





Synergistic Effects of Lauryl Gallate and Tamoxifen on Human Breast Cancer Cell

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Abstract

Background: Tamoxifen (TAM) is widely used for adjuvant therapy in breast cancer patients. Tamoxifen therapy may lead to serious side effects. Anti-apoptotic substances in combination with chemotherapy drugs can result in additive or synergistic effects. Lauryl gallate (LG), a Gallic acid derivative, has been proven to inhibit tumor growth, without affecting normal cells. This study aimed to investigate the synergistic effect of TAM and LG in breast cancer cell line (MCF-7).

Methods: In this experimental study, performed in ShahreKord University of Medical Science, Iran in 2017, the MCF-7 cells were treated by final concentrations of 10 μM TAM alone, and in combination with 200 μM of LG. We also used EX-527, as SIRT-1 inhibitor to examine the role of SIRT1 in cell apoptosis. *BCL-2* and *SIRT1* gene expression were measured by real-time PCR method, and cell apoptosis was investigated by flow cytometry.

Results: Tamoxifen alone and in combination with LG decreased BCL-2 expression 2.64 ± 0.75 and 6.38 ± 1.9 fold, respectively, after 48 h (P<0.05). SIRT1 expression was increased 1.67 ± 0.22 and 2.47 ± 0.34 - fold by TAM alone and in combination with LG, respectively (P<0.05). TAM alone and in combination with LG increased the percentage of apoptotic cells 15.79 ± 2.81 and 60.67 ± 6.23 percent, respectively after 48 h (P<0.001).

Conclusion: The combination of LG and TAM is more effective for induction of apoptosis of breast cancer cells, compared to individual use of each. Thus, our data pave the way for new therapeutic options for suppressing breast cancer growth.

Keywords: Breast cancer; Lauryl gallate; Gene expression; Apoptosis; Tamoxifen

Introduction

Breast cancer is the most prevalent causes of cancer mortality, among the female population in developing countries (1). Tamoxifen (TAM) is an anti-estrogenic drug widely used for adjuvant therapy in breast cancer patients (2). In addition, TAM is also used to prevent breast cancer in healthy high-risk women (3). Long term TAM therapy may lead to serious side effects, such as cardiovascular events, hepatic injury, etc. (4, 5).

Hence, the use of herbal medicine with anticancer effects, in combination with low concentrations of TAM may help to reduce TAM side effects. Lauryl gallate (LG) is a Gallic acid derivative with antioxidant effects, used as a food additive (6). LG has been proven to protect human cells from oxidative damage and to inhibit tumor growth, without affecting normal cells (7). In addition, LG has also been reported to induce apoptosis and inhibit the proliferation of cancer cells (8, 9).

The combination therapy might be a significant advantage for cancer patients, to increase the response rate and reduce side effects of cancer therapeutics. Combination therapies, promoting the effectiveness of TAM, have been previously investigated in several studies, using compounds, such as caffeic acid phenethyl ester (10), green tea (11) and thymoquinone (12).

Concurrent administration of LG has additive or synergistic effects with TAM, widely has been used for the treatment of breast cancer.

Accordingly, in this study we investigated the efficacy of TAM and LG in breast cancer cells, with regard to apoptosis in these cells.

Materials and Methods

Cell culture

In this experimental study, the human breast cancer cell lines MCF-7 were obtained from the Pasteur Institute of Iran in 2017. Cells were routinely maintained in RPMI medium 1640 (Sigma, USA), containing 10% fetal bovine serum (FBS; Gibco, USA), 100 U/ml penicillin and 100 mg/ml streptomycin (Sigma, USA). Cells were incubated in a humidified incubator, containing 5% CO2 at 37 °C. Cell growth and cell division were daily controlled.

The study protocol was approved by the Shahrekord University of Medical Science Ethics Committee (IR.SKUMS.REC.1395.98).

Cell proliferation assay

The MTT assay was used to determine LG concentration, causing 50% cell death (IC50). Briefly, the cells were seeded at a density of 15×10³ in 96-well plates, and allowed to attach for 24 h, before incubation with drugs.

Cells were then exposed to different doses of LG alone and in combination with 10 µM final concentration of TAM (10), for 48 hours. For each sample and for every concentration three wells were loaded. Thereafter, 20 µl MTT (Sigma-Aldrich; 5 mg/ml) solution was added to each

well. After 4h incubation at 37 °C, the medium was replaced with 100 μL of 0.1 N HCl/isopropanol, in each well. The optical density of each well was determined at 570 nm, using an ELISA plate reader (Awareness, USA).

Treatment

The cells were seeded in a 6-well plate at a density of 1×10⁵ cells per well. After 24 h, they were treated by 10 μM final concentration of TAM alone and in combination with 200 μM final concentration of LG, for 24 and 48 hours. Another group was also considered with TAM and LG, in combination with 2 μM final concentration of EX-527, as SIRT-1 inhibitor. A group was considered as control group cultured without any drug treatment. We used EX-527 to examine the role of SIRT1 in cell apoptosis. By this, we determined the contribution and role of each gene (*BCL-2* and *SIRT-1*) in the apoptosis induction.

RNA isolation and cDNA synthesis

Total RNA was isolated from the cells, using Trizol (Invitrogen, California), according to the manufacturer's instructions. RNA was quantified, using a Nanodrop 2000 C spectrophotometer (Thermo Scientific-USA). Then cDNA was synthesized from 0.4 mg total RNA, using random primer and the cDNA synthesis kit (Thermo Fisher Scientific-Waltham, MA).

Real-Time Polymerase Chain Reaction (real-time PCR)

BCL-2 and SIRT-1 gene Expression were measured by quantitative real-time PCR. The experiments were performed, using Rotor-Gene 3000 real-time DNA amplification system (Corbett Research, Australia), and SYBR Green Method. Primers used for real-time PCR are listed in Table 1. Experiments were performed in triplicate, using 5 μL SYBR Green PCR Master Mix, 0.2 μL primer sets (10 μMol), 40 ng cDNA, and 3.6 μL nuclease-free H2O, in a final volume of 10 μL. The amplification was carried out under the following conditions: initial denaturing at 95 °C for 10 min, then 40 cycles of 95 °C for 15 sec, 59 °C for 20 sec and 72 °C for 25 sec. Quantitation of

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data was performed, using the comparative expression CT ($\Delta\Delta$ CT) method. GAPDH was used as a housekeeping gene. The gel electrophoresis

of PCR product and melting curve analysis were investigated to ensure the specificity of the reactions.

Table 1: Primer sequences and product length

Genes	Primer sequences (5'-3')	Amplicon length (bp)	GeneBank reference
BCL-2	Forward: GTGCTGAAGATTGATGGGATCG	118	NM_000633.2
	Reverse: TCAGTCTACTTCCTCTGTGATGTTG		
SIRT-1	Forward: TGCTGGCCTAATAGAGTGGCA	102	NM_012238.4
	Reverse: CTCAGCGCCATGGAAAATGT		
GAPDH	Forward: ACACCCACTCCTCCACCTTTG	112	NM-002046.5
	Reverse: TCCACCACCCTGTTGCTGTAG		

Flow cytometric analysis of cell apoptosis

The apoptosis kit (FITC Annexin V Apoptosis Detection Kit I, BD Biosciences, USA) was used to detect the apoptotic cells, according to the manufacturer's recommendation. Briefly, MCF-7 cells were seeded in the 6-well plates (1 \times 10⁵ cells per well) and incubated for 24 hours. Then they were treated similar to those previously mentioned. After 24 h and 48 h, the cells were trypsinized and washed in PBS. Overall, 100 µL of cells were transferred to a tube, 5 µL of FITCconjugated Annexin V (Annexin V-FITC) and 5 µL of propidium iodide (PI) were added, followed by incubation for 15 min at room temperature, in the dark. The cells were diluted by the binding buffer and directly analyzed, using flow cytometry (Partec GmbH, Munster, Germany).

Statistical analysis

All experiments were performed in triplicate. Data are expressed as mean \pm SEM. Statistical analysis was performed, using nonparametric Kruskal-Wallis test. Differences between two groups were investigated by the Mann-Whitney test. P<0.05 was considered statistically significant.

Results

Cell viability in MCF-7 cell line was determined by MTT assay (Fig. 1). The IC50 of the LG and in combination with 10 μ M TAM was about 200 μ mol/L. This concentration of LG was chosen for further experiments.

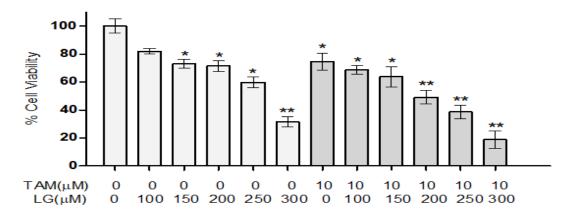


Fig. 1: Cell viability in MCF-7 cell line. Effect of different doses of lauryl gallate (LG) alone and in combination with 10 μ M tamoxifen (TAM) on cell viability, were determined by MTT assay for 48 hours. Data are presented as mean \pm SEM of three independent experiments *P<0.05, **P<0.01

We investigated the LG effects on BCL-2 and SIRT-1 expression in MCF-7 cells, in combination with TAM and EX-527(inhibitor of SIRT1), after 24 h and 48 hours. Our results showed TAM alone and in combination with 200 μ M of LG significantly decreased BCL-2 expression 2.64 \pm 0.75 and 6.38 \pm 1.9 –fold after 48 h, respectively, compared with the control group (P<0.001) (Fig. 2A). An increase in BCL-2 expression was

achieved when SIRT1 was inhibited by EX-527 (P<0.05).

After 48h, SIRT1 expression was significantly increased 1.67±0.22 and 2.47±0.34 fold by TAM alone and in combination with 200 μ M of LG, respectively (P<0.05) (Fig. 2B). EX527 inhibited deacetylase activity of SIRT1 in MCF-7 cell lines and thereby SIRT1 expression was decreased 2.71±0.76 fold when SIRT1 was inhibited by EX-527 (P<0.05).

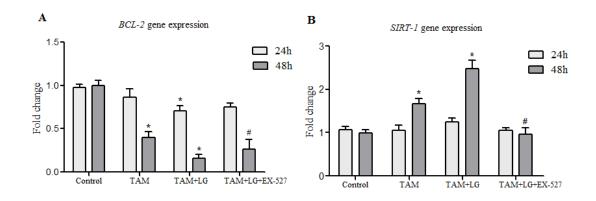


Fig. 2: BCL-2 and SIRT1 gene expression in MCF-7 cells, after 24 and 48 hours TAM: Tamoxifen (10 μM), TAM+LG: Tamoxifen (10 μM) in combination with 200 μM of LG. TAM+LG+EX-527: Tamoxifen (10 μM) in combination with 200 μM of LG and 2 μM EX-527

Data are presented as mean ± SEM of three independent experiments. *Compared to the control group, # compared to TAM + LG group

We further investigated the extent of apoptosis, by determining the percentage of Annexin V-stained cells. TAM alone and in combination with 200 μ M of LG increased the percentage of apoptotic cells to 15.79 \pm 2.81% and 60.67 \pm 6.23%, respectively, after 48 h (Fig. 3) (P<0.001). On the other hand, the increase in apoptosis was reduced by EX-527, as SIRT1 inhibitor (P<0.05). However, by SIRT1 inhibition, the apoptosis rate was decreased only about 6%.

Discussion

In this study, LG, as an antioxidant plays a role in reducing the growth of breast cancer cells. It does this by enhancing apoptosis through down-regulation of the anti-apoptotic gene expression, such as *BCL-2*.

The complex biology of cancer development requires relatively complex treatment approaches. Thus, the application of synergistic combinations of drugs, play an important role in the treatment of cancer (13).

In this study, the combination of LG and TAM is more potent, compared to either agent alone, in preventing breast cancer cell growth. This prevention occurs through apoptotic effects of LG and TAM.

Anti-apoptotic substances in combination with chemotherapy drugs can result in additive or synergistic effects. For example, the combination of green tea and TAM have a synergistic effect on inhibiting the growth of cancer cell (11). Moreover, curcumin in combination with TAM may have therapeutic benefit for prevention of breast cancer cells growth (14).

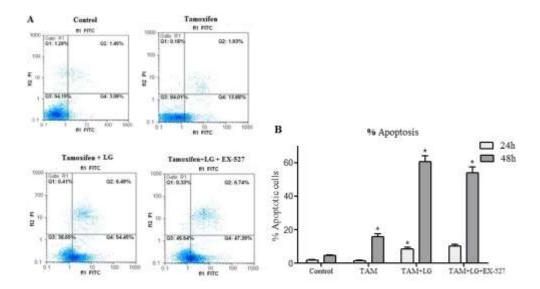


Fig. 3: Flow cytometry analysis of Annexin V-FITC/PI stained apoptotic cells. A). Dot plot profile of cells after 48 h. B) Bar diagram of percent apoptotic cells after 24 and 48 hours

TAM: Tamoxifen (10 μ M), TAM+LG: Tamoxifen (10 μ M) in combination with 200 μ M of lauryl gallate. TAM+LG+EX-527: Tamoxifen (10 μ M) in combination with 200 μ M lauryl gallate and 2 μ M EX-527

Data are presented as mean ± SEM of three independent experiments. *Compared to the control group, # compared to TAM + LG group

LG is a Gallic acid derivative with antioxidant effects, used as a food additive. It may be possible to use lower doses of chemical drugs and minimize the associated side effects, by using concurrent intake of herbal medicine that has fewer side effects, compared to chemotherapy drugs.

The synergistic effects of other chemotherapy drugs with Gallic acid derivative have also been reported in a study (15). Dhima et al. have shown pre-treatment with epigallocatechin-3-gallate (EGCG) sensitised leiomyosarcoma cells to cisplatin (15). Overall, synergistic combinations of herbal medicines and chemotherapy drugs may use for enhancing efficacy, reducing side effects, immune modulation, and abrogating drug resistance (13).

SIRT1 is a member of the sirtuin family involved in genomic stability and cell survival, acting by deacetylation of histone and affecting some cell cycle regulators. There is a controversy about the role of sirtoin, as a tumor suppressor gene or as a proto-oncogene (16, 17).

In our study, TAM increased the *SIRT-1* gene expression, and LG intensified this effect. This effect was confirmed, by using EX527 as inhibit-

ing the activity of *SIRT1*. But, by *SIRT1* inhibition, the apoptosis rate was decreased only about 6%. The synergistic effects of tamoxifen and lauryl gallate have a greater effect on the *BCL-2* than *SIRT1* gene at induction of apoptosis. But, it seems that the increase in TAM and LG -induced apoptosis, is partly due to the effect of *SIRT-1* on *BCL-2* expression. Kuo et al. showed that *SIRT1* suppresses breast cancer growth, through downregulation of the *BCL-2* gene (18).

Conclusion

Overall, compounds, such as LG strengthen the effects of drugs such as TAM, in inhibition of cancer cell growth. Further in vivo experiments needed, to confirm and validate these findings. Our data pave the way for using low-dose chemotherapy drugs and reducing the associated side effects of these drugs.

Ethical considerations

Ethical issues (Including plagiarism, informed consent, misconduct, data fabrication and/or falsification, double publication and/or submission,

redundancy, etc.) have been completely observed by the authors.

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Conflict of interest

The authors of the present work declare no conflict of interest.

References

- 1. Jemal A, Siegel R, Ward E et al (2009). Cancer statistics, 2009. *CA Cancer J Clin*, 59 (4): 225-49.
- 2. Furr BJ, Jordan VC (1984). The pharmacology and clinical uses of tamoxifen. *Pharm Ther*, 25 (2): 127-205.
- Fisher B, Costantino JP, Wickerham DL et al (1998). Tamoxifen for prevention of breast cancer: report of the National Surgical Adjuvant Breast and Bowel Project P-1 Study. J Natl Cancer Inst, 90 (18): 1371-88.
- 4. Alagozlu H, Cindoruk M, Unal S (2006). Tamoxifen-induced severe hypertriglyceridaemia and acute pancreatitis. *Clin Drug Investig*, 26 (5): 297-302.
- 5. Gao FF, Lv JW, Wang Y et al (2016). Tamoxifen induces hepatotoxicity and changes to hepatocyte morphology at the early stage of endocrinotherapy in mice. *Biomed Rep*, 4 (1): 102-6.
- Aruoma OI, Murcia A, Butler J et al (1993). Evaluation of the antioxidant and prooxidant actions of gallic acid and its derivatives. J Agricu Food Chem, 41 (11): 1880-5.
- 7. Jagan S, Ramakrishnan G, Anandakumar P et al (2008). Antiproliferative potential of gallic acid against diethylnitrosamine-induced rat hepatocellular carcinoma. *Mol Cell Biochem*, 319 (1-2): 51-9.
- 8. Cheng CH, Cheng YP, Chang IL et al (2016).

 Dodecyl gallate induces apoptosis by

- upregulating the caspase-dependent apoptotic pathway and inhibiting the expression of anti-apoptotic Bcl-2 family proteins in human osteosarcoma cells. *Mol Med Rep*, 13 (2): 1495-500.
- de Cordova CA, Locatelli C, Assunção LS et al (2011). Octyl and dodecyl gallates induce oxidative stress and apoptosis in a melanoma cell line. *Toxicol In Vitro*, 25 (8): 2025-34.
- Motawi TK, Abdelazim SA, Darwish HA et al (2016). Modulation of tamoxifen cytotoxicity by caffeic acid phenethyl ester in MCF-7 breast cancer cells. Oxid Med Cellular Longev, 2016: 3017108
- 11. Sartippour MR, Pietras R, Marquez-Garban DC et al (2006). The combination of green tea and tamoxifen is effective against breast cancer. *Carcinogenesis*, 27 (12): 2424-33.
- 12. Ganji-Harsini S, Khazaei M, Rashidi Z et al (2016). Thymoquinone could increase the efficacy of tamoxifen induced apoptosis in human breast cancer cells: An in vitro study. *Cell J*, 18 (2): 245-54.
- 13. Malongane F, McGaw LJ, Mudau FN (2017). The synergistic potential of various teas, herbs and therapeutic drugs in health improvement: a review. *Sci Food Agric*, 97 (14): 4679-4689.
- 14. Jiang M, Huang O, Zhang X et al (2013). Curcumin induces cell death and restores tamoxifen sensitivity in the antiestrogenresistant breast cancer cell lines MCF-7/LCC2 and MCF-7/LCC9. *Molecules*, 18 (1): 701-20.
- Dhima IT, Peschos D, Simos YV et al (2018).
 Modulation of cisplatin cytotoxic activity against leiomyosarcoma cells by epigallocatechin-3-gallate. Nat Prod Res, 32 (11): 1337-1342.
- Kabra N, Li Z, Chen L et al (2009). SirT1 is an inhibitor of proliferation and tumor formation in colon cancer. *J Biol Chem*, 284 (27): 18210-7.
- 17. Santolla MF, Avino S, Pellegrino M et al (2015). SIRT1 is involved in oncogenic signaling mediated by GPER in breast cancer. *Cell Death Dis*, 6 (7): e1834.
- 18. Kuo S-J, Lin H-Y, Chien S-Y et al (2013). SIRT1 suppresses breast cancer growth through downregulation of the Bcl-2 protein. *Oncol Rep*, 30(1): 125-30.

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