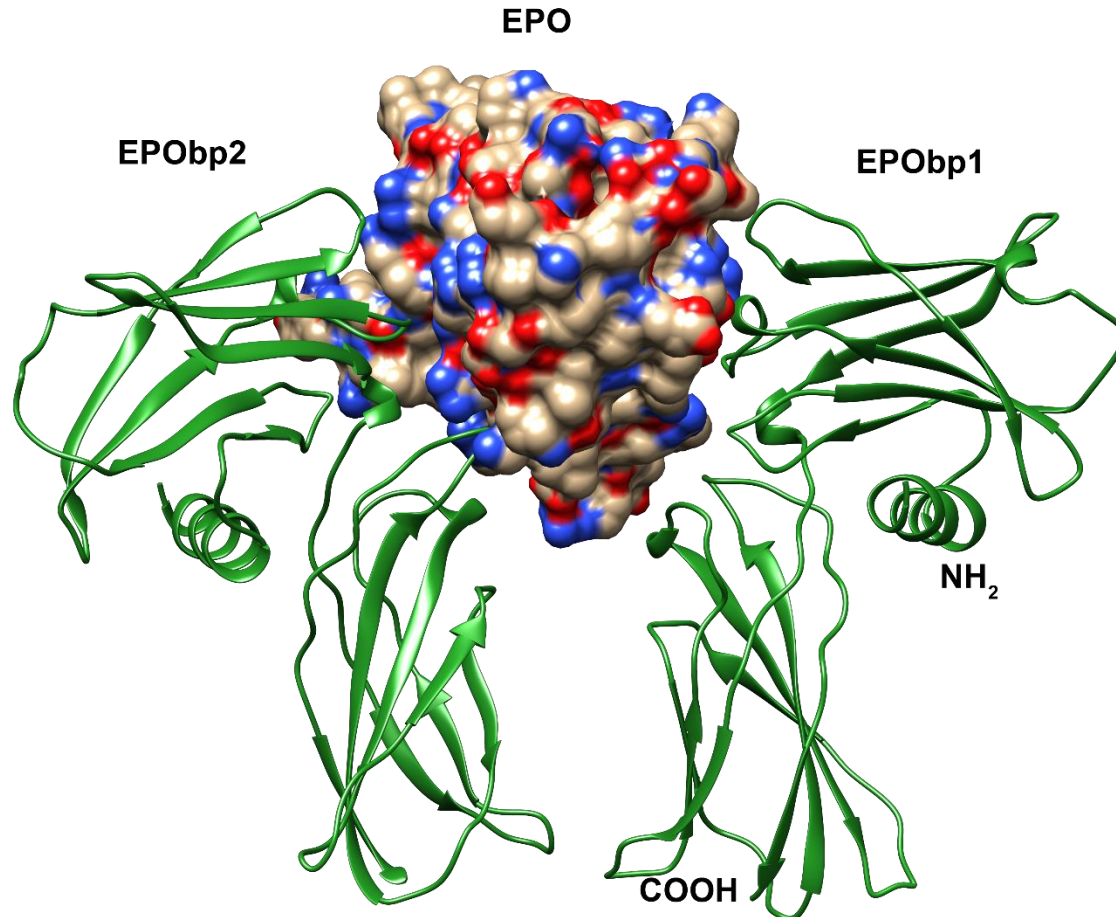


Figure 1. Cartoon overview presentation of the crystallographic structure of EPO-R in complex with surface presentation of the EPO (PDB identifier 1EER). EPO-R is shown in green color, while EPO is in color defined for each atom.

administration of EPO in the first 24 hours of brain ischemia results in induction of angiogenesis and neurogenesis, reduced neural loss and prevention of BBB disruption (94, 95). Moreover, EPO could induce expression of EPO-R, downregulate HIF-1 α expression in ischemic region, decline the level of IL- β , increase sensorimotor and cognitive responses and decrease traumatic axonal injury specially in hypoxic context. (96, 97). Furthermore, upregulation of mitochondrial respiratory complex III and IV as well as regulation of neural energetics and stabilization of mitochondrial membrane potential by EPO prevents mitochondrial damage, inhibits oxygen free radicals, and cytochrome C release (98, 99). Activation of

PI3K/Akt pathway by EPO was found to be effective against neural cell apoptosis (100). Janus kinase-2 (JAK-2) signaling pathway downstream to EPO-R activation in a positive cross talk with NF- κ B promotes transcription of neuro-protective genes ⁽¹⁰¹⁾. In addition, EPO-mediated activation of the signal transducers and activators of transcription 5 (STAT5) protein, downstream to JAK-2 signaling, leads to upregulation of B-cell lymphoma extra-large (Bcl-xL) and X-linked inhibitor of apoptosis protein (XIAP) with anti-apoptotic properties ⁽¹⁰²⁾. Elevation of Bcl/Bax ratio and prevention of release of caspase-3 and -9 by EPO lead to microglia survival without affecting their pro-inflammatory characteristics (103-105). In a

cell culture study, EPO was shown to have protective effect on microglia and astrocytes against oxidative stress injury (106). On the other hand, Inhibition of (AQP-4)-induced astrocyte swelling and downregulation of MMP-9 via increasing the expression of TIMP-1 by EPO aids in reducing neuroinflammation (107, 108).

EPO-R and EPO found on the plasma membrane of human CD4⁺ and CD8⁺ T cells suppress alloreactive human T-cell immunity through inhibition of downstream T-cell and IL-2 receptor signaling pathways (109).

EPO and Neuroinflammation in COVID-19

Generally, EPO administration in sepsis and infections is controversial due to its suppressive effect against the function of macrophages, documented in Salmonella infection.(110, 111) However, EPO in an animal study could improve survival in sepsis due to restoration of the aorta responsiveness to norepinephrine (NE), upregulation of eNOS and decline in iNOS (112). According to the outcome of several clinical trials that have shown similar cell protective effect and an improvement in the status of the COVID-19 patients after administration of erythropoietin, a randomized clinical trial is currently underway (113, 114). Although EPO exerts neuroprotective effects, its administration in COVID19-induced neuroinflammation as a special, yet imprecisely identified subject, needs meticulous consideration.

1. Renin-Angiotensin System (RAS) and COVID-19

A novel hypothesis, supported by a large body of literature, could attribute the pathophysiology of cytokine storm in COVID19 to the virus-induced downregulation of angiotensin converting

enzyme 2 (ACE2). ACE2, while as a member of RAS family metabolizes Ang II to angiotensin [1-7], is the receptor of SARS-CoV2 on the host cell. Downregulation of ACE2 subsequent to SARS-CoV2 entry to cells leads to acute accumulation of Ang II. This sudden surplus of Ang II (whether locally produced or spilled over from the cells into the local or general circulation) results in supra-activation of type 1 angiotensin II receptor (AT1R) with all the pro-inflammatory, prothrombotic and pro-apoptotic effects eventuating into cytokine storm and tissue pathological changes (115, 116). A recent animal study showed that infusion of Ang II to swine resulted in pathological changes in the lungs similar to that of ARDS in COVID-19 (117). ACE2 deficiency was also shown to increase brain swelling and cell death in an animal model of brain ischemia (118).

It has been reported that Ang II blood level in patients with COVID-19, especially in moderate to severe form, is higher than that seen in milder cases or non-infected healthy people. The severity of the disease has also been correlated to the circulating level of Ang II (119, 120). Accordingly, administration of Ang II or applying any measure which may increase the level or effect of Ang II in patients with COVID-19 is not recommended (121).

2. RAS and Brain involvement in COVID-19

It merits mentioning that local RAS has been discovered in the brain and even in extensions of neural tissue like retina(122). Ang II contributes to some physiological functions in the brain such as the control of synaptic transmission and neuronal excitability, cognition and memory processing, regulation of autonomic responses and hormone secretion (123-125). Moreover, an angiotensinergic sympatho-

excitatory pathway in the brain connects the circumventricular area (lacking competent BBB) including subfornical organ (SFO) and the organum vasculosum of the lamina terminalis (OVLT) to rostral ventrolateral medulla (RVLM) via paraventricular nucleus (PVN) in the hypothalamus or directly to the intermediolateral cell column in the spinal cord. Nevertheless, Ang II content in the brain should be regulated delicately as its excess may have devastating effects like oxidative stress and endothelial dysfunction (126, 127). ACE2, beyond its wide expression in the neurons, astrocytes, cerebral vascular smooth muscle cells and endothelium, has been reported in mice to predominate in circumventricular area, hypothalamus and brain stem (128-130). As this receptor of SARS-CoV2 downregulates with entry of the virus to the host cells, Ang II content increases especially in these areas if the virus reaches the brain. In addition, excess of circulating Ang II in COVID-19 may reach the brain especially at sites where BBB is incompetent or disrupted which results in AT1R over-stimulation. This effect increases ROS through activation of NADPH oxidase which leads to an increase in intracellular calcium concentration (Ca^{2+}_i) followed by inhibition of voltage-gated potassium channels resulting in inhibition of GABAergic interneurons and depolarization of glutaminergic with an excitotoxic effect (131-133). AT1R is located on both glutaminergic neurons in RVLM and GABAergic interneurons in caudal ventrolateral medulla (CVLM) (134). Furthermore, massive release of glutamate in excitotoxicity damages the function of ACE2 via shedding of its ectodomain by a member of “A Disintegrin And Metalloprotease (ADAM)” family called ADAM17 (135). Additionally, this results in further cell injury

due to an increase in Ang II due to lack of available ACE2 and in exaggerated AT1R activation.

Activation of AT1R located in circumventricular area and cerebrovascular pericytes and endothelial cells by circulating or locally produced Ang II compromises neurovascular coupling and diminishes cerebral blood flow (low dose Ang II of 0.1 pmol/min leads to 23% reduction in CBF) (136, 137). Intriguingly, it has been reported that Ang II-mediated activation of AT1R of microglial cells in the retina may result in neurovascular uncoupling and inflammation in this special type of neural tissue (138, 139). This effect of Ang II on blood perfusion and inflammatory response in the neural tissue may explain the neuroimaging findings observed in most cases of COVID-19 (140).

EPO and Ang II

A cell culture study showed that recombinant human EPO exhibits synergistic effects with Ang II and NE on mobilization of intracellular Ca^{2+} in vascular smooth muscle cells lasting up to 60 minutes. This effect of EPO results in enhanced vasoconstriction and hypertension in the presence of Ang II (33, 141). Almost two decades ago, EPO was shown to affect Ca homeostasis in neural cells (142). Furthermore, EPO dose-dependently induces expression of TRPC channel protein up to 70% which facilitates Ca^{2+} influx. Moreover, Ang II exhibits similar TRPC-mediated Ca^{2+} current which soars up in synergism with EPO (143). Although cytosolic Ca^{2+} mediates several homeostatic pathways such as gene regulation, neural excitability, neurosecretion and synaptic plasticity, its intracellular concentration is finely regulated because any increase in its content, if sustained enough, may induce apoptosis-mediated loss of

neurons and other cells (144, 145). Besides, Ca^{2+} signaling has been linked to neuroinflammation as $\text{TNF-}\alpha$ was reported to enhance release of Ca^{2+} from its intracellular deposits. In addition, glutamate, ATP, cytokines and other inflammatory mediators increase the content of this ion in glial cells (146, 147). However, the interaction among EPO, Ang II and intracellular Ca^{2+} content needs further investigation due to its complexity.

Neuroinflammation generally is accompanied by excitotoxicity due to ineffective reuptake of glutamate in the synaptic cleft by excitatory amino acid transporter (EAAT) (148, 149). As mentioned earlier Ang II via AT1R contribute to this phenomenon by inhibiting glutamate transporter function in astrocytes (131, 150, 151). Intriguingly, a cell culture study elucidated that EPO in the absence of excitotoxic condition could increase calcium influx in cell culture of cortical neurons. However, EPO in excitotoxic conditions tends to repress elevation of $[\text{Ca}^{2+}_i]$, yet insignificantly. Besides, EPO could not repress the increase in $[\text{Ca}^{2+}_i]$ resulting from stimulation of metabotropic glutamate receptor (152). Bearing in mind that Ang II surplus is likely the cause of inflammatory responses in COVID-19 (115) and, on the other hand, Ang II dose dependently increases Ca^{2+} transient content in neural and glial cells (153, 154), it is legitimately expected that solitary EPO administration in COVID-19, in face of elevation of Ang II, may raise $[\text{Ca}^{2+}_i]$ in the absence of excitotoxic condition or at least may not be able to repress effectively the rise of $[\text{Ca}^{2+}_i]$ in excitotoxicity, both with probable hazardous outcome.

Ang II acts as a direct EPO secretagogue (155). Activation of both AT1R and AT2R

by Ang II upregulates EPO (155, 156). Furthermore, activation of AT1R induces hypoxia-inducible factor α -1 (HIF α 1) gene even in normoxia (157). This effect opposes the degradation of hydroxylated HIF through ubiquitination by von Hippel-Lindau protein (VHL) in normoxia (fig. 2) (158). Stimulation of AT2R has also been shown in a cell culture study to increase HIF1- α through a post-transcriptional regulatory pathway (159). In addition, another cell culture study demonstrated that intermittent hypoxia could activate HIF by NADPH oxidase which is by itself induced by Ang II in the CNS (160, 161). HIF1- α beyond its

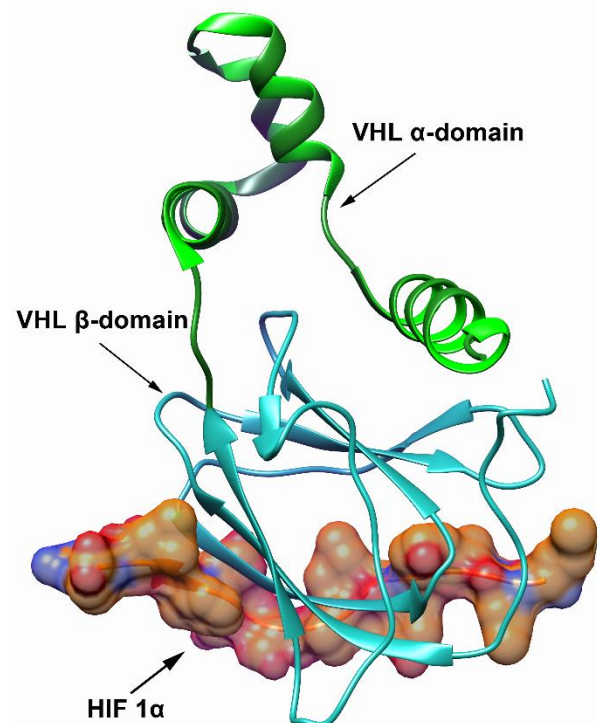


Figure 2. VHL with HIF-1 α (Pdb identifier: 1lm8). VHL- α domain is depicted in green, VHL- β in blue, HIF (surface view) in defined color of each atom.

pro-inflammatory effects upregulates EPO expression (162). Thus, it is hypothetically expected that in conditions associated with higher content of Ang II (circulating, local or intracellular), like that happens in COVID-

19, local or circulating EPO production may rise.

Consequently, as to the positive feedback interactions between EPO and Ang II, EPO should be administered in patients with COVID-19 cautiously (141, 163).

Two Novel Hypotheses

EPO production may increase in COVID-19 due to the probable rise of HIF in the presence of PLpro

As mentioned before, in the presence of sufficient pressure of oxygen in the tissues, hydroxylated HIF1- α is degraded after being ubiquitinated by von Hippel-Lindau protein (VHL) (fig. 2), which is a recognition substrate component of an E3 ubiquitin ligase (158, 164). Considering deubiquitinating property of papain-like protease (PLpro) of coronaviruses (165-168), it is wise to investigate whether PLpro in cytosol of infected cells with SARS-CoV2 prevents ubiquitination-mediated degradation of HIF1- α in normoxia which may lead to a rise in local EPO, as well. If this hypothesis can be validated through in vitro and in vivo studies, discovering potential inhibitors of PLpro, as being currently attempted vigorously, might provide the means to suppress the pro-inflammatory effect of HIF in COVID-19.

EPO alone might be hazardous in COVID-19-Induced Neuroinflammation; losartan may reverse the harmful pathway

In an animal study synergistic neuroprotective effect of EPO and olmesartan (an AT1R blocker) in stroke was reported (169). In addition, neuroprotective and anti-inflammatory effects of losartan in retinal ischemia-reperfusion injury and diabetic retinopathy have been explained in animal studies (170, 171). Accordingly, it

seems that due to the complex cellular interactions of Ang II and EPO, administration of a combination of EPO and an angiotensin receptor blocker (ARB) is more rational than solitary administration of EPO in attenuating COVID-19-induced neuroinflammation. Wisely, while EPO exerts anti-apoptotic and anti-inflammatory effects, ARBs block post-receptor untoward pathways of the surplus of Ang II in COVID-19. Considering the following data, it is legitimate to add losartan (instead of olmesartan) to EPO in treating neuroinflammation in COVID-19:

1. losartan apart from being an ARB was demonstrated in an in silico study to have the ability to change the conformational shape of PLpro of SARS-CoV2; other ARBs did not show such favorable results (fig. 3) ⁽¹¹⁵⁾. In this context, losartan can be considered a potential inhibitor of PLpro.

2. losartan according to a bio-informatic study could decline the affinity of SARS-CoV2 to ACE2; other ARBs did not show sufficient docking energy (115),

3. losartan has low water solubility and like olmesartan which is lipid soluble may penetrate into the brain tissue effectively (172, 173).

4. losartan compared to olmesartan has modest antihypertensive property so that the patients with COVID-19 may experience less probability of hypotension with losartan (174).

5. Considering retina as a neural tissue, losartan was demonstrated to attenuate retinal neurovascular decoupling in patients with diabetic retinopathies (170).

Subclinical and randomized clinical trials are needed to prove these hypotheses.

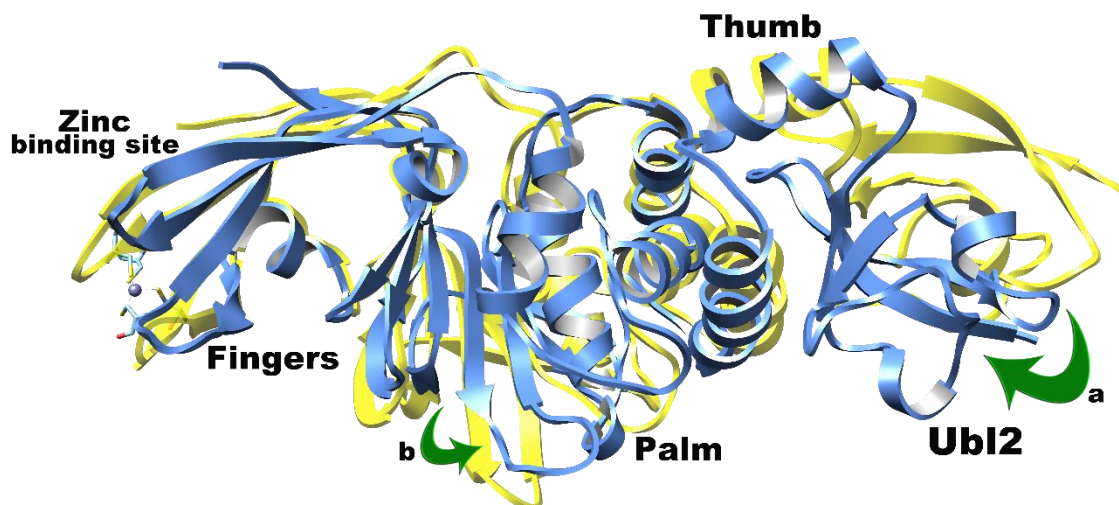


Figure 3. Superimposition of the structures of papain-like protease (PLpro) obtained in 100ns molecular dynamic (MD) simulation before (yellow) and after (blue) being affected by losartan. MD simulations of PLpro with and without losartan was performed by Gromacs 2018 package with gromos43a1 force field and the structural changes of PLpro were investigated: a. structural changes in UBL 2 b. structural changes in the cleavage site of the protein between finger and palm area.

Conclusion

According to a large number of studies, EPO's effect in subsiding neuroinflammation may reduce mortality and morbidity in many neurological disorders. Based on the hypothesis attributing COVID-19 pathophysiology to the excess of Ang II due to downregulation of ACE2, in which solitary administration of EPO might be harmful, combination of EPO and losartan might be promising in ameliorating neuroinflammatory reactions in COVID-19-induced brain disease.

Acknowledgement:

The authors would like to dedicate this article to their families. Moreover, they would like to express their greatest thanks and gratitude to physicians including intensivists, anesthesiologists, radiologists, emergency medicine, infectious disease specialists and other healthcare workers including committed nurses, radiology technicians, and all the personnel of hospitals who give so much by sacrificing their lives with dignity to aid the ill in this CORONA pandemic.

References:

1. DiSabato, D.J., N. Quan, and J.P. Godbout, Neuroinflammation: the devil is in the details. *Journal of neurochemistry*, 2016. **139**: p. 136-153.
2. Sochocka, M., B.S. Diniz, and J. Leszek, Inflammatory response in the CNS: friend or foe? *Molecular neurobiology*, 2017. **54**(10): p. 8071-8089.
3. Shastri, A., D.M. Bonifati, and U. Kishore, Innate immunity and neuroinflammation. *Mediators of inflammation*, 2013. **2013**.
4. Ratter, J.M., et al., Proinflammatory effects of hypoglycemia in humans with or without diabetes. *Diabetes*, 2017. **66**(4): p. 1052-1061.
5. Fang, M., et al., Effect of Inflammation on the Process of Stroke Rehabilitation and poststroke depression. *Frontiers in Psychiatry*, 2019. **10**.
6. Liu, R., et al., Role of neuroinflammation in ischemic stroke. *Neuroimmunol Neuroinflammation*, 2017. **4**: p. 158-66.
7. Tschöe, C., et al., Neuroinflammation after intracerebral hemorrhage and potential

- therapeutic targets. *Journal of stroke*, 2020. **22**(1): p. 29.
- 8.Mracsko, E. and R. Veltkamp, Neuroinflammation after intracerebral hemorrhage. *Frontiers in cellular neuroscience*, 2014. **8**: p. 388.
- 9.Wofford, K.L., D.J. Loane, and D.K. Cullen, Acute drivers of neuroinflammation in traumatic brain injury. *Neural regeneration research*, 2019. **14**(9): p. 1481.
- 10.Dinet, V., K.G. Petry, and J. Badaut, Brain-immune interactions and neuroinflammation after traumatic brain injury. *Frontiers in neuroscience*, 2019. **13**: p. 1178.
- 11.Hensley, K., Neuroinflammation in Alzheimer's disease: mechanisms, pathologic consequences, and potential for therapeutic manipulation. *Journal of Alzheimer's disease*, 2010. **21**(1): p. 1-14.
- 12.Calsolaro, V. and P. Edison, Neuroinflammation in Alzheimer's disease: current evidence and future directions. *Alzheimer's & Dementia*, 2016. **12**(6): p. 719-732.
- 13.Millington, C., et al., Chronic neuroinflammation in Alzheimer's disease: new perspectives on animal models and promising candidate drugs. *BioMed research international*, 2014. **2014**.
- 14.Wang, Q., Y. Liu, and J. Zhou, Neuroinflammation in Parkinson's disease and its potential as therapeutic target. *Translational Neurodegeneration*, 2015. **4**(1): p. 19.
- 15.Gelders, G., V. Baekelandt, and A. Van der Perren, Linking neuroinflammation and neurodegeneration in Parkinson's disease. *Journal of immunology research*, 2018. **2018**.
- 16.Liu, J. and F. Wang, Role of neuroinflammation in amyotrophic lateral sclerosis: cellular mechanisms and therapeutic implications. *Frontiers in Immunology*, 2017. **8**: p. 1005.
- 17.Koudriavtseva, T. and C. Mainero, Neuroinflammation, neurodegeneration and regeneration in multiple sclerosis: intercorrelated manifestations of the immune response. *Neural regeneration research*, 2016. **11**(11): p. 1727.
- 18.Al-Badri, G. and A. Castorina, Insights into the Role of Neuroinflammation in the Pathogenesis of Multiple Sclerosis. *Journal of Functional Morphology and Kinesiology*, 2018. **3**(1): p. 13.
- 19.Vezzani, A., *Epilepsy and Inflammation in the Brain: Overview and Pathophysiology: Epilepsy and Inflammation in the Brain*. *Epilepsy currents*, 2014. **14**(2_suppl): p. 3-7.
- 20.Jeon, S.W. and Y.K. Kim, Neuroinflammation and cytokine abnormality in major depression: cause or consequence in that illness? *World journal of psychiatry*, 2016. **6**(3): p. 283.
- 21.Troubat, R., et al., Neuroinflammation and depression: A review. *European Journal of Neuroscience*, 2020.
- 22.Gerentes, M., et al., Obsessive-Compulsive Disorder: Autoimmunity and Neuroinflammation. *Current psychiatry reports*, 2019. **21**(8): p. 78.
- 23.Aricioglu, F., et al., Neuroinflammation in schizophrenia: a critical review and the future. *Klinik Psikofarmakoloji Bülteni-Bulletin of Clinical Psychopharmacology*, 2016. **26**(4): p. 429-437.
- 24.Shin, T., et al., Erythropoietin and autoimmune neuroinflammation: lessons from experimental autoimmune encephalomyelitis and experimental autoimmune neuritis. *Anatomy & cell biology*, 2012. **45**(4): p. 215-220.
- 25.Moransard, M., et al., Erythropoietin reduces experimental autoimmune encephalomyelitis severity via

- neuroprotective mechanisms. *Journal of neuroinflammation*, 2017. **14**(1): p. 202.
- 26.Chong, Z.Z., J.-Q. Kang, and K. Maiese, Apaf-1, Bcl-xL, cytochrome c, and caspase-9 form the critical elements for cerebral vascular protection by erythropoietin. *Journal of Cerebral Blood Flow & Metabolism*, 2003. **23**(3): p. 320-330.
- 27.Liu, W.-C., et al., Therapeutic effect of erythropoietin in patients with traumatic brain injury: a meta-analysis of randomized controlled trials. *Journal of neurosurgery*, 2016. **127**(1): p. 8-15.
- 28.Nirula, R., et al., Safety and efficacy of erythropoietin in traumatic brain injury patients: a pilot randomized trial. *Critical care research and practice*, 2010. **2010**.
- 29.Ehrenreich, H., et al., Erythropoietin therapy for acute stroke is both safe and beneficial. *Molecular medicine*, 2002. **8**(8): p. 495-505.
- 30.Tang, Y.-D., H.M. Rinder, and S.D. Katz, Effects of recombinant human erythropoietin on antiplatelet action of aspirin and clopidogrel in healthy subjects: results of a double-blind, placebo-controlled randomized trial. *American heart journal*, 2007. **154**(3): p. 494. e1-494. e7.
- 31.Demetz, G., et al., The influence of Erythropoietin on platelet activation, thrombin generation and FVII/active FVII in patients with AMI. *Thrombosis journal*, 2014. **12**(1): p. 18.
- 32.Tobu, M., et al., Erythropoietin-induced thrombosis as a result of increased inflammation and thrombin activatable fibrinolytic inhibitor. *Clinical and applied thrombosis/hemostasis*, 2004. **10**(3): p. 225-232.
- 33.Agarwal, R., Mechanisms and mediators of hypertension induced by erythropoietin and related molecules. *Nephrology Dialysis Transplantation*, 2018. **33**(10): p. 1690-1698.
- 34.Nejat, R., et al., Daily Low Dose of Erythropoietin in Neuroinflammation; EPO Might Be Hazardous in COVID-19. 2020: preprints.org. DOI: 10.20944/preprints202006.0107.v1
- 35.Yatsiv, I., et al., Erythropoietin is neuroprotective, improves functional recovery, and reduces neuronal apoptosis and inflammation in a rodent model of experimental closed head injury. *The FASEB journal*, 2005. **19**(12): p. 1701-1703.
- 36.Banjara, M. and C. Ghosh, Sterile neuroinflammation and strategies for therapeutic intervention. *International journal of inflammation*, 2017. **2017**.
- 37.Noctor, S.C., et al., Neurons derived from radial glial cells establish radial units in neocortex. *Nature*, 2001. **409**(6821): p. 714-720.
- 38.Barry, D.S., J.M. Pakan, and K.W. McDermott, Radial glial cells: key organisers in CNS development. *The international journal of biochemistry & cell biology*, 2014. **46**: p. 76-79.
- 39.Lima, C.M.d., et al., Differential change in hippocampal radial astrocytes and neurogenesis in shorebirds with contrasting migratory routes. *Frontiers in neuroanatomy*, 2019. **13**: p. 82.
- 40.Cho, H., et al., Differential innate immune response programs in neuronal subtypes determine susceptibility to infection in the brain by positive-stranded RNA viruses. *Nature medicine*, 2013. **19**(4): p. 458.
- 41.Ising, C. and M.T. Heneka, Functional and structural damage of neurons by innate immune mechanisms during neurodegeneration. *Cell death & disease*, 2018. **9**(2): p. 1-8.
- 42.Ma, Y., et al., TLR8: an innate immune receptor in brain, neurons and axons. *Cell Cycle*, 2007. **6**(23): p. 2859-2868.

43. González-Reyes, R.E. and M.G. Rubiano, Astrocyte' s RAGE: More than just a question of mood. *Central Nervous System Agents in Medicinal Chemistry (Formerly Current Medicinal Chemistry-Central Nervous System Agents)*, 2018. **18**(1): p. 39-48.
44. Kigerl, K.A., et al., Pattern recognition receptors and central nervous system repair. *Experimental neurology*, 2014. **258**: p. 5-16.
45. Kong, Y. and Y. Le, Toll-like receptors in inflammation of the central nervous system. *International immunopharmacology*, 2011. **11**(10): p. 1407-1414.
46. Peltier, D.C., et al., Human neuronal cells possess functional cytoplasmic and TLR-mediated innate immune pathways influenced by phosphatidylinositol-3 kinase signaling. *The Journal of Immunology*, 2010. **184**(12): p. 7010-7021.
47. Canton, J., D. Neculai, and S. Grinstein, Scavenger receptors in homeostasis and immunity. *Nature Reviews Immunology*, 2013. **13**(9): p. 621-634.
48. Vorup-Jensen, T. and R.K. Jensen, Structural immunology of complement receptors 3 and 4. *Frontiers in immunology*, 2018. **9**: p. 2716.
49. Jahagirdar, P., et al., Mannose Receptor and Targeting Strategies, in *Targeted intracellular drug delivery by receptor mediated endocytosis*. 2019, Springer. p. 433-456.
50. Ransohoff, R.M. and M.A. Brown, Innate immunity in the central nervous system. *The Journal of clinical investigation*, 2012. **122**(4): p. 1164-1171.
51. Liu, T., et al., NF- κ B signaling in inflammation. *Signal transduction and targeted therapy*, 2017. **2**(1): p. 1-9.
52. Rui, W., et al., PM2. 5-induced oxidative stress increases adhesion molecules expression in human endothelial cells through the ERK/AKT/NF- κ B-dependent pathway. *Journal of Applied Toxicology*, 2016. **36**(1): p. 48-59.
53. Sun, B., et al., Activation of NF κ B and Expression of ICAM-1 in Ischemic-reperfused Canine Myocardium. *Journal of molecular and cellular cardiology*, 2001. **33**(1): p. 109-119.
54. Milstone, D.S., et al., Differential role of an NF- κ B transcriptional response element in endothelial versus intimal cell VCAM-1 expression. *Circulation research*, 2015. **117**(2): p. 166-177.
55. Singh, S.S., et al., NF- κ B-Mediated Neuroinflammation in Parkinson's Disease and Potential Therapeutic Effect of Polyphenols. *Neurotoxicity Research*, 2019: p. 1-17.
56. Ju Hwang, C., et al., NF-KB as a Key Mediator of Brain Inflammation in Alzheimer's Disease. *CNS & Neurological Disorders-Drug Targets (Formerly Current Drug Targets-CNS & Neurological Disorders)*, 2019. **18**(1): p. 3-10.
57. Flood, P.M., et al., Transcriptional factor NF- κ B as a target for therapy in Parkinson's disease. *Parkinson's disease*, 2011. **2011**.
58. Jones, S.V. and I. Kounatidis, Nuclear factor-kappa B and Alzheimer disease, unifying genetic and environmental risk factors from cell to humans. *Frontiers in immunology*, 2017. **8**: p. 1805.
59. Li, Y., O. Sibon, and P. Dijkers, Inhibition of NF- κ B in astrocytes is sufficient to delay neurodegeneration induced by proteotoxicity in neurons. *Journal of neuroinflammation*, 2018. **15**(1): p. 261.
60. Ridder, D. and M. Schwaninger, NF- κ B signaling in cerebral ischemia. *Neuroscience*, 2009. **158**(3): p. 995-1006.
61. Theus, M.H., et al., Loss of NLRX1 exacerbates neural tissue damage and NF- κ B signaling following brain injury. *The Journal*

- of Immunology, 2017. **199**(10): p. 3547-3558.
62. Bertheloot, D. and E. Latz, HMGB1, IL-1 α , IL-33 and S100 proteins: dual-function alarmins. *Cellular & molecular immunology*, 2017. **14**(1): p. 43-64.
63. Xu, J., et al., Microglial activation induced by the alarmin S100B is regulated by poly (ADP-ribose) polymerase-1. *Glia*, 2016. **64**(11): p. 1869-1878.
64. Shih, R.-H., C.-Y. Wang, and C.-M. Yang, NF-kappaB signaling pathways in neurological inflammation: a mini review. *Frontiers in molecular neuroscience*, 2015. **8**: p. 77.
65. Hollborn, M., et al., Positive feedback regulation between MMP-9 and VEGF in human RPE cells. *Investigative ophthalmology & visual science*, 2007. **48**(9): p. 4360-4367.
66. McGeehan, G.M., et al., Regulation of tumour necrosis factor- α processing by a metalloproteinase inhibitor. *Nature*, 1994. **370**(6490): p. 558-561.
67. Cox, J.H. and C.M. Overall, Cytokine substrates: MMP regulation of inflammatory signaling molecules, in *The Cancer Degradome*. 2008, Springer. p. 519-539.
68. Arpino, V., M. Brock, and S.E. Gill, The role of TIMPs in regulation of extracellular matrix proteolysis. *Matrix Biology*, 2015. **44**: p. 247-254.
69. Meli, R., C. Pirozzi, and A. Pelagalli, New perspectives on the potential role of aquaporins (AQPs) in the physiology of inflammation. *Frontiers in physiology*, 2018. **9**: p. 101.
70. Tourdias, T., et al., Differential aquaporin 4 expression during edema build-up and resolution phases of brain inflammation. *Journal of Neuroinflammation*, 2011. **8**(1): p. 143.
71. Van Hoecke, M., et al., Evidence of HIF-1 functional binding activity to caspase-3 promoter after photothrombotic cerebral ischemia. *Molecular and Cellular Neuroscience*, 2007. **34**(1): p. 40-47.
72. Chandel, N.S., et al., Reactive oxygen species generated at mitochondrial complex III stabilize hypoxia-inducible factor-1 α during hypoxia a mechanism of O₂ sensing. *Journal of Biological Chemistry*, 2000. **275**(33): p. 25130-25138.
73. Klimova, T. and N. Chandel, Mitochondrial complex III regulates hypoxic activation of HIF. *Cell Death & Differentiation*, 2008. **15**(4): p. 660-666.
74. Haibara, A.S., N.M. Sharma, and K.P. Patel, Hypoxia-Inducible Factor (HIF)-1 Alpha Expression Induced by Proinflammatory Cytokines in NG108-15 Neuronal Cells. *The FASEB Journal*, 2016. **30**(1_supplement): p. 1234.3-1234.3.
75. Otto, T. and J. Fandrey, Thyroid hormone induces hypoxia-inducible factor 1 α gene expression through thyroid hormone receptor β /retinoid x receptor α -dependent activation of hepatic leukemia factor. *Endocrinology*, 2008. **149**(5): p. 2241-2250.
76. Qutub, A.A. and A.S. Popel, Reactive oxygen species regulate hypoxia-inducible factor 1 α differentially in cancer and ischemia. *Molecular and cellular biology*, 2008. **28**(16): p. 5106-5119.
77. Gorlach, A., et al., Thrombin activates the hypoxia-inducible factor-1 signaling pathway in vascular smooth muscle cells: role of the p22 phox-containing NADPH oxidase. *Circulation research*, 2001. **89**(1): p. 47-54.
78. Feldser, D., et al., Reciprocal positive regulation of hypoxia-inducible factor 1 α and insulin-like growth factor 2. *Cancer research*, 1999. **59**(16): p. 3915-3918.

79. Jiang, F., et al., The role of insulin-like growth factor I and hypoxia inducible factor 1 α in vascular endothelial growth factor expression in type 2 diabetes. *Annals of Clinical & Laboratory Science*, 2013. **43**(1): p. 37-44.
80. Yoshida, T., et al., Transcriptional upregulation of HIF-1 α by NF- κ B/p65 and its associations with β -catenin/p300 complexes in endometrial carcinoma cells. *Laboratory investigation*, 2013. **93**(11): p. 1184-1193.
81. Bartczek, P., et al., Neuronal HIF-1 α and HIF-2 α deficiency improves neuronal survival and sensorimotor function in the early acute phase after ischemic stroke. *Journal of Cerebral Blood Flow & Metabolism*, 2017. **37**(1): p. 291-306.
82. Chen, W., et al., Prodeath or prosurvival: two facets of hypoxia inducible factor-1 in perinatal brain injury. *Experimental neurology*, 2009. **216**(1): p. 7-15.
83. Gawad, A., et al., Antiapoptotic properties of erythropoietin: novel strategies for protection of retinal pigment epithelial cells. *Eye*, 2009. **23**(12): p. 2245-2250.
84. Alural, B., et al., EPO mediates neurotrophic, neuroprotective, anti-oxidant, and anti-apoptotic effects via downregulation of miR-451 and miR-885-5p in SH-SY5Y neuron-like cells. *Frontiers in immunology*, 2014. **5**: p. 475.
85. MacRedmond, R., G.K. Singhera, and D.R. Dorscheid, Erythropoietin inhibits respiratory epithelial cell apoptosis in a model of acute lung injury. *European Respiratory Journal*, 2009. **33**(6): p. 1403-1414.
86. Olsen, N.V., Central nervous system frontiers for the use of erythropoietin. *Clinical infectious diseases*, 2003. **37**(Supplement_4): p. S323-S330.
87. Nejat, R., Neuroprotective Effect of Erythropoietin. *International Journal of Emergency Mental Health & Human Resilience*, 2018. **20**: p. 21.
88. Banks, W.A., et al., Passage of erythropoietic agents across the blood-brain barrier: a comparison of human and murine erythropoietin and the analog darbepoetin alfa. *European journal of pharmacology*, 2004. **505**(1-3): p. 93-101.
89. Brines, M.L., et al., Erythropoietin crosses the blood-brain barrier to protect against experimental brain injury. *Proceedings of the National Academy of Sciences*, 2000. **97**(19): p. 10526-10531.
90. Nagai, A., et al., Erythropoietin and erythropoietin receptors in human CNS neurons, astrocytes, microglia, and oligodendrocytes grown in culture. *Journal of Neuropathology & Experimental Neurology*, 2001. **60**(4): p. 386-392.
91. Noguchi, C.T., et al., Role of erythropoietin in the brain. *Critical reviews in oncology/hematology*, 2007. **64**(2): p. 159-171.
92. Alnaeli, M., et al., Erythropoietin in brain development and beyond. *Anatomy research international*, 2012. **2012**.
93. Hassouna, I., et al., Revisiting adult neurogenesis and the role of erythropoietin for neuronal and oligodendroglial differentiation in the hippocampus. *Molecular psychiatry*, 2016. **21**(12): p. 1752-1767.
94. Wang, L., et al., Treatment of stroke with erythropoietin enhances neurogenesis and angiogenesis and improves neurological function in rats. *Stroke*, 2004. **35**(7): p. 1732-1737.
95. Thériault, P., et al., Sub-acute systemic erythropoietin administration reduces ischemic brain injury in an age-dependent manner. *Oncotarget*, 2016. **7**(24): p. 35552.

- 96.Souvenir, R., et al., Erythropoietin inhibits HIF-1 α expression via upregulation of PHD-2 transcription and translation in an in vitro model of hypoxia-ischemia. *Translational stroke research*, 2014. **5**(1): p. 118-127.
- 97.Hellewell, S.C., et al., Erythropoietin improves motor and cognitive deficit, axonal pathology, and neuroinflammation in a combined model of diffuse traumatic brain injury and hypoxia, in association with upregulation of the erythropoietin receptor. *Journal of neuroinflammation*, 2013. **10**(1): p. 1-21.
- 98.Wang, Y., et al., Erythropoietin (EPO) protects against high glucose-induced apoptosis in retinal ganglionic cells. *Cell biochemistry and biophysics*, 2015. **71**(2): p. 749-755.
- 99.Singhal, N., et al., Erythropoietin upregulates brain hemoglobin expression and supports neuronal mitochondrial activity. *Molecular neurobiology*, 2018. **55**(10): p. 8051-8058.
- 100.Weishaupt, J.H., et al., Effect of erythropoietin axotomy-induced apoptosis in rat retinal ganglion cells. *Investigative ophthalmology & visual science*, 2004. **45**(5): p. 1514-1522.
- 101.Digicaylioglu, M. and S.A. Lipton, Erythropoietin-mediated neuroprotection involves cross-talk between Jak2 and NF- κ B signalling cascades. *Nature*, 2001. **412**(6847): p. 641-647.
- 102.Zhang, F., et al., Signal transducers and activators of transcription 5 contributes to erythropoietin-mediated neuroprotection against hippocampal neuronal death after transient global cerebral ischemia. *Neurobiology of disease*, 2007. **25**(1): p. 45-53.
- 103.Liao, Z., et al., Erythropoietin can promote survival of cerebral cells by downregulating Bax gene after traumatic brain injury in rats. *Neurology India*, 2009. **57**(6): p. 722.
- 104.Sekiguchi, N., et al., Effect of erythropoietin on endothelial cell apoptosis induced by high glucose. *Diabetes research and clinical practice*, 2004. **66**: p. S103-S107.
- 105.Wenker, S.D., et al., Protective action of erythropoietin on neuronal damage induced by activated microglia. *The FEBS journal*, 2013. **280**(7): p. 1630-1642.
- 106.Pathipati, P. and D.M. Ferriero, The differential effects of erythropoietin exposure to oxidative stress on microglia and astrocytes in vitro. *Developmental neuroscience*, 2017. **39**(1-4): p. 310-322.
- 107.Tang, G. and G.-Y. Yang, Aquaporin-4: A potential therapeutic target for cerebral edema. *International journal of molecular sciences*, 2016. **17**(10): p. 1413.
- 108.Souvenir, R., et al., Janus kinase 2 and tissue inhibitor of matrix metalloproteinase-1 mediate the protective effects of erythropoietin in in-vitro model of hypoxia ischemia. 2009, Federation of American Societies for Experimental Biology.
- 109.Cravedi, P., et al., Immunosuppressive effects of erythropoietin on human alloreactive T cells. *Journal of the American Society of Nephrology*, 2014. **25**(9): p. 2003-2015.
- 110.Nairz, M., et al., The pleiotropic effects of erythropoietin in infection and inflammation. *Microbes and infection*, 2012. **14**(3): p. 238-246.
- 111.Chousterman, B.G. and M. Arnaud, Is there a role for hematopoietic growth factors during sepsis? *Frontiers in Immunology*, 2018. **9**: p. 1015.
- 112.Kandasamy, K., et al., Erythropoietin reverses sepsis-induced vasoplegia to norepinephrine through preservation of α 1D-adrenoceptor mRNA expression and inhibition of GRK2-mediated desensitization

- in mouse aorta. *Journal of cardiovascular pharmacology and therapeutics*, 2016. **21**(1): p. 100-113.
- 113.Hadadi, A., et al., Does recombinant human erythropoietin administration in critically ill COVID-19 patients have miraculous therapeutic effects? *Journal of Medical Virology*, 2020.
- 114.Ehrenreich, H., et al., Erythropoietin as candidate for supportive treatment of severe COVID-19. *Molecular Medicine*, 2020. **26**(1): p. 1-9.
- 115.Nejat, R. and A.S. Sadr, Are losartan and Imatinib Effective Against SARS-CoV2 Pathogenesis? A Pathophysiologic-Based in Silico Study. *chemRxiv*. preprint, 2020. DOI: 10.26434/chemrxiv.12271865.v1
- 116.Miesbach, W., Pathological Role of Angiotensin II in Severe COVID-19. *TH Open: Companion Journal to Thrombosis and Haemostasis*, 2020. **4**(2): p. e138.
- 117.Rysz, S., et al., COVID-19 pathophysiology may be driven by a loss of inhibition of the Renin-Angiotensin-Aldosterone System. PREPRINT (Version 1) available at Research Square, 2020.
- 118.Alenina, N. and M. Bader, ACE2 in brain physiology and pathophysiology: Evidence from transgenic animal models. *Neurochemical research*, 2019. **44**(6): p. 1323-1329.
- 119.Liu, Y., et al., Clinical and biochemical indexes from 2019-nCoV infected patients linked to viral loads and lung injury. *Science China Life Sciences*, 2020. **63**(3): p. 364-374.
- 120.Wu, Z., et al., Elevation of plasma angiotensin II level is a potential pathogenesis for the critically ill COVID-19 patients. *Critical Care*, 2020. **24**(1): p. 1-3.
- 121.Speth, R.C., Angiotensin II administration to COVID-19 patients is not advisable. *Critical Care*, 2020. **24**(1): p. 1-2.
- 122.Wilkinson-Berka, J.L., et al., Angiotensin II and aldosterone in retinal vasculopathy and inflammation. *Experimental eye research*, 2019. **187**: p. 107766.
- 123.Pan, H.-L., Brain angiotensin II and synaptic transmission. *The Neuroscientist*, 2004. **10**(5): p. 422-431.
- 124.Jackson, L., et al., Within the brain: the renin angiotensin system. *International journal of molecular sciences*, 2018. **19**(3): p. 876.
- 125.Senanayake, P.d., et al., Angiotensin II and its receptor subtypes in the human retina. *Investigative ophthalmology & visual science*, 2007. **48**(7): p. 3301-3311.
- 126.Landmesser, U., et al., Angiotensin II induces endothelial xanthine oxidase activation: role for endothelial dysfunction in patients with coronary disease. *Arteriosclerosis, Thrombosis, and Vascular Biology*, 2007. **27**(4): p. 943-948.
- 127.Chriissobolis, S., et al., Role of Nox isoforms in angiotensin II-induced oxidative stress and endothelial dysfunction in brain. *Journal of applied physiology*, 2012. **113**(2): p. 184-191.
- 128.Elased, K.M., et al., Brain angiotensin-converting enzymes: role of angiotensin-converting enzyme 2 in processing angiotensin II in mice. *Experimental physiology*, 2008. **93**(5): p. 665-675.
- 129.Xia, H. and E. Lazartigues, Angiotensin-converting enzyme 2 in the brain: properties and future directions. *Journal of neurochemistry*, 2008. **107**(6): p. 1482-1494.
- 130.Doobay, M.F., et al., Differential expression of neuronal ACE2 in transgenic mice with overexpression of the brain renin-angiotensin system. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology*, 2007. **292**(1): p. R373-R381.

131. Leenen, F.H., Actions of circulating angiotensin II and aldosterone in the brain contributing to hypertension. *American journal of hypertension*, 2014. **27**(8): p. 1024-1032.
132. Görlach, A., et al., Calcium and ROS: a mutual interplay. *Redox biology*, 2015. **6**: p. 260-271.
133. Shah, V.N., B. Chagot, and W.J. Chazin, Calcium-dependent regulation of ion channels. *Calcium binding proteins*, 2006. **1**(4): p. 203.
134. Bourassa, E.A. and R.C. Speth, Longitudinal analysis of AT-1 angiotensin receptor binding in the rostral and caudal ventrolateral medulla and nucleus of the solitary tract in the rat brainstem. 2008, Federation of American Societies for Experimental Biology. p. 1169.13.
135. Xu, J., S. Sriramula, and E. Lazartigues, Excessive glutamate stimulation impairs ACE2 activity through ADAM17-mediated shedding in cultured cortical neurons. *Cellular and molecular neurobiology*, 2018. **38**(6): p. 1235-1243.
136. Saavedra, J.M., Brain angiotensin II: new developments, unanswered questions and therapeutic opportunities. *Cellular and molecular neurobiology*, 2005. **25**(3-4): p. 485-512.
137. Kramár, E.A., J.W. Harding, and J.W. Wright, Angiotensin II-and IV-induced changes in cerebral blood flow: Roles of AT1 AT2, and AT4 receptor subtypes. *Regulatory peptides*, 1997. **68**(2): p. 131-138.
138. Phipps, J.A., et al., The renin-angiotensin system and the retinal neurovascular unit: A role in vascular regulation and disease. *Experimental Eye Research*, 2019. **187**: p. 107753.
139. Kurihara, T., et al., Renin-Angiotensin system hyperactivation can induce inflammation and retinal neural dysfunction. *International journal of inflammation*, 2012. **2012**.
140. Jain, R., et al., COVID-19 related neuroimaging findings: A signal of thromboembolic complications and a strong prognostic marker of poor patient outcome. *Journal of the Neurological Sciences*, 2020: p. 116923.
141. Akimoto, T., et al., Erythropoietin modulates angiotensin II-or noradrenaline-induced Ca²⁺ mobilization in cultured rat vascular smooth-muscle cells. *Nephrology Dialysis Transplantation*, 2001. **16**(3): p. 491-499.
142. Assandri, R., et al., Erythropoietin modulates intracellular calcium in a human neuroblastoma cell line. *The Journal of physiology*, 1999. **516**(Pt 2): p. 343.
143. Liu, Y., et al., Erythropoietin increases expression and function of transient receptor potential canonical 5 channels. *Hypertension*, 2011. **58**(2): p. 317-324.
144. Harr, M.W. and C.W. Distelhorst, Apoptosis and autophagy: decoding calcium signals that mediate life or death. *Cold Spring Harbor perspectives in biology*, 2010. **2**(10): p. a005579.
145. Bollimuntha, S., S. Selvaraj, and B.B. Singh, Emerging roles of canonical TRP channels in neuronal function, in *Transient Receptor Potential Channels*. 2011, Springer. p. 573-593.
146. Sama, D.M. and C.M. Norris, Calcium dysregulation and neuroinflammation: discrete and integrated mechanisms for age-related synaptic dysfunction. *Ageing research reviews*, 2013. **12**(4): p. 982-995.
147. Mizoguchi, Y. and A. Monji, Microglial intracellular Ca²⁺ signaling in synaptic development and its alterations in neurodevelopmental disorders. *Frontiers in Cellular Neuroscience*, 2017. **11**: p. 69.

148. Tilleux, S. and E. Hermans, Neuroinflammation and regulation of glial glutamate uptake in neurological disorders. *Journal of neuroscience research*, 2007. **85**(10): p. 2059-2070.
149. Magi, S., et al., Excitatory amino acid transporters (EAATs): Glutamate transport and beyond. *International journal of molecular sciences*, 2019. **20**(22): p. 5674.
150. Justin, A., S. Divakar, and M. Ramanathan, Cerebral ischemia induced inflammatory response and altered glutaminergic function mediated through brain AT1 and not AT2 receptor. *Biomedicine & Pharmacotherapy*, 2018. **102**: p. 947-958.
151. Stern, J.E., et al., Astrocytes contribute to angiotensin II stimulation of hypothalamic neuronal activity and sympathetic outflow. *Hypertension*, 2016. **68**(6): p. 1483-1493.
152. Andoh, T., et al., Effects of erythropoietin on intracellular calcium concentration of rat primary cortical neurons. *Brain research*, 2011. **1387**: p. 8-18.
153. Gebke, E., et al., Angiotensin II-induced calcium signalling in neurons and astrocytes of rat circumventricular organs. *Neuroscience*, 1998. **85**(2): p. 509-520.
154. Zimmerman, M.C., R.V. Sharma, and R.L. Davisson, Superoxide mediates angiotensin II-induced influx of extracellular calcium in neural cells. *Hypertension*, 2005. **45**(4): p. 717-723.
155. Calò, L.A., et al., Assessing the relationship of angiotensin II Type 1 receptors with erythropoietin in a human model of endogenous angiotensin II Type 1 receptor antagonism. *Cardiorenal medicine*, 2016. **6**(1): p. 16-24.
156. Gossmann, J., et al., Angiotensin II infusion increases plasma erythropoietin levels via an angiotensin II type 1 receptor-dependent pathway. *Kidney international*, 2001. **60**(1): p. 83-86.
157. Diebold, I., et al., The NADPH oxidase subunit NOX4 is a new target gene of the hypoxia-inducible factor-1. *Molecular biology of the cell*, 2010. **21**(12): p. 2087-2096.
158. Li, Z., et al., Identification of a deubiquitinating enzyme subfamily as substrates of the von Hippel-Lindau tumor suppressor. *Biochemical and biophysical research communications*, 2002. **294**(3): p. 700-709.
159. Wolf, G., R. Schroeder, and R.A. Stahl, Angiotensin II induces hypoxia-inducible factor-1 α in PC 12 cells through a posttranscriptional mechanism: role of AT2 receptors. *American journal of nephrology*, 2004. **24**(4): p. 415-421.
160. Nanduri, J., et al., HIF-1 α activation by intermittent hypoxia requires NADPH oxidase stimulation by xanthine oxidase. *PloS one*, 2015. **10**(3): p. e0119762.
161. Garrido, A.M. and K.K. Griendling, NADPH oxidases and angiotensin II receptor signaling. *Molecular and cellular endocrinology*, 2009. **302**(2): p. 148-158.
162. Imtiyaz, H.Z. and M.C. Simon, Hypoxia-inducible factors as essential regulators of inflammation, in *Diverse Effects of Hypoxia on Tumor Progression*. 2010, Springer. p. 105-120.
163. Kim, Y.-C., et al., Mechanism of erythropoietin regulation by angiotensin II. *Molecular pharmacology*, 2014. **85**(6): p. 898-908.
164. Haase, V.H., Regulation of erythropoiesis by hypoxia-inducible factors. *Blood reviews*, 2013. **27**(1): p. 41-53.
165. Nejat, R. and A.S. Sadr, SARS Virus Papain-like Protease: A Mysterious Weapon. *Journal of Biostatistics and Epidemiology*, 2019. **5**(4): p. 288-295.

- 166.Freitas, B.T., et al., Characterization and noncovalent inhibition of the deubiquitinase and deISGylase activity of SARS-CoV-2 papain-like protease. *ACS Infectious Diseases*, 2020. **6**(8): p. 2099-2109.
- 167.Clemente, V., P. D'Arcy, and M. Bazzaro, Deubiquitinating Enzymes in Coronaviruses and Possible Therapeutic Opportunities for COVID-19. *International Journal of Molecular Sciences*, 2020. **21**(10): p. 3492.
- 168.Lindner, H.A., Deubiquitination in virus infection. *Virology*, 2007. **362**(2): p. 245-256.
- 169.Faure, S., et al., Synergistic protective effects of erythropoietin and olmesartan on ischemic stroke survival and post-stroke memory dysfunctions in the gerbil. *Journal of hypertension*, 2006. **24**(11): p. 2255-2261.
- 170.Mori, F., et al., Inhibitory effect of losartan, an AT1 angiotensin II receptor antagonist, on increased leucocyte entrapment in retinal microcirculation of diabetic rats. *British journal of ophthalmology*, 2002. **86**(10): p. 1172-1174.
- 171.Fukuda, K., et al., Neuroprotection against retinal ischemia–reperfusion injury by blocking the angiotensin II type 1 receptor. *Investigative ophthalmology & visual science*, 2010. **51**(7): p. 3629-3638.
- 172.Souza, J.B.d., et al., Evaluation of the losartan solubility in the biowaiver context by shake-flask method and intrinsic dissolution. *Pharmaceutical Development and Technology*, 2019. **24**(3): p. 283-292.
- 173.Shakeel, F., et al., Measurement and correlation of solubility of olmesartan medoxomil in six green solvents at 295.15–330.15 K. *Industrial & Engineering Chemistry Research*, 2014. **53**(7): p. 2846-2849.
- 174.Oparil, S., et al., Comparative efficacy of olmesartan, losartan, valsartan, and irbesartan in the control of essential hypertension. *The Journal of Clinical Hypertension*, 2001. **3**(5): p. 283-318.