

Cross-Reacting Material 197, a Specific Inhibitor of HB-EGF, and Its Anticancer Effects

Maryam Tanhapour^{1,2}, Abolfazl Golestani¹, Asad Vaisi-Raygani², Mahdi Aminian^{1*}

1

1 Department of Clinical Biochemistry, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran

2 Department of Clinical Biochemistry, School of Medicine, Kermanshah University of Medical Sciences, Kermanshah, Iran

Corresponding Author: Mahdi Aminian

Associate Professor of Clinical Biochemistry, Department of Clinical Biochemistry, School of Medicine, Tehran University of Medical Sciences, Poursina Ave. Tehran, Iran

Email: amminian@tums.ac.ir

<https://orcid.org/-3997-0002-00006954>

Tel: (+98)2164053296

ABSTRACT

Genetic heterogeneity and metastasis remain the most challenging aspects of cancer treatment. Recent studies have introduced a mutant of diphtheria toxin, cross-reacting material 197 (CRM197), as a new promising biological anticancer drug to improve cancer therapy in patients previously resistant to chemotherapy. Weak toxicity of CRM197 accounts for the stimulation of cell apoptosis and the antitumor effect. Increasing evidence has indicated that the expression of heparin-binding epidermal growth factor-like (HB-EGF) growth factor increases in most cancer cells, and CRM197 is its specific inhibitor. This study has focused on the structure, properties, and anticancer activity of CRM197.

Keywords: Cancer, Cross-reacting materials 197 (CRM197), Diphtheria Toxin (DT), Epidermal growth factor receptor (EGFR), Heparin-binding EGF-like growth factor (HB-EGF)

Diphtheria Toxin:

During the 1820s, corynebacterium diphtheriae was identified as a pathogen causing a distinct form of sore throat (1). Visualization and isolation of the organism were performed in 1883 and 1884 by Klebs and Loeffler, respectively (1, 2). The DT gene expression is presented by lysogenic corynebacterium diphtheriae, which is regulated by the diphtheria toxin repressor (DtxR), a member of a family of metal ion-regulated repressors (1, 3, 4). Qian et al. reported that high levels of iron coupled with DtxR bind to the gene's operator and suppress the DT expression (3). Diphtheria toxin precursor includes a 26-amino acid signal sequence, and following cleavage by the processing enzyme, produces a 535-amino acid single-chain protein (58kDa). This structure has been discovered by Roux and Yersin in 1888 (5-8). Proteolytic cleavage of the toxin by trypsin results in the generation of two fragments, an N-terminal, enzymatically active fragment A of 193 aa (DTA, 21-kDa) involved in the inactivation of elongation factor 2 (EF2), and a C-terminal, fragment B of 342 aa (DTB, 37-kDa) which includes both the transmembrane (T) and the receptor-binding (R) domains (Fig. 1) (9-11).

DTA and DTB are connected via a 14-amino acid pro-

tease-sensitive loop and a disulfide bond between Cys168 and Cys201 (12). Murphy reported that the ADP-ribosyltransferase activity of the toxin is activated by proteolytic “nicking” of the α-carbon backbone at Arg193 in disulfide containing loop (13). DT binding to the human heparin-binding epidermal growth factor precursor (pro-HB-EGF) as a high-affinity receptor for the toxin and to the specific DT receptor is mediated by the R-domain (residues 432–535) (13-17). The T domain in the plasma membrane consists of nine helices organized in one amphipathic and two hydrophobic layers (12). These three layers form a pore across the endosomal membrane, in which the second layer stabilizes the structure (12).

Clathrin-mediated endocytosis is involved in the endocytosis of DT bound to its receptor and its internalization into early endosomal vesicles (18). Although protease furin is responsible for DTA and DTB domain cleavage in the endosomal vesicles, their linkage remains intact via a disulfide bond between Cys168 in DTA and Cys201 in DTB (19). Translocation of DTA from the endosome to the cytosol requires the insertion of the T domain into the endosomal membrane (20). Thus, the key protein components of clathrin on the vesicle membrane are substituted with some different protein components

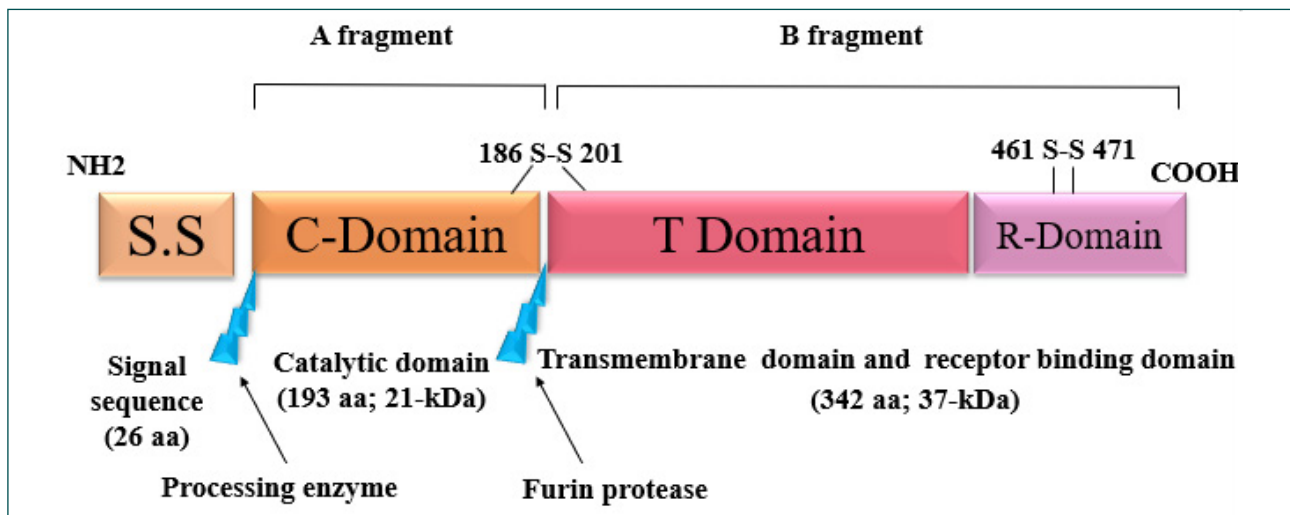


Figure.1. Diphtheria toxin structure. Diphtheria toxin synthesized in precursor form following the cleavage of a signal sequence by the processing enzyme has been observed in a Y-shaped molecule including A and B fragments. Fragment A includes the catalytic domain (C domain) of DT, cleaved by furin protease, and fragment B consists of two fragments, transmembrane (T) and receptor binding (R) domains. Two disulfide bonds have been demonstrated in DT structure, Cys186 to Cys201, between fragments A and B and an internal disulfide bridge (Cys461 to Cys471) within fragment B (11,12).

such as ADP-ribosylation factor 1 (Arf-1), coat protein complex I (COPI), ras-related protein (Rab-5), and vacuolar H⁺-ATPase (V-ATPase) (20). The V-ATPase activity leads to acidification of endosomes, unfolding of T domain, insertion into the endosomal membrane, formation of 18–22Å cation-selective membrane pore, and finally, DTA translocation from endosome to cytosol (12, 21). In this case, according to Murphy, the catalytic domain (C-domain) of diphtheria toxin located in fragment A is threaded through the pore by a process facilitated by a cytosolic translocation factor (CTF) complex. In a second hypothesis, he suggested that the nascent chaperone-like activity of the partially unfolded T domain mediates the autonomous delivery of the C-domain across the membrane (13). The rate-limiting step for diphtheria toxin activation is reducing the disulfide bond between DTA and DTB that is catalyzed by thioredoxin reductases (TrxRs) or protein disulfide isomerase (PDI) (22). Following the delivery of DTA into the cytoplasm, activation of enzyme depends on refolding it and subsequently NAD⁺-dependent ADP-ribosylation of EF-2, leading to the inhibition of protein synthesis and ultimately apoptosis (Fig. 2) (23).

The current study has focused on the structure, proper-

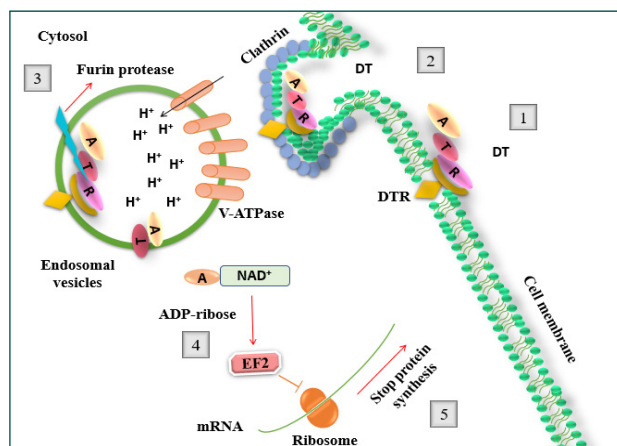


Figure 2. Mechanism of action of diphtheria toxin. DT binding to a receptor on the surface of sensitive cells (1) leads to internalization into early endosomal vesicles via clathrin-mediated endocytosis (2, 3). Following the DT insertion into endosomal vesicles, the key protein components of clathrin are substituted with different proteins such as V-ATPase. The V-ATPase activity provides the low endosomal pH essential for inserting the T domain into the endosomal membrane and the membrane pore formation. T domain helps in the delivery of the A domain to the cytosol (3). Once the A domain is released into the cytoplasm of sensitive cells via interaction with NAD⁺ (4) and transformation of an adenosine diphosphate ribosyl (ADPR) moiety from NAD⁺ to EF2, irreversibly terminates the protein synthesis (5). DT: diphtheria toxin, DTR: diphtheria toxin receptor (HB-EGF), V-ATPase: vacuolar H⁺-ATPase, NAD; nicotinamide dinucleotide, EF2: elongation factor 2. The current study focuses on the structure, properties, and anticancer activity of CRM197.

ties, and anticancer activity of CRM197.

Structure of CRM197

The nontoxic or partially toxic forms of diphtheria toxin, including CRMs 197, 228, and 176 with the same size as an intact toxin, results from the missense mutations (24). In addition, CRMs 30 and 45 with lower molecular weight than the native molecule and an N-terminal fragment may result from chain-termination mutations, according to Collier (24). In 1973, for the first time, Uchida et al. isolated these five mutant proteins from *C. diphtheriae* strain C7(β) induced by ultraviolet.

CRM 40, another nontoxic form of DT toxin, with a C to T transition resulting in the generation of (TAA) termination codon, lacks the last 149 carboxyterminal amino acid residues (approximately 17kDa) and the ability to bind to the cell surface (25). The nontoxic effect of CRM30 is due to the absence of binding and translocation domains, but its enzymatic activity remains intact (26, 27). Decreased binding capacity and enzymatic activity of CRM176 and CRM228 are attributed to the presence of several mutations and changes in the structure of Fragment A (26). The G/A transition in the DTA sequence leads to Gly52/Glu substitution and lack of enzymatic activity, perhaps due to alteration in the NAD⁺ binding site (28). Different methods for structure analysis of CRM197 have revealed two main domains; fragment A or the catalytic domain and fragment B or the transmembrane domain, which contains two subdomains, an all-β-sheet structure that forms a subdomain for binding to the HB-EGF and anine α-helices which forms transmembrane subdomain for translocation into the target cell. Two S-S bonds have been shown in the protein structure, the former (Cys186 to Cys201) links A and B fragments together, and the latter joins Cys461 to Cys471 within fragment B (29).

Diphtheria toxin and CRM197 interaction with cell surface

In addition to interaction with protein receptors, DT binds to the negative charge of phosphate groups of phospholipids (28). In other words, even in the absence of a receptor, DT interacts with phospholipids with high affinity (30).

Due to alteration in the protonation state of the imidazole rings and carboxylic residues within both A and B, the pH ranges from neutrality to 5, and consequently, the conformational changes occur in the DT structure. Thus, pH has a vital role in DT translocation across the membrane. Papiniet al. showed that at neutral pH, DT interacts with the lipid bilayer surface only. However, at low pH, it binds to the hydrocarbon chains of phospholipids. Moreover, at low pH, CRM45 with only an N-terminal fragment is significantly less toxic than the native diphtheria toxin. These results suggest that the COOH-terminal segment of DT has a crucial role in interactions with cell receptors and the translocation of toxin across the membrane (30). Consistent with this, the importance of acidic conditions in penetration of diphtheria toxin into the cell has been reported by Sandvig et al. They reported that protein synthesis is blocked at low pH. In cells exposed to DT at pH 4.5 for 10 min, protein production decreased as fast as in the cells exposed to the toxin with 1000 times concentration at neutral pH (31). In addition, Sandvig et al. hypothesized that in an acidic environment, the possible mechanism by which the DT enters into the cell is as follows; after entering the cell through pinocytosis, the toxin breaks down to its constituent amino acids in lysosomes, but some of them pass directly through vesicle membrane and escape from digestive lysosomal enzymes as soon as the vesicles become acidic. Low pH may also change the conformation of some protein structures in the plasma membrane, including receptors, or may induce opening of different channels and even conformational changes in DT. When the channels are opened, DT, hormones, and some growth factors flow into the cell, and significant amounts of ions and intracellular compounds leak out of the cell and ultimately lead to cell damage (31).

The CRM197 interaction with membrane lipids and cell surface receptors is stronger than DT (32). Another study has reported that both CRM197 and DT interact with diphtheria toxin receptors with the same affinity (28).

The partially nontoxic properties of CRM197 are due to similar DT or CRM197 B fragment effects that competitively inhibit the DT interaction with the cell surface receptor and enzymatically inactive A fragment (27). Also, the permeability of CRM197 into the cell is temperature- and pH-independent, suggesting that CRM197 has an open conformation similar to the acidified DT, in which the R domain is separated from the C and T domains (33, 34).

Diphtheria toxin interaction with HB-EGF

DT interacts with HB-EGF precursor at the surface of various mammalian cell types (34). The specific DT receptor, mainly found in monkey Vero cells, was first isolated from the conditioned medium of macrophages and macrophage-like U937 cells (35). This particular receptor on the cell surface is similar to proHB-EGF with an identical amino acid sequence (35).

For the first time, EGF was isolated from parotid glands of male mice and purified from human urine. Because of suppressive effects on gastric acid secretion in humans, it was termed urogastrone (36, 37). Epidermal growth factor (EGF), a 53 aa protein (6 kDa) with three internal disulfide bonds, is secreted mainly by monocytes, macrophages, vascular endothelial, and smooth muscle cells. The protein has a high affinity for heparin, termed heparin-binding epidermal growth factor-like growth factor (HB-EGF) (38, 39). Moreover, HB-EGF, a member of the EGF family, is generated as a membrane-anchored precursor molecule (proHB-EGF). This precursor protein is cleaved by ADAM (which acts as a disintegrin and is a metalloprotease) in a process named ectodomain shedding leading to generation of soluble HB-EGF. ProHB-EGF cleavage is made by either matrix metalloproteases 7 (MMP7, expressed in tumor cells) at a position similar to that of ADAM or at a position in the N-terminal portion of the EGF-like domain (fig. 3) (40-44).

According to the previous studies, amino acid sequence within the EGF-like domain of the receptor (residues

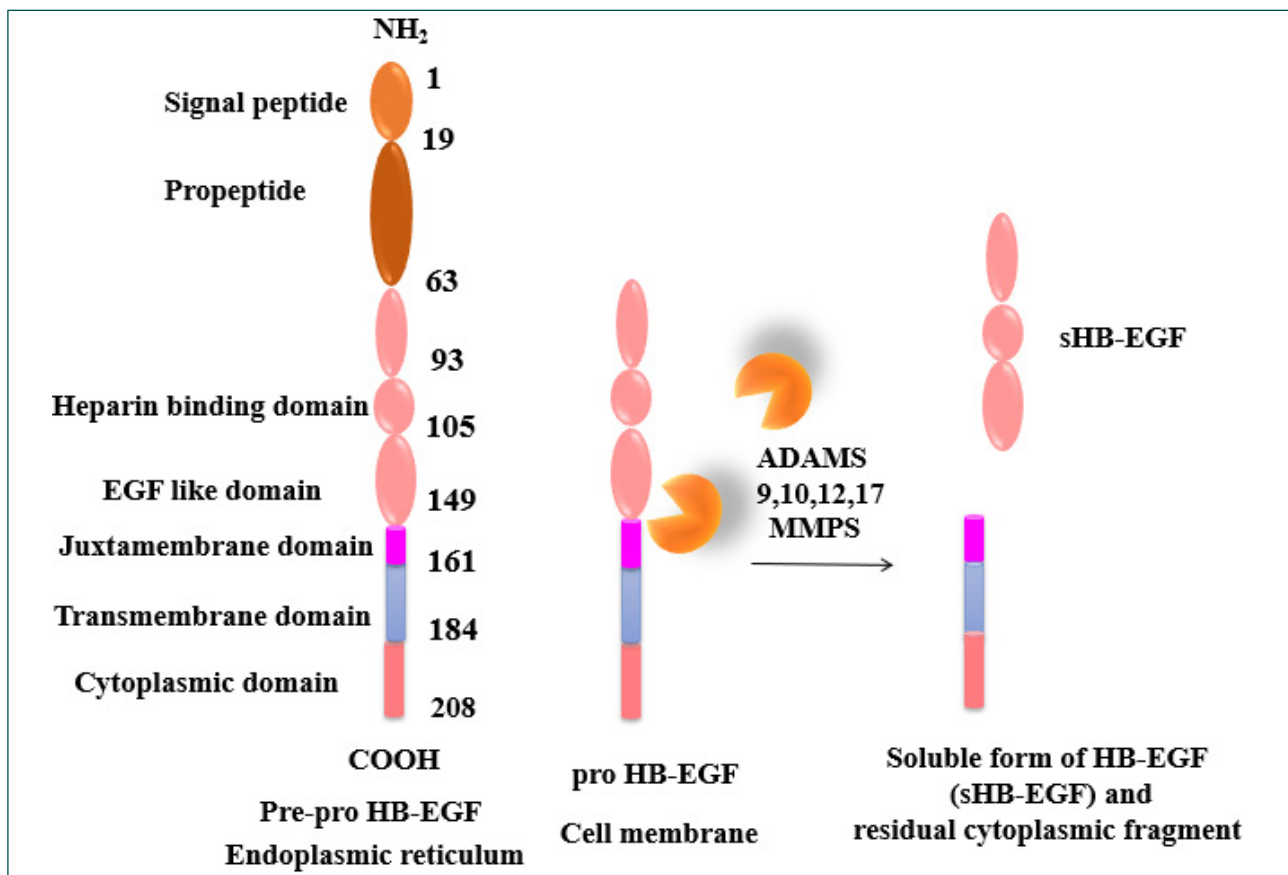


Figure.3. HB-EGF-like growth factor structure and ectodomain shedding of epidermal growth factor receptor (EGFR) ligand. Human HB-EGF-like growth factor is synthesized as a 208-amino acid membrane-anchored precursor molecule (pro-HB-EGF) cleaved by ADAM or MMPs to yield the mature protein or the soluble form of approximately 86 amino acids. ADAM: A disintegrin and metalloprotease, MMPs: matrix metalloproteases (45).

115-148) is specifically essential for DT binding (27). In addition, ATP and other polyphosphates can inhibit DT toxicity by binding to a cationic region within the receptor-binding domain of the B-fragment, i.e., P-site (45). Based on this, Eidelset al. postulated that the P-site on the toxin molecule should be involved in both polyphosphate and receptor binding. Also, highly cationic molecules such as histone, lysozyme, poly (L-ornithine), putrescine, spermidine, and spermine may inhibit DT from binding to HB-EGF due to the similar effects of P-site (45). On the other hand, Brooke et al. reported that the negatively charged 141 Glu of the DT receptor and positively charged 516 Lys of diphtheria toxin have the most critical role in crucial interactions between the toxin and the receptor. Besides, the replacement of 141 Glu with His in the pro-HB-EGF binding site results in a significant reduction in

affinity binding and the toxicity of DT (46).

Because the DT binding to HB-EGF forms a complex with DRAP27, the monkey homolog of human CD9 antigen, and regulates the number of functional HB-EGF binding to the toxin. Thus, the 27-kD diphtheria toxin receptor-associated protein (DRAP27) is considered an essential factor in cell sensitivity to DT (16, 27).

Epidermal growth factor receptor is known as a protein tyrosine kinase receptor (PTKR). The receptors of the tyrosine kinase (RTK) family include four members: ErbB1 or (HER1), ErbB2 (Neu or HER2), ErbB3 (HER3), and ErbB4 (HER4) (47). Ligand binding leads to homo- or hetero-dimerization of the receptors and the kinase activity (48). EGFR signaling pathway results in the transition from the G1 phase of the cell cycle to the S phase and several different biological responses such as

migration, motility, proliferation, cellular differentiation, survival, and apoptosis (49). EGFR, firstly reported by Purba et al. (50), is a single-pass transmembrane protein consisting of an extracellular domain, a transmembrane domain, a juxtamembrane (JM) segment, a kinase domain, and a C-terminal regulatory tail. Various ligands such as epidermal growth factor (EGF), transforming growth factor- α (TGFA), amphiregulin (AREG), epigen (EPGN), betacellulin (BTC), heparin-binding EGF (HB-EGF), and epiregulin (EPR) have been known as activators for EGFR (51). The extracellular ligand-binding domain of EGFR has four motifs I-IV. The I (L1) and III (L2) members of the leucine-rich repeat superfamily are characterized by β -helix solenoid structure, which interacts with their ligands, whereas II (CR1) and IV (CR2) motifs contain cysteine-rich regions (50). This structure is followed by one transmembrane domain (25 aa) and

an intracellular domain (about 550 aa) with 20 tyrosine residues which 12 of which are targeted for phosphorylation (52). Phosphorylated tyrosine residues serve as docking sites for soluble or membrane-anchored effector proteins and stimulate multiple signal transduction pathways, mediated by Ras/Raf/mitogen-activated protein kinase (MAPK), phosphoinositide-3-kinase (PI3K)/Akt, and phospholipase C (fig. 4) (50).

CRM197 and anti-Cancer effect

EGFR-mediated signaling must be tightly regulated to provide proper cell growth, proliferation, differentiation, survival, or wound healing (53). EGFR signaling depends on the regulation of gene expression and liberation of their ligands (54). Excess EGFR signaling, increased production of ligands, overproduction of the EGFR protein, EGFR gene mutations, and lack of EGFR

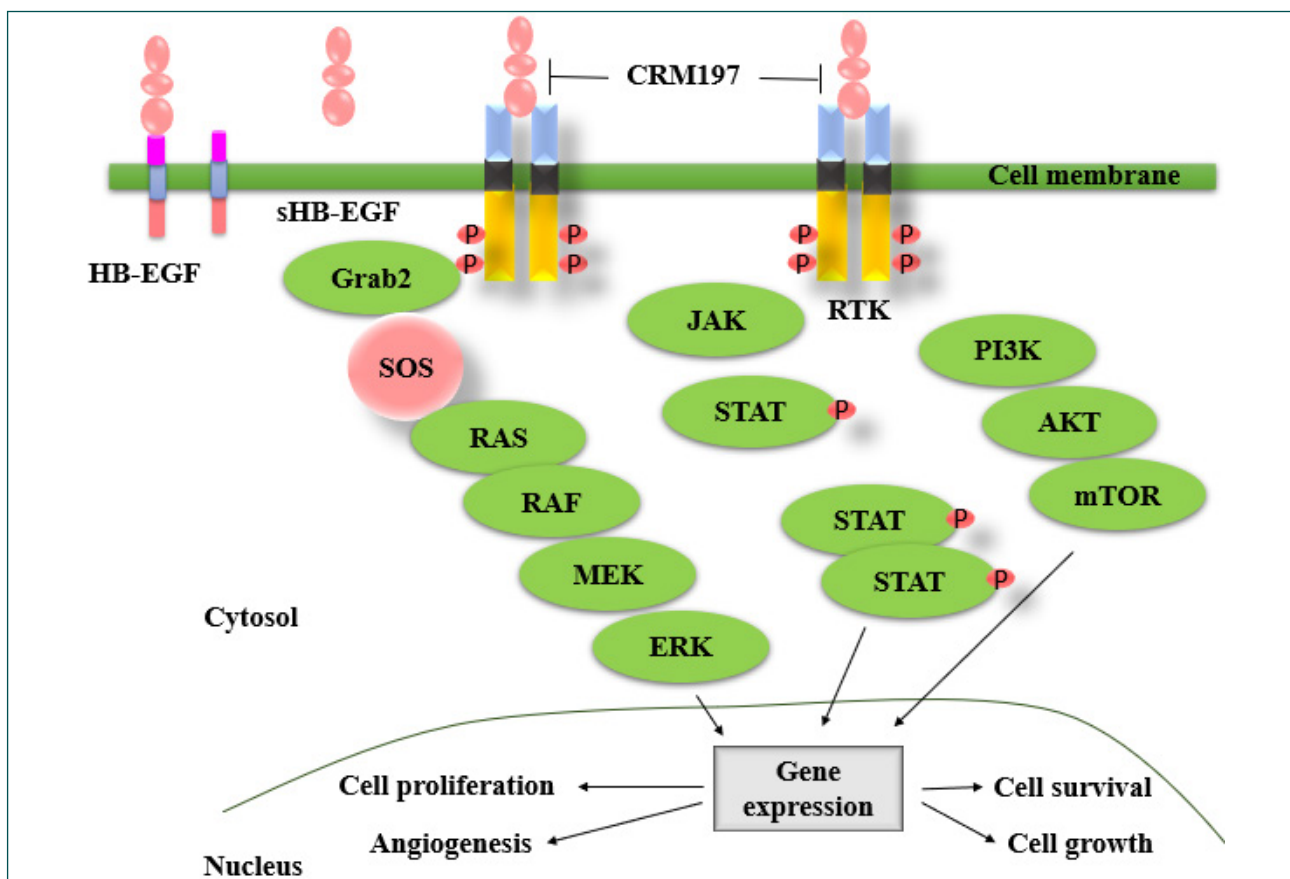


Figure 4. CRM197 as an inhibitor that abrogates the EGFR activation pathway. sHB-EGF: soluble Heparin-binding epidermal growth factor-like growth factor, RTK: receptor tyrosine kinase, EGFR: epidermal growth factor receptor, CRM197: cross-reacting material.

downregulation may lead to cancer development and progression (55, 56). CRM197 binds to HB-EGF, inhibits its mitogenic activity, and inhibits the ectodomain shedding of proHB-EGF (fig. 4) (40, 57). In addition to the partially toxic nature of the molecule, CRM197 acts as an immunological adjuvant and carrier (58-64). Therefore, CRM197 may improve the efficacy of cancer therapy when combined with chemotherapeutic drugs (65).

Like the intact DT, none of the EGFR ligands except for HB-EGF can interact with CRM197 (54). Therefore, HB-EGF suppression leads to inhibition of metastasis, survival, cell adhesion, invasion, angiogenesis, and tumorigenicity of this growth factor (54).

The antitumor effects of CRM197 in the treatment of different cancer types are listed in table 1. Buzzi et al. investigated the anticancer effect of CRM197 on patients with various advanced tumors. According to the results, subcutaneous injection of CRM197 accompanied by tolerable side effects such as inflammatory skin reaction, increased neutrophil count, and significantly enhanced serum levels of TNF- α (66). Yagiet al. assessed the synergistic effects of paclitaxel as an antitumor chemotherapy drug and CRM197 as an inhibitor of HB-EGF on ovarian cancer (OC). The results of in vitro and in vivo studies suggested that resistance to paclitaxel can occur as follows: Although paclitaxel stimulates the proapoptotic signals such as c-Jun N-terminal kinase (JNK) and p38 MAPK, it activates antiapoptotic signals via enhancement of the ectodomain shedding of proHB-EGF and activation of the ERK and Akt signaling. After the ectodomain shedding of proHB-EGF, the carboxyl-terminal fragments of HB-EGF (HB-EGF-CTFs) translocate into the nucleus and trigger cell growth signals. CRM197 blocks the EGFR and subsequent ERK and Akt signaling and may suppress the HB-EGF-CTFs translocation into the nucleus (67).

Epithelial OC metastasis occurs via multiple pathways such as the peritoneal cavity, blood, or lymphatic vessels termed transcoelomic, hematogenous, or lymphatic metastasis, respectively (68, 69). The multi-step process in OC metastasis is divided into several distinct stages; (a) infiltration and survival of cancer cells in the peritoneal

fluid, (b) adherence to the peritoneal surface, and finally (c) acquisition of potential for motility, angiogenesis, and invasion into the peritoneum to subsequent migration and proliferation in other organs resulting in colonization (70). Survival of individual ovarian tumor cells depends on the secretion of lysophosphatidic acid, HB-EGF, and vascular endothelial growth factor (VEGF) into the ascitic fluid. CRM197 has adverse effects on cell proliferation, growth, and survival in peritoneal fluid of patients with OC. CRM197 inhibits cell adhesion via integrins, protein tyrosine kinase 2 (PTK2), which is known as focal adhesion kinase (FAK), and EGF R (54, 71). CRM197 decreases the VEGF and interleukin 8 (IL-8) gene expression significantly and reduces the metastatic potential of OC cells (72). Yotsumoto et al. reviewed the efficacy of ligand-based targeting for the EGF system in cancer therapy. They reported that although DT interacts with the EGF domain of HB-EGF and inhibits cell proliferation, it is not considered an HB-EGF inhibitor because of the high toxicity effects against normal cells (73). In contrast, CRM197 interacts with HB-EGF more strongly than DT or at least as strong as a native molecule without significant side effects (35). Coadministration of CRM197 with paclitaxel blocks the gene expression of ERK and the Akt signaling pathway. It activates p38, c-Jun N-terminal kinases (JNK), and MAPK signaling pathways involved in cell proliferation and apoptosis. Therefore, combining chemotherapeutic drugs with CRM197 enhances apoptosis rate and inhibits cell proliferation (74).

According to Martarelli et al. (75), the therapeutic effect of CRM197 in adrenocortical carcinoma is attributed to its inhibitory effect on HB-EGF, its residual toxicity, and its potent properties in triggering the immune response. The role of HB-EGF-EGFR signaling in gastric cancer development and significant improvement in anticancer effect of coadministration of 5-FU, cisplatin, or paclitaxel with CRM197 was approved in both in vitro and in vivo experiments performed by Sanui et al. (76). Consistent with these results, Yotsumoto et al. reported that HB-EGF was predominantly expressed in breast cancer, and CRM197 was introduced as both a specific inhibitor

Type of cancer	Drug dosage and type of injection	Human, animal or cell culture	The aims of a study	REff
			Results	
Various* advanced cancer	According to the patient's degree of immunological reactivity to DT/CRM197 (none, moderate, or high) three different dosages (1.7, 2.6, or 3.5 mg/day) of CRM197 injected subcutaneously in the abdominal wall, on alternate days, for 6 days.	outpatients	Anti-tumor effect of CRM 197	Buzzi 2004
			Different degree of inflammatory response and significant increase in circulating neutrophils and serum TNF- α concentration was observed.	
Ovarian cancer	10 nM-10 μ M concentrations of paclitaxel and 1 μ g/ml CRM197 for 24 hr	SKOV3 and SK-HB-1 cells	The effect of paclitaxel and CRM197 on cell proliferation	Yagi 2009
			The more effective suppressed SKOV3 cells viability was observed following paclitaxel co-administration with CRM197 than paclitaxel only.	
	10 nM-1 μ M concentrations of paclitaxel in the presence or absence of 1 μ g/ml CRM197 for 48 hr	SKOV3 and SK-HB-1 cells	The effect of paclitaxel and CRM197 on apoptosis	
			The effect of concurrent administration of paclitaxel and CRM197 on both SKOV3 and SK-HB-1 cells death was significantly higher than the paclitaxel only.	
	Dose-dependent administration of 5 mg/kg and 50 mg/kg of CRM197 in nude of mice	Tumorigenesis by SKOV3 or SK-HB-1 cells injection into mice	In vivo anti-tumor effects of CRM197	
			The tumor formation was partially inhibited following administration of 5 mg/kg CRM197 and completely inhibited following administration of 50 mg/kg CRM197.	
	Co-administration of 20 mg/kg and 10 mg/kg of paclitaxel in nude mice by	Tumorigenesis by SKOV3 or SK-HB-1 cells injection into mice	In vivo anti-tumor effects of paclitaxel	
			Tumor formation was suppressed by SKOV3 cells (20 mg/kg), while this effect was not seen about SK-HB-1 cells (20 mg/kg), SKOV3 cells and SK-HB-1 cells (10 mg/kg).	
Co-administration of 10 mg/kg paclitaxel and 5 mg/kg CRM197	Tumorigenesis by SKOV3 or SK-HB-1 cells injection into mice	In vivo anti-tumor effects of paclitaxel and CRM197		
		Tumor growth by both SKOV3 cells and SK-HB-1 cells Completely was suppressed.		
Adrenocortical carcinoma	1 mg/week CRM197 every week was injected intraperitoneally	Tumorigenesis by Subcutaneous injection of H95R and SW-13 cells into mice	Effect of CRM197 on tumor formation	Martarelli 2009
			At presence of CRM197 tumor formation by H295R and SW-13 cells was significantly suppressed	

Adrenocortical carcinoma	For cell proliferation assays and cell cytotoxic assays CRM197 dissolved in DMSO at various concentrations: 0,1, 10 and 100 g/ml.	H295R and SW-13 cells	In vitro anti-tumor effect and cytotoxicity of CRM197	Martarelli 2009
			At a concentration higher than 1 g/ml of CRM197 the growth of tumor cells was considerably suppressed and at presence of 100 g/ml of CRM197 apoptosis was accrued.	
	CRM197 (0, 1, 10 and 100 g/ml)	H295R cells	Effects of CRM197 on cell migration and invasion	
			The cell invasion significantly was suppressed following the treatment with noncytotoxic doses of CRM197 and completely was inhibited at cytotoxic dosage of CRM197 (100 g/ml).	
	100 g/ml of CRM197 for 24 h	H295R and SW-13	CRM197 effect on apoptosis	
		Cell death was seen following treatment with CRM197 at both H295R and SW-13 cells		
		Tumor cell implanted in nude mice.	CRM197 effect on angiogenesis	
			Angiogenesis was suppressed in subcutaneous tumors.	
Gastric Cancer	Transfection of siRNA at concentration of 25 nM and incubation for 72 h for HB-EGF, amphiregulin and EGFR, and after incubation with CRM197 (1 µg/ml) and inhibitory antibody against AR (AR;10 µg/ml) and EGFR (10 µg/ml) and incubation for 72 h	NUGC3 and MKN28 cells	Investigation of the tyrosine phosphorylation of EGFR, ERK and AKT	Sanui 2010
			At presence of a siRNA, inhibitory anti-EGFR antibody or CRM197 separately, the EGFR, ERK and AKT phosphorylation in NUGC3 and MKN28 cells was decreased but no change in AR phosphorylation was observed at presence of amphiregulin siRNA.	
	No alteration in EGFR, ERK and AKT phosphorylation was seen at presence the inhibitory anti- AR antibody.			
			The significantly high rate of cell death was observed at presence of the CRM197 and inhibitory anti-EGFR antibody.	
Different concentrations (0-10 µM) of 5-FU, cisplatin or paclitaxel	NUGC3 And MKN28 cells	5-FU, cisplatin or paclitaxel effect on concentration of HB-EGF and apoptosis	A significant increase in HB-EGF expression and apoptosis was observed following the treatment with paclitaxel at concentrations of 1 nM and 10 nM in compared to 5-FU and cisplatin in two types of cells.	

Gastric Cancer	1 µg/ml of CRM197 with different concentration (0-10 µM) of 5-FU, cisplatin or paclitaxel	NUGC3 And MKN28 cells	CRM197, 5-FU, cisplatin or paclitaxel effect on concentration of HB-EGF cell apoptotic rate The synergistic antitumor effect was reported between CRM197, 5-FU, cisplatin or paclitaxel.	Sanui 2010
	CRM197 (50 mg/kg) or paclitaxel (10 mg/kg), or their combination was injected intraperitoneally each week for 6 weeks into mice	Tumorigenesis by Injection of NUGC3 And MKN28 cells into mice	Investigation of antitumor effects following co-administration of CRM197 and paclitaxel in xenografted mice CRM197 alone significantly inhibited tumor growth. Partial suppression of NUGC3 cell growth and no inhibition of MKN28 cell growth reported following the treatment with paclitaxel. At presence the CRM197 and paclitaxel tumor formation by both NUGC3 and MKN28 cells completely was suppressed.	
Breast cancer	1 µM of cetuximab or 100 nM of CRM197	BT549 and MDA-MB-231 cells	Anti-tumor effect of cetuximab or CRM197 Unlike the treatment with cetuximab, the significant higher rate of cell death and lower HB-EGF expression and EGFR, ERK and AKT activation was reported following the treatment with CRM197.	Yotsumoto 2010
	CRM197 (50 mg/kg) was injected intraperitoneally every day for 10 days after the tumor volume exceeded 100 mm ³	Tumorigenesis by Injection of MDA-MB-231 cells into NOD/SCID mice	Anti-tumor effect of CRM197 Completely tumor formation was suppressed	
	CRM197 (100 nM), trastuzumab (1 µM) trastuzumab (1 µM) plus recombinant human HB-EGF (rHB-EGF) (1 µg/ml)	SK-BR-3 and BT474 cells	HB-EGF is involved in trastuzumab-resistant signals At presence of trastuzumab or CRM197 increase in cell death and decrease in HB-EGF expression was induced, but addition of HB-EGF suppressed the cell death following treatment with trastuzumab.	
Acute lymphoblastic leukemia T-ALL	10 µg/ml of CRM197 for 72 h	Jurkat E6-1 cells compared with untreated cells	CRM197 effect on HB-EGF expression and apoptosis in T-ALL cells Although CRM197 induces apoptosis and decreases HB-EGF protein expression in Jurkat E6-1 cells, no association was found between RNA expression and treatment with CRM197.	Kunami 2011

Acute lymphoblastic leukemia T-ALL	25 nM doxorubicin and/or 10 µg/ml CRM197 for 72 hours	Jurkat E6-1 cells compared with untreated cells	Antitumor effects of doxorubicin co-administration with CRM197	Kunami 2011
			At presence of CRM 197 or doxorubicin separately the rate of apoptosis was 10.8±3.2% and 65.1±2.5%, respectively. Co-administration of CRM 197 and doxorubicin increased the rate of apoptosis by 75.6±2.4%.	
Glioma	5 mg/ml of cisplatin combined with 1mg/ml of CRM197	Glioma cell (U251) and human astrocytes (HA)	Effect of cisplatin and CRM197 on cell growth	Wang 2012
			The treatment with cisplatin and CRM 197 significantly blocked the U251 cells growth compared to HA cells.	
	No information has been mentioned	U251 cells	Effects of co-administration of cisplatin with CRM197 on apoptosis	
			Following the treatment with both cisplatin and CRM197 significantly cell apoptosis increased in compared to cisplatin alone.	
	No information has been mentioned	U251 cells	CRM197 suppressed the Akt phosphorylation induced by cisplatin	
			At presence the cisplatin Akt phosphorylation was increased, while treatment with either CRM197 or combination of cisplatin with CRM197 considerably blocked the Akt phosphorylation compared with the control group.	
10 mmol/l of LY294002, 1 mmol/l of wortmannin, cisplatin combined with CRM197 and combination of cisplatin with CRM197 plus 10 mmol/l of LY294002 or 1 mmol/l of wortmannin		Anti-tumor effect of the combination of cisplatin with CRM197 plus LY294002 or wortmannin		
		Co-administration of cisplatin with CRM197 plus either LY294002 or wortmannin, Akt activation significantly blocked and cell death significantly induced compared to the co-administration of cisplatin with CRM197.		
Oral cancer	CRM197 at various concentrations (0, 0.5, 1.0, 1.5, and 2.0 µg/ml)	HSC3 and SAS cells	Effect of CRM197 on HSC3 cell growth	Dateoka 2012
			Cell growth at a concentration higher than 0.5 µg/ml of CRM197 significantly was inhibited.	

Oral cancer	2.0 µg/ml of CRM197 for 48 h	HSC3 cell vs. untreated cells	The effect of CRM197 on expression of HB-EGF Decreased the HB-EGF mRNA expression was found in CRM 197 treated cells.	Dateoka 2012
	2.0 µg/ml of CRM197 for 48 h	HSC3 and SAS cells vs. Un-treatment cells	CRM197 effect on HSC3 and SAS cells invasion At presence of CRM197 partially inhibited cell invasion was reported.	
	2.0 µg/ml of CRM197	HSC3 cells	CRM 197 effect on MMP-9 and VEGF expression Significantly reduced MMP-9 expression and VEGF and MMP-9 activity was reported.	
	Groups were divided as follow; control (1), CRM197 1 mg/kg/day (2), CDDP 1 mg/kg/day (3), and CRM197 and CDDP 1 mg/kg/day (4)	Tumorigenesis by HSC3 cells implanted subcutaneously into female BALB/c nu/nu mice	CRM 197 effect on tumor formation Partially inhibition of tumor growth was seen following the treatment with either CDDP (1 mg/kg/day) or CRM197 (1 mg/kg/day) and completely inhibition was seen at presence of coadministration of 1.0 mg/kg CDDP and 1.0 mg/kg CRM197.	
Neuroblastoma	10 µg/ml of CRM197 for 48h	SK-N-SH cells	CRM197 effect on apoptosis The rate of cell apoptosis significantly increased in cells with high HB-EGF expression	Nam 2015
Triple-negative breast cancer	2 models of treatment were as follow; 1. the adjuvant therapy model CRM197 was injected intravenously at the same time as the subcutaneously injected MDA-MB-231 cells (1, 5, 10 and 50 mg/kg) 2. advanced cancer model CRM197 was injected after the tumor reached an estimated volume >100 mm ³ . (1, 5, 10 and 50 mg/kg)	Tumorigenesis by Subcutaneously injection of MDA-MB-231cells into NOD/SCID mice	Effect of CRM197 doses on tumor volume The anti-tumor effect of CRM197 was more effective in the advanced cancer therapy model compared to the adjuvant therapy model.	Nam 2016
lung cancer	1 µg/ml of CRM197	RERF-LC-A1 and NCI-H1975 cells	CRM197 effect on apoptosis Increased rate of cell death was reported.	Yotsumoto 2017
	1 mg/kg of CRM197 was administered daily for 10 consecutive days	Tumorigenesis by RERF-LC-A1 and NCI-H1975 cells injected subcutaneous into female BALB/c nu/nu mice	Anti-tumor effect of CRM197 Completely inhibition of tumor formation by RERF-LC-A1 cells, and significantly suppression by NCI-H1975 cells was demonstrated.	

Ovarian cancer (OC) or peritoneal cancer (PC)	(1.0, 2.0, 3.3, and 5.0 mg/m ²) dose of CRM 197, intraperitoneal	Patients with advanced or recurrent OC or PC	CRM 197 effect on HB-EGF	Miyamoto 2017
			Reduced HB-EGF concentration in serum and abdominal fluid was observed. 8 of 11 patients completed treatment.	
Ovarian cancer	The A2780 cells and the SKOV3 cells that were infected with AdCRM197	A2780 cells vs. SKOV3 cells	Assessment the A2780 cells and SKOV3 cells sensitivity to AdCRM197	Dai 2018
			The A2780 cells were sensitive to AdCRM197, but the SKOV3 cells were resistant to AdCRM197	
	Wild type p53 A2780 cells and knockdown p53 A2780 cells were infected with AdCRM197	wild type p53 A2780 cells vs.	Cell sensitivity investigation to AdCRM197	
			Although the A2780 cells were sensitive, the knockdown p53 A2780 cells were resistance to AdCRM197.	
	p53 gene was restored in SKOV3 cells with Adp53 and SKOV3 cells and SKOV3-p53 cells infected with AdCRM197	SKOV3 cells vs. SKOV3-p53 cells	Assessment the cell sensitivity to AdCRM197	
			Only the SKOV3-p53 cells were sensitive to AdCRM197.	
	A2780 cells and A2780 cells with p53 knocked down infected with AdCRM197	A2780 cells vs. A2780 cells with p53 knocked down	The role of p53 in AdCRM197-induced apoptosis	
			Unlike the A2780 cells with p53 knocked down gene, caspase-dependent apoptosis was activated in A2780 cells.	
A2780 cells and mutant EF2 (A2780-mEF2) cells were infected with AdCRM197.	wild type EF2 A2780 cells vs. the A2780- mutant EF2 cells	Effect of the EF2-ADP-ribosyl transferase activity of CRM197 in p53 activation		
		Unlike the A2780-mEF2 cells, apoptosis via p53 pathway was activated in wild type EF2 A2780 cells.		
A2780 cells and A2780-mEF2 cells were infected with AdCRM197.	wild type EF2 A2780 cells vs. the A2780- mutant EF2 cells	Assessment the p53 and HB-EGF gene expression		
		The higher level of p53 and HB-EGF genes expression in AdCRM197-infected A2780 cells was reported, while no significant alteration of gene expression was observed in the AdCRM197-infected A2780-mEF2 cells.		
The combined therapy of AdCRM197 plus Adp53 was used on A2780 xenograft tumors	Tumorigenesis by A2780 cells were injected into mice	Antitumor effect of the combined therapy with AdCRM197 plus Adp53		
		Reduced tumor volume and increased the cells death were reported in the mice that received combined therapy with AdCRM197 plus Adp53 in compared to that of the AdCRM197 group.		

Ovarian cancer	The combined therapy of AdCRM197 plus Adp53 was used on SKOV3 xenograft tumors	Tumorigenesis by SKOV3 were injected into mice	<p>Antitumor effect of the combined therapy with AdCRM197 plus Adp53 to overcome resistance of SKOV3 xenograft tumors to AdCRM197 in vivo</p> <p>Reduced tumor volume and increased the cells death were reported in the mice that received combined therapy with AdCRM197 plus Adp53 in compared to that of the AdCRM197 group.</p>	Dai 2018
----------------	--------------------------------------------------------------------------------	------------------------------------------------	--------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------------	----------

*Breast, Colon, Mouth, Kidney, Bladder, Ovary, Uterus, Prostate, Brain, neuroblastoma, and non-small cell lung cancer

Table 1. Anticancer effect of CRM197

of the EGFR signaling pathway and an effective drug in breast cancer patients resistant to trastuzumab (77). The evidence has indicated that activation of PI3K/Akt signaling pathway via phosphorylation of 308Thr and 473Ser in Akt is involved in glioma progression, invasion, and angiogenesis (78-80). Wang et al. showed that growth and apoptosis of human U251 glioma cells were induced after treatment with cisplatin combined with CRM197 plus LY294002 or wortmannin. Their research revealed that this activation occurred through Akt signaling pathway only in cisplatin-treated U251 cells. They suggested that Akt activation in the cells exposed to cisplatin is due to cisplatin resistance. In contrast, inhibition of PI3K/Akt signaling pathway in CRM197-treated U251 cells is done by inhibiting proteolytic cleavage of pro-HB-EGF and secretion of the soluble form of the receptor. This soluble form has mitogenic and chemoattractant properties for various cell types by activating the PI3K/Akt pathway. Moreover, they found that combining cisplatin, CRM197, and some specific PI3K inhibitors such as LY294002 or wortmannin could significantly suppress the PI3K/Akt pathway leading to inhibition of U251 cell growth and induction of apoptosis when compared to control groups (65). Dateo et al. reported that CRM197 could suppress the invasiveness and motility of cultured oral cancer cell line through inhibition of HB-EGF and reduction in MMP-9 expression, which has a crucial role in VEGF produced by endothelial cells (81). The pivotal role of HB-EGF in neuroblastoma, triple-negative breast cancer, and OC was reported by Nam et al. These researchers introduced CRM197 as a safe and promising anticancer drug that targets HB-EGF (82-84). Yotsumoto

reported that T790M mutation in exons 18-24 of EGFR gene, responsible for its tyrosine kinase activity, leads to induced resistance to pretreatment with small-molecule EGFR tyrosine kinase inhibitors (TKIs), stimulation of EGFR signaling, and increased expression of EGFR ligands such as HB-EGF in patients with lung cancer. So, the administration of CRM197 may be the best therapeutic approach in these patients (85). To explain the role of CRM197 in apoptosis through the P53 pathway, Dai et al. recruited two types of cells, A2780 cells with p53 wild-type gene, and p53 deleted SKOV3 cells, and then treated them with adenovirus-mediated CRM197 (AdCRM197). In addition, P53 was knocked-down in A2780 cells by lentivirus-mediated RNA interference, restored in SKOV3 cells with Adp53, and treated with AdCRM197 (86). Unlike the SKOV3 and A2780 cells with a deletion in gene encoding p53, caspase-dependent apoptosis was activated in A2780 and SKOV3 cells with the wild-type p53 gene. Although for a long time, until 2018, CRM197 had been considered a nontoxic variant of the diphtheria toxin, Dai et al. approved the crucial effect of weak EF2-ADP-ribosyl activity of CRM197 in the induction of apoptosis in OC cells through activation of the p53 pathway. With this aim, they treated A2780 cells with wild-type EF2 gene and A2780- mutant EF2 (A2780-mEF2) cells by AdCRM197. They demonstrated that cell death occurs only in A2780 cells. A2780-mEF2 cells are resistant to AdCRM197 and apoptosis. Thus, they hypothesized that the weak toxicity of CRM197 has led to induction of the ADP-ribosylation of EF2 and cell death in A2780 cells with wild-type EF2 gene. In addition, their results showed that p53 and HB-EGF expres-

sion were increased in AdCRM197 infected A2780 cells. But the expression of p53 and HB-EGF genes were inhibited in mutant EF2 A2780 cells, and no response was observed following treatment with AdCRM197.

DISCUSSION

Cancer has been considered one of the most significant causes of death around the world (87). Based on a report published in 2018, over half of the global cancer mortality has occurred among the Asian population (88). The survival rate of cancer patients remains poor despite impressive progress in cancer screening, diagnosis, and treatment (89, 90). More research must be conducted to investigate the problems related to cancer and its treatment. This review has focused on the role of CRM197 in the treatment of different types of cancers. Most of the studies demonstrated that treatment with CRM197 has a promising positive effect on inhibition of growth, angiogenesis, and metastasis and induction of apoptosis in

various cancer types, such as oral, ovary, lung, prostate, breast, gastric, pancreatic, adrenocortical, and colon cancer, neuroblastoma, glioblastoma, acute lymphoblastic leukemia (ALL), leukemia, melanoma and solid tumors (Fig 5) (58, 62, 75, 76, 81, 83-85, 91-103).

Although overexpression of EGFR in different types of cancers (104-106) has attracted the increasing interest of researchers as a therapeutic target, the drug resistance leading to poor outcome and severe complications during therapy is currently a problem (107). It has been shown that the coadministration of CRM197 with chemotherapeutic agents has a significantly stronger antitumor effect when compared to each of the agents separately (86, 108). In a clinical trial study performed by Miyamoto et al., the antitumor effect of CRM197 via blocking HB-EGF in eleven Japanese patients with advanced or recurrent ovarian cancer (OC) or peritoneal cancer (PC) was approved (109). In addition, Miyamoto et al. compared the side effects of intraperitoneal injection of CRM197

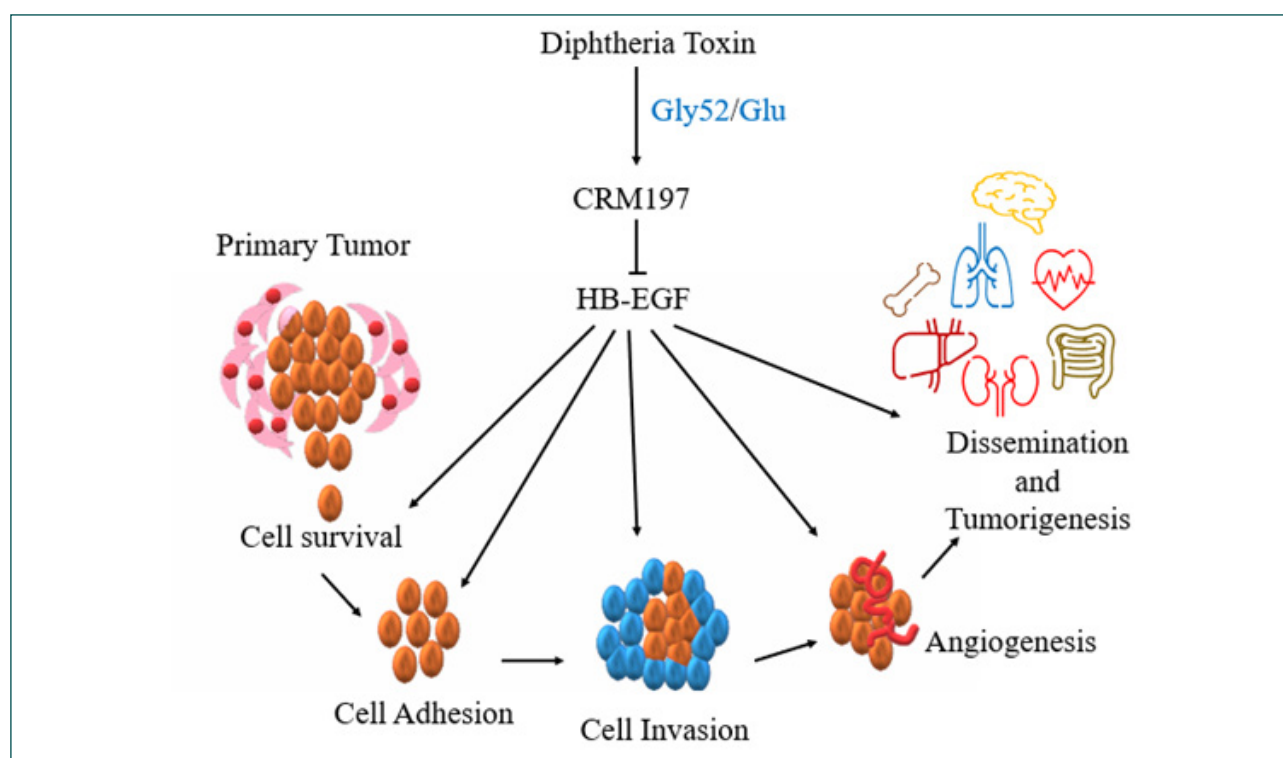


Figure.5. Mechanism of anticancer effect of CRM197; The G/A transition in the fragment A diphtheria toxin leads to Gly52/Glu substitution and CRM197 production. CRM197 can inhibit the HB-EGF-mediated signaling pathway, which leads to cell growth, invasion, proliferation, differentiation, survival, angiogenesis, and metastasis

with cisplatin. They observed that following 60–100 mg/m² intraperitoneal (IP) injection of cisplatin, the plasma concentration of the drug was approximately 0.7–2.3 µg/mL. In contrast, after IP injection of 1–3.3 mg/m² of CRM197, the plasma concentration of this drug was undetectable. They suggested that the high molecular weight of CRM197 prevents its passing across the peritoneum, and due to the high level of HB-EGF in the peritoneum, CRM197 easily interacts with it and enters into the lymphatic capillaries (109). Another clinical trial study showed that administration of CRM197 via the subcutaneous route is safe and relatively well tolerated. Other side effects of DT/CRM197, including skin irritation in the injection sites and a flu-like syndrome with fever, were observed only in patients with hypersensitivity (66). In addition, CRM197 is one of the best carriers for stimulating the immune response in cancer immunotherapy. To assess the immune response, Globo H (GH), a hexasaccharide overexpressed in a variety of cancer cells, was attached to different carriers such as keyhole limpet hemocyanin, CRM197, tetanus toxoid, and BSA combined with an adjuvant and were injected into mice. Huang et al. reported that combination of GH-CRM197 adjuvant with α -galactosylceramide C34 induces the highest production of antibodies with IgG-dominant. These antibodies act very specific and selective against cancer cells that overexpressed GH and the GH-related epitopes such as breast cancer cells (110). Interestingly, Dai et al. revealed that the weak toxicity of CRM197 and ADP-ribosylation of EF2 caused p53 pathway activation, but in the A2780 cells carrying the EF2 mutant gene, p53 expression reduced, and the cells were resistant to Ad-CRM197 significantly (86).

It has been reported that the overexpression of HB-EGF and membrane-type matrix metalloproteinase-1 (MT1-MMP) is correlated with different types of malignancies (111, 112). MT1-MMP expression leads to extracellular matrix degradation and facilitates tumor cell invasion and growth (113). Also, Naohiko et al. demonstrated that concurrent overexpression of MT1-MMP and HB-EGF in gastric cancer cells resulted in HB-EGF cleavage and enhanced cell proliferation (57). The increased production

of soluble HB-EGF is associated with MT1-MMP processing found in malignant ascites in patients with metastatic ovarian carcinoma (114). Therefore, the administration of CRM197 with MT1-MMP inhibitors might provide an effective treatment for metastatic carcinoma (57).

Although many significant advances have taken place in this respect, many vital questions remain unanswered. While most studies have introduced CRM197 as a safe antitumor drug, further clinical trial studies should be conducted to confirm that long-term treatment with this agent has no serious adverse consequences. This review reveals that CRM197 induced cell apoptosis through the activation of the p53 pathway. However, to the best of our knowledge, there has been no study on the effects of CRM197 on the expression of other tumor suppressor genes or the functions of other proteins.

CONCLUSION

This review indicates that CRM197 binding to HB-EGF and soluble HB-EGF inhibits the mitogen-activated protein kinase (MAPK) and protein kinase B (Akt) signaling pathways and suppresses cell proliferation, survival, and tumorigenicity. CRM197 inhibits angiogenesis, invasion, and metastasis through silencing the genes of protein tyrosine kinase 2 (PTK2), vascular endothelial growth factor (VEGF), and interleukin 8 (IL-8). The toxicity of CRM197 depends on activating the p53 pathway in cancer cells carrying a wild-type elongation factor 2 (EF2) gene. The combination of chemotherapeutic drugs and CRM197 enhances the effectiveness of cancer treatment. CRM197 as an ideal carrier protein induces the highest production of antibodies with specific and selective functions against cancer cells.

The weak or lack of enzymatic activity of CRM197 compared with diphtheria toxin decreases the survival of cancer cells via overexpression of HB-EGF, inhibiting HB-EGF's interaction with EGFR, or by residual enzymatic activity in cells transfected with CRM197. Regarding the positive anticancer effect of CRM197 on different types of cancer cells, appropriate clinical trial studies are required to investigate the efficacy of its combination with other therapies such as surgery and radiotherapy.

Further studies in gene therapy aimed at restoring the defective tumor suppressor genes combined with targeted drug delivery methods such as injecting virus-mediated CRM197 to human cancer cells are needed to confirm the precise anticancer effect of CRM197 and assess the life-threatening side effects.

Conflict of Interest Statement: The authors have declared that no competing interests exist.

Role of the funding source: The authors received no specific funding for this work.

Ethical approval: Not applicable

Informed consent: Not applicable.

Abbreviations:

HB-EGF: Heparin-binding epidermal growth factor-like growth factor

CRM197 cross-reacting materials 197

EGFR: epidermal growth factor receptor

DtxR: diphtheria toxin repressor

DT: diphtheria toxin

EF2: elongation factor 2

Cys: Cysteine

Arf-1: ADP-ribosylation factor 1

COPI: coat protein complex I

Rab-5: ras-related protein

V-ATPase: vacuolar H⁺-ATPase

CTF: cytosolic translocation factor

TrxRs: thioredoxin reductases

PDI: protein disulfide isomerase

ADPR: adenosine diphosphate ribosyl

Gly: Glycine

Glu: Glutamic acid

CTF: cytosolic translocation factor

TrxRs: thioredoxin reductases

PDI: protein disulfide isomerase

ADPR: adenosine diphosphate ribosyl

Gly: Glycine

Glu: Glutamic acid

ADAM: a disintegrin and metalloproteinases

MMP: matrix metalloproteinases

Lys: lysine

His: histidine

DRAP27: diphtheria toxin receptor-associated protein

CD9: cluster of differentiation 9

PTKR: protein tyrosine kinase receptor

RTK: receptors of tyrosine kinase

JM: juxtamembrane

TGFA: transforming growth factor- α

AREG: amphiregulin

EPGN: epigen

BTC: betacellulin

EPR: epiregulin

aa: amino acid

MAPK: mitogen-activated protein kinase

PI3K/Akt: phosphoinositide-3-kinase/protein kinase B

sHB-EGF: soluble Heparin-binding epidermal growth factor-like growth factor

RTK: receptor tyrosine kinase

TNF- α : tumor necrosis factor alpha

JNK: Jun N-terminal kinase

HB-EGF-CTFs: carboxyl-terminal fragments of HB-EGF

VEGF: vascular endothelial growth factor

PTK2: protein tyrosine kinase 2

FAK: focal adhesion kinase

IL- 8: interleukin 8

5-FU: 5-Fluorouracil

Thr: Threonine

Ser: Serine

TKIs: tyrosine kinase inhibitors

mEF2: mutant EF2

ALL: acute lymphoblastic leukemia

OC: ovarian cancer

PC: peritoneal cancer

ip: intraperitoneal

GH: Globo H

BSA: bovine serum albumin

SSEA3: stage-specific embryonic antigen-3

MT1-MMP: membrane-type matrix metalloproteinase-1

REFERENCES

- Holmes RK. Biology and molecular epidemiology of diphtheria toxin and the tox gene. *Journal of Infectious Diseases*. 2000;181(Supplement_1):S156-S67.
- Trost E, Blom J, Soares Sde C, Huang IH, Al-Dilaimi A, Schroder J, et al. Pangenomic study of *Corynebacterium diphtheriae* that provides insights into the genomic diversity of pathogenic isolates from cases of classical diphtheria, endocarditis, and pneumonia. *J Bacteriol*. 2012;194(12):3199-215.
- Qian Y, Lee JH, Holmes RK. Identification of a DtxR-regulated operon that is essential for siderophore-dependent iron uptake in *Corynebacterium diphtheriae*. *Journal of bacteriology*. 2002;184(17):4846-56.
- Guedon E, Helmann JD. Origins of metal ion selectivity in the DtxR/MntR family of metalloregulators. *Molecular microbiology*. 2003;48(2):495-506.
- Collier R. Understanding the mode of action of diphtheria toxin: a perspective on progress during the 20th century. *Toxicon*. 2001;39(11):1793-803.
- Kaczorek M, Delpeyroux F, Chenciner N, Streeck RE, Boquet P, Tiollais P. Nucleotide sequence and expression of the diphtheria tox228 gene in *Escherichia coli*. *Science*. 1983;221(4613):855-8.
- Smith W, Tai P, Murphy J, Davis B. Precursor in cotranslational secretion of diphtheria toxin. *Journal of bacteriology*. 1980;141(1):184-9.
- Greenfield L, Bjorn MJ, Horn G, Fong D, Buck GA, Collier RJ, et al. Nucleotide sequence of the structural gene for diphtheria toxin carried by corynebacteriophage beta. *Proceedings of the National Academy of Sciences*. 1983;80(22):6853-7.
- Zhang J, Wei H, Guo X, Hu M, Gao F, Li L, et al. Functional verification of the diphtheria toxin A gene in a recombinant system. *J Anim Sci Biotechnol*. 2012;3(1):29.
- Shafiee F, Aucoin MG, Jahanian Najafabadi A. Targeted Diphtheria Toxin Based Therapy: A Review Article. *Frontiers in microbiology*. 2019;10:2340.
- Chenal A, Nizard P, Gillet D. Structure and function of diphtheria toxin: from pathology to engineering. *Journal of Toxicology: Toxin Reviews*. 2002;21(4):321-59.
- h B. uptake and trafficking of protein toxins book. K A, editor: springer international publishing 2017.
- Murphy JR. Mechanism of diphtheria toxin catalytic domain delivery to the eukaryotic cell cytosol and the cellular factors that directly participate in the process. *Toxins (Basel)*. 2011;3(3):294-308.
- Louie GV, Yang W, Bowman ME, Choe S. Crystal structure of the complex of diphtheria toxin with an extracellular fragment of its receptor. *Molecular cell*. 1997;1(1):67-78.
- Edwards JP, Zhang X, Mosser DM. The expression of heparin-binding epidermal growth factor-like growth factor by regulatory macrophages. *J Immunol*. 2009;182(4):1929-39.
- Freeman MR, Yoo JJ, Raab G, Soker S, Adam RM, Schneck FX, et al. Heparin-binding EGF-like growth factor is an autocrine growth factor for human urothelial cells and is synthesized by epithelial and smooth muscle cells in the human bladder. *The Journal of clinical investigation*. 1997;99(5):1028-36.
- Thorne BA, Plowman GD. The heparin-binding domain of amphiregulin necessitates the precursor pro-region for growth factor secretion. *Molecular and cellular biology*. 1994;14(3):1635-46.
- Lanzrein M, Garred O, Olsnes S, Sandvig K. Diphtheria toxin endocytosis and membrane translocation are dependent on the intact membrane-anchored receptor (HB-EGF precursor): studies on the cell-associated receptor cleaved by a metalloprotease in phorbol-ester-treated cells. *Biochem J*. 1995;310 (Pt 1):285-9.
- Chiron MF, Fryling CM, FitzGerald DJ. Cleavage of pseudomonas exotoxin and diphtheria toxin by a furin-like enzyme prepared from beef liver. *J Biol Chem*. 1994;269(27):18167-76.
- Lemichez E, Bomsel M, Devilliers G, van der Spek J, Murphy JR, Lukianov EV, et al. Membrane translocation of diphtheria toxin fragment A exploits early to late endosome trafficking machinery. *Molecular microbiology*. 1997;23(3):445-57.
- Watson P, Spooner RA. Toxin entry and trafficking in mammalian cells. *Adv Drug Deliv Rev*. 2006;58(15):1581-96.
- Papini E, Rappuoli R, Murgia M, Montecucco C. Cell penetration of diphtheria toxin. Reduction of the interchain disulfide bridge is the rate-limiting step of translocation in the cytosol. *Journal of Biological Chemistry*. 1993;268(3):1567-74.
- Simon NC, Aktories K, Barbieri JT. Novel bacterial ADP-ribosylating toxins: structure and function. *Nat Rev Microbiol*. 2014;12(9):599-611.
- Collier RJ. Diphtheria toxin: mode of action and structure. *Bacteriological reviews*. 1975;39(1):54.
- Giannini G, Rappuoli R, Ratti G. The amino-acid sequence of two non-toxic mutants of diphtheria toxin: CRM45 and CRM197. *Nucleic Acids Res*. 1984;12(10):4063-9.
- Uchida T, Pappenheimer AM, Greany R. Diphtheria toxin and related proteins I. Isolation and properties of mutant proteins serologically related to diphtheria toxin. *Journal of Biological Chemistry*. 1973;248(11):3838-44.
- Brooke JS, Cha J-H, Eidels L. Diphtheria toxin: receptor interaction: association, dissociation, and effect of pH. *Biochemical and biophysical research communications*. 1998;248(2):297-302.
- Broker M, Costantino P, DeTora L, McIntosh ED, Rappuoli R. Biochemical and biological characteristics of cross-reacting material 197 CRM197, a non-toxic mutant of diphtheria toxin: use as a conjugation protein in vaccines and other potential clinical applications. *Biologicals*. 2011;39(4):195-204.
- Mishra RPN, Yadav RSP, Jones C, Nocadello S, Minasov G, Shuvalova LA, et al. Structural and immunological characterization of *E. coli* derived recombinant CRM197 protein used as carrier in conjugate vaccines. *Biosci Rep*. 2018;38(5).
- PAPINI E, SCHIAVO G, TOMASI M, COLOMBATTI M, RAPPULI R, MONTECUCCO C. Lipid interaction of diphtheria toxin and mutants with altered fragment B: 2. Hydrophobic photolabelling and cell intoxication. *European journal of biochemistry*. 1987;169(3):637-44.
- Sandvig K, Olsnes S. Diphtheria toxin entry into cells is facilitated by low pH. *The Journal of cell biology*. 1980;87(3):828-32.
- DEMEL R, SCHIAVO G, KRUIJFF Bd, MONTECUCCO C. Lipid interaction of diphtheria toxin and mutants: a study with phospholipid and protein monolayers. *European journal of biochemistry*. 1991;197(2):481-6.
- Hu VW, Holmes RK. Single mutation in the A domain of diphtheria toxin results in a protein with altered membrane insertion behavior. *Biochimica et Biophysica Acta (BBA)-Biomembranes*. 1987;902(1):24-30.
- Kent MS, Yim H, Murton JK, Satija S, Majewski J, Kuzmen-

- ko I. Oligomerization of membrane-bound diphtheria toxin (CRM197) facilitates a transition to the open form and deep insertion. *Biophys J*. 2008;94(6):2115-27.
- 35 Mitamura T, Higashiyama S, Taniguchi N, Klagsbrun M, Mekada E. Diphtheria toxin binds to the epidermal growth factor (EGF)-like domain of human heparin-binding EGF-like growth factor/diphtheria toxin receptor and inhibits specifically its mitogenic activity. *Journal of Biological Chemistry*. 1995;270(3):1015-9.
- 36 Gregory H. Isolation and structure of urogastrone and its relationship to epidermal growth factor. *Nature*. 1975;257(5524):325-7.
- 37 Iwamoto R, Takagi M, Akatsuka J, Ono K, Kishi Y, Mekada E. Characterization of a Novel Anti-Human HB-EGF Monoclonal Antibody Applicable for Paraffin-Embedded Tissues and Diagnosis of HB-EGF-Related Cancers. *Monoclon Antib Immunodiagn Immunother*. 2016;35(2):73-82.
- 38 Zhao G, Liu L, Peek RM, Jr., Hao X, Polk DB, Li H, et al. Activation of Epidermal Growth Factor Receptor in Macrophages Mediates Feedback Inhibition of M2 Polarization and Gastrointestinal Tumor Cell Growth. *J Biol Chem*. 2016;291(39):20462-72.
- 39 Negahdari B, Shahosseini Z, Baniyasi V. Production of human epidermal growth factor using adenoviral based system. *Res Pharm Sci*. 2016;11(1):43-8.
- 40 Miyazono K. Ectodomain shedding of HB-EGF: a potential target for cancer therapy. *J Biochem*. 2012;151(1):1-3.
- 41 Kim GY, Besner GE, Steffen CL, McCarthy DW, Downing MT, Luquette MH, et al. Purification of heparin-binding epidermal growth factor-like growth factor from pig uterine luminal flushings, and its production by endometrial tissues. *Biology of reproduction*. 1995;52(3):561-71.
- 42 Higashiyama S, Lau K, Besner G, Abraham JA, Klagsbrun M. Structure of heparin-binding EGF-like growth factor. Multiple forms, primary structure, and glycosylation of the mature protein. *Journal of Biological Chemistry*. 1992;267(9):6205-12.
- 43 Uchiyama-Tanaka Y, Matsubara H, Mori Y, Kosaki A, Kishimoto N, Amano K, et al. Involvement of HB-EGF and EGF receptor transactivation in TGF- β -mediated fibronectin expression in mesangial cells. *Kidney international*. 2002;62(3):799-808.
- 44 Vinante F, Rigo A. Heparin-binding epidermal growth factor-like growth factor/diphtheria toxin receptor in normal and neoplastic hematopoiesis. *Toxins*. 2013;5(6):1180-201.
- 45 Eidels L, Ross LL, Hart DA. Diphtheria toxin-receptor interaction: a polyphosphate-insensitive diphtheria toxin-binding domain. *Biochemical and biophysical research communications*. 1982;109(2):493-9.
- 46 Brooke JS, Cha J-H. Molecular characterization of key diphtheria toxin: receptor interactions. *Biochemical and biophysical research communications*. 2000;275(2):374-81.
- 47 Wieduwilt MJ, Moasser MM. The epidermal growth factor receptor family: biology driving targeted therapeutics. *Cell Mol Life Sci*. 2008;65(10):1566-84.
- 48 Wang Q, Chen X, Wang Z. Dimerization drives EGFR endocytosis through two sets of compatible endocytic codes. *J Cell Sci*. 2015;128(5):935-50.
- 49 Bodnar RJ. Epidermal Growth Factor and Epidermal Growth Factor Receptor: The Yin and Yang in the Treatment of Cutaneous Wounds and Cancer. *Adv Wound Care (New Rochelle)*. 2013;2(1):24-9.
- 50 Purba ER, Saita EI, Maruyama IN. Activation of the EGF Receptor by Ligand Binding and Oncogenic Mutations: The "Rotation Model". *Cells*. 2017;6(2).
- 51 Rayego-Mateos S, Rodrigues-Diez R, Morgado-Pascual JL, Valentijn F, Valdivielso JM, Goldschmeding R, et al. Role of Epidermal Growth Factor Receptor (EGFR) and Its Ligands in Kidney Inflammation and Damage. *Mediators Inflamm*. 2018;2018:8739473.
- 52 Roskoski R, Jr. Small molecule inhibitors targeting the EGFR/ ErbB family of protein-tyrosine kinases in human cancers. *Pharmacol Res*. 2019;139:395-411.
- 53 Walsh AM, Lazzara MJ. Regulation of EGFR trafficking and cell signaling by Sprouty2 and MIG6 in lung cancer cells. *J Cell Sci*. 2013;126(Pt 19):4339-48.
- 54 Miyamoto S, Yagi H, Yotsumoto F, Horiuchi S, Yoshizato T, Kawarabayashi T, et al. New approach to cancer therapy: heparin binding-epidermal growth factor-like growth factor as a novel targeting molecule. *Anticancer Res*. 2007;27(6A):3713-21.
- 55 Zandi R, Larsen AB, Andersen P, Stockhausen MT, Poulsen HS. Mechanisms for oncogenic activation of the epidermal growth factor receptor. *Cell Signal*. 2007;19(10):2013-23.
- 56 Normanno N, De Luca A, Bianco C, Strizzi L, Mancino M, Maiello MR, et al. Epidermal growth factor receptor (EGFR) signaling in cancer. *Gene*. 2006;366(1):2-16.
- 57 Koshikawa N, Mizushima H, Minegishi T, Eguchi F, Yotsumoto F, Nabeshima K, et al. Proteolytic activation of heparin-binding EGF-like growth factor by membrane-type matrix metalloproteinase-1 in ovarian carcinoma cells. *Cancer Sci*. 2011;102(1):111-6.
- 58 Amedei A, Asadzadeh F, Papi F, Vannucchi MG, Ferrucci V, Bermejo IA, et al. A Structurally Simple Vaccine Candidate Reduces Progression and Dissemination of Triple Negative Breast Cancer. *iScience*. 2020;101250.
- 59 Song C, Zheng X-J, Guo H, Cao Y, Zhang F, Li Q, et al. Fluorine-modified sialyl-Tn-CRM197 vaccine elicits a robust immune response. *Glycoconjugate journal*. 2019;36(5):399-408.
- 60 Mawas F, Niggemann J, Jones C, Corbel MJ, Kamerling JP, Vliegenthart JF. Immunogenicity in a mouse model of a conjugate vaccine made with a synthetic single repeating unit of type 14 pneumococcal polysaccharide coupled to CRM197. *Infection and immunity*. 2002;70(9):5107-14.
- 61 Zhang H-L, Yuan C, Zhang D-M, Shi H-S, Li M, Luo Z-C, et al. A novel combined conjugate vaccine: enhanced immunogenicity of bFGF with CRM197 as a carrier protein. *Molecular medicine reports*. 2011;4(5):857-63.
- 62 Huiyong Z, Yong L, Didier M, Yu Z, Jing F, Rongyue C, et al. Enhanced inhibition of murine prostatic carcinoma growth by immunization with or administration of viable human umbilical vein endothelial cells and CRM197. *Brazilian Journal of Medical and Biological Research*. 2011;44(2):140-8.
- 63 Jaffe J, Wucherer K, Sperry J, Zou Q, Chang Q, Massa MA, et al. Effects of conformational changes in peptide-CRM197 conjugate vaccines. *Bioconjugate chemistry*. 2018;30(1):47-53.
- 64 Amedei A, Asadzadeh F, Papi F, Vannucchi MG, Ferrucci V, Bermejo IA, et al. A Structurally Simple Vaccine Candidate Reduces Progression and Dissemination of Triple-Negative Breast Cancer. *Isience*. 2020;23(6):101250.
- 65 Wang L, Wang P, Liu Y, Xue Y. Regulation of cellular growth, apoptosis, and Akt activity in human U251 glioma cells by a combination of cisplatin with CRM197. *Anti-cancer drugs*. 2012;23(1):81-9.
- 66 Buzzi S, Rubboli D, Buzzi G, Buzzi AM, Morisi C, Pironi F. CRM197 (nontoxic diphtheria toxin): effects on advanced

- cancer patients. *Cancer Immunology, Immunotherapy*. 2004;53(11):1041-8.
- 67 Yagi H, Yotsumoto F, Sonoda K, Kuroki M, Mekada E, Miyamoto S. Synergistic anti-tumor effect of paclitaxel with CRM197, an inhibitor of HB-EGF, in ovarian cancer. *International journal of cancer*. 2009;124(6):1429-39.
 - 68 Barbolina MV. Molecular Mechanisms Regulating Organ-Specific Metastases in Epithelial Ovarian Carcinoma. *Cancers (Basel)*. 2018;10(11).
 - 69 Mitra AK. *Ovarian Cancer Metastasis: A Unique Mechanism of Dissemination*. Tumor Metastasis: IntechOpen; 2016.
 - 70 Slack-Davis JK, Atkins KA, Harrer C, Hershey ED, Conaway M. Vascular cell adhesion molecule-1 is a regulator of ovarian cancer peritoneal metastasis. *Cancer research*. 2009;69(4):1469-76.
 - 71 Desgrosellier JS, Cheresch DA. Integrins in cancer: biological implications and therapeutic opportunities. *Nat Rev Cancer*. 2010;10(1):9-22.
 - 72 Yagi H, Yotsumoto F, Miyamoto S. Heparin-binding epidermal growth factor-like growth factor promotes transcoelomic metastasis in ovarian cancer through epithelial-mesenchymal transition. *Molecular cancer therapeutics*. 2008;7(10):3441-51.
 - 73 Yotsumoto F, Sanui A, Fukami T, Shirota K, Horiuchi S, Tsujioka H, et al. Efficacy of ligand-based targeting for the EGF system in cancer. *Anticancer Res*. 2009;29(11):4879-85.
 - 74 Yagi H, Yotsumoto F, Sonoda K, Kuroki M, Mekada E, Miyamoto S. Synergistic anti-tumor effect of paclitaxel with CRM197, an inhibitor of HB-EGF, in ovarian cancer. *International journal of cancer*. 2009;124(6):1429-39.
 - 75 Martarelli D, Pompei P, Mazzoni G. Inhibition of adrenocortical carcinoma by diphtheria toxin mutant CRM197. *Chemotherapy*. 2009;55(6):425-32.
 - 76 Sanui A, Yotsumoto F, Tsujioka H, Fukami T, Horiuchi S, Shirota K, et al. HB-EGF inhibition in combination with various anticancer agents enhances its antitumor effects in gastric cancer. *Anticancer research*. 2010;30(8):3143-9.
 - 77 Yotsumoto F, Oki E, Tokunaga E, Maehara Y, Kuroki M, Miyamoto S. HB-EGF orchestrates the complex signals involved in triple-negative and trastuzumab-resistant breast cancer. *International journal of cancer*. 2010;127(11):2707-17.
 - 78 Pu P, Kang C, Zhang Z, Liu X, Jiang H. Downregulation of PIK3CB by siRNA suppresses malignant glioma cell growth in vitro and in vivo. *Technology in cancer research & treatment*. 2006;5(3):271-80.
 - 79 Pu P, Kang C, Li J, Jiang H. Antisense and dominant-negative AKT2 cDNA inhibits glioma cell invasion. *Tumor Biology*. 2004;25(4):172-8.
 - 80 Pu P, Kang C, Li J, Jiang H, Cheng J. The effects of antisense AKT2 RNA on the inhibition of malignant glioma cell growth in vitro and in vivo. *Journal of neuro-oncology*. 2006;76(1):1.
 - 81 Dateoka S, Ohnishi Y, Kakudo K. Effects of CRM197, a specific inhibitor of HB-EGF, in oral cancer. *Medical molecular morphology*. 2012;45(2):91-7.
 - 82 Tanaka Y, Miyamoto S, Suzuki SO, Oki E, Yagi H, Sonoda K, et al. Clinical significance of heparin-binding epidermal growth factor-like growth factor and a disintegrin and metalloprotease 17 expression in human ovarian cancer. *Clinical Cancer Research*. 2005;11(13):4783-92.
 - 83 Nam SO, Yotsumoto F, Miyata K, Souzaki R, Taguchi T, Kuroki M, et al. Validity of HB-EGF as target for human neuroblastoma therapy. *Anticancer research*. 2015;35(8):4433-40.
 - 84 Nam SO, Yotsumoto F, Miyata K, Fukagawa S, Odawara T, Manabe S, et al. Anti-tumor effect of intravenous administration of CRM197 for triple-negative breast cancer therapy. *Anticancer research*. 2016;36(7):3651-7.
 - 85 Yotsumoto F, Fukagawa S, Miyata K, Nam SO, Katsuda T, Miyahara D, et al. HB-EGF Is a Promising Therapeutic Target for Lung Cancer with Secondary Mutation of EGFRT790M. *Anticancer research*. 2017;37(7):3825-31.
 - 86 Dai L, Pan Q, Peng Y, Huang S, Liu J, Chen T, et al. p53 Plays a Key Role in the Apoptosis of Human Ovarian Cancer Cells Induced by Adenovirus-Mediated CRM197. *Hum Gene Ther*. 2018;29(8):916-26.
 - 87 Zaorsky NG, Churilla TM, Egleston BL, Fisher SG, Ridge JA, Horwitz EM, et al. Causes of death among cancer patients. *Ann Oncol*. 2017;28(2):400-7.
 - 88 Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin*. 2018;68(6):394-424.
 - 89 Escaff S, Fernandez J, Gonzalez L, Suárez A, Gonzalez-Reyes S, Gonzalez J, et al. Study of matrix metalloproteinases and their inhibitors in prostate cancer. *British journal of cancer*. 2010;102(5):922-9.
 - 90 Society AC. *Cancer Treatment & Survivorship Facts & Figures 2019–2021*. American Cancer Society Atlanta, GA; 2019.
 - 91 Tang XH, Deng S, Li M, Lu MS. Cross-reacting material 197 reverses the resistance to paclitaxel in paclitaxel-resistant human ovarian cancer. *Tumour Biol*. 2016;37(4):5521-8.
 - 92 Tang X, Lu M, Li C, Deng S, Li M. [Expression and significance of heparin binding-epidermal growth factor-like growth factor in paclitaxel-resistant ovarian cancer]. *Zhonghua Fu Chan Ke Za Zhi*. 2014;49(7):517-22.
 - 93 Hu Y, Lin X, Wang P, Xue YX, Li Z, Liu LB, et al. CRM197 in Combination With shRNA Interference of VCAM-1 Displays Enhanced Inhibitory Effects on Human Glioblastoma Cells. *J Cell Physiol*. 2015;230(8):1713-28.
 - 94 Fiorentini G, Banfi R, Dentico P, Moriconi S, Turrisi G, Pelagotti F, et al. Clinical experience of treatment of metastatic melanoma and solid tumours adopting a derivative of diphtheria toxin: cross-reacting material 197. *In Vivo*. 2013;27(2):197-202.
 - 95 Rivetti S, Lauriola M, Voltattorni M, Bianchini M, Martini D, Ceccarelli C, et al. Gene expression profile of human colon cancer cells treated with cross-reacting material 197, a diphtheria toxin non-toxic mutant. *Int J Immunopathol Pharmacol*. 2011;24(3):639-49.
 - 96 Kunami N, Yotsumoto F, Ishitsuka K, Fukami T, Odawara T, Manabe S, et al. Antitumor effects of CRM197, a specific inhibitor of HB-EGF, in T-cell acute lymphoblastic leukemia. *Anticancer Res*. 2011;31(7):2483-8.
 - 97 Cheng L-m, Jiang J-g, Sun Z-y, Chen C, Dackor RT, Zeldin DC, et al. The epoxyeicosatrienoic acid-stimulated phosphorylation of EGF-R involves the activation of metalloproteinases and the release of HB-EGF in cancer cells. *Acta pharmacologica Sinica*. 2010;31(2):211-8.
 - 98 Tsujioka H, Yotsumoto F, Shirota K, Horiuchi S, Yoshizato T, Kuroki M, et al. Emerging strategies for ErbB ligand-based targeted therapy for cancer. *Anticancer research*. 2010;30(8):3107-12.
 - 99 Tang Xh, Li H, Zheng Xs, Lu Ms, An Y, Zhang Xl. CRM197 reverses paclitaxel resistance by inhibiting the NAC-1/Gadd45 pathway in paclitaxel-resistant ovarian cancer cells. *Cancer medicine*. 2019;8(14):6426-36.
 - 100 Fogar P, Navaglia F, Basso D, Zambon C, Moserle L, Indraccolo S, et al. Heat-induced transcription of diphtheria toxin A or its var-

- iants, CRM176 and CRM197: implications for pancreatic cancer gene therapy. *Cancer Gene Therapy*. 2010;17(1):58-68.
- 101 Tsujioka H, Yotsumoto F, Hikita S, Ueda T, Kuroki M, Miyamoto S. Targeting the heparin-binding epidermal growth factor-like growth factor in ovarian cancer therapy. *Current Opinion in Obstetrics and Gynecology*. 2011;23(1):24-30.
- 102 Cheng K, Xie G, Raufman J-P. Matrix metalloproteinase-7-catalyzed release of HB-EGF mediates deoxycholytaurine-induced proliferation of a human colon cancer cell line. *Biochemical pharmacology*. 2007;73(7):1001-12.
- 103 Weerakkody LR, Witharana C. The role of bacterial toxins and spores in cancer therapy. *Life sciences*. 2019;235:116839.
- 104 Agarwal V, Subash A, Nayar RC, Rao V. Is EGFR really a therapeutic target in head and neck cancers? *J Surg Oncol*. 2019.
- 105 Anderson NG, Ahmad T, Chan K, Dobson R, Bundred NJ. ZD1839 (Iressa), a novel epidermal growth factor receptor (EGFR) tyrosine kinase inhibitor, potently inhibits the growth of EGFR-positive cancer cell lines with or without erbB2 overexpression. *International journal of cancer*. 2001;94(6):774-82.
- 106 Sigismund S, Avanzato D, Lanzetti L. Emerging functions of the EGFR in cancer. *Mol Oncol*. 2018;12(1):3-20.
- 107 Tang XH, Deng S, Li M, Lu MS. The anti-tumor effect of cross-reacting material 197, an inhibitor of heparin-binding EGF-like growth factor, in human resistant ovarian cancer. *Biochem Biophys Res Commun*. 2012;422(4):676-80.
- 108 Miyata K, Yotsumoto F, Nam SO, Odawara T, Manabe S, Ishikawa T, et al. Contribution of transcription factor, SP1, to the promotion of HB-EGF expression in defense mechanism against the treatment of irinotecan in ovarian clear cell carcinoma. *Cancer Med*. 2014;3(5):1159-69.
- 109 Miyamoto S, Yotsumoto F, Ueda T, Fukami T, Sanui A, Miyata K, et al. BK-UM in patients with recurrent ovarian cancer or peritoneal cancer: a first-in-human phase-I study. *BMC Cancer*. 2017;17(1):89.
- 110 Huang YL, Hung JT, Cheung SK, Lee HY, Chu KC, Li ST, et al. Carbohydrate-based vaccines with a glycolipid adjuvant for breast cancer. *Proc Natl Acad Sci U S A*. 2013;110(7):2517-22.
- 111 Seiki M. Membrane-type 1 matrix metalloproteinase: a key enzyme for tumor invasion. *Cancer letters*. 2003;194(1):1-11.
- 112 Fukushima R, Kasamatsu A, Nakashima D, Higo M, Fushimi K, Kasama H, et al. Overexpression of Translocation Associated Membrane Protein 2 Leading to Cancer-Associated Matrix Metalloproteinase Activation as a Putative Metastatic Factor for Human Oral Cancer. *J Cancer*. 2018;9(18):3326-33.
- 113 Conlon GA, Murray GI. Recent advances in understanding the roles of matrix metalloproteinases in tumour invasion and metastasis. *J Pathol*. 2019;247(5):629-40.
- 114 Koshikawa N. MT1-MMP cleaves off the NH2-terminal portion of HB-EGF and converts it into a heparin-independent growth factor. *Cancer Res*. 2010;70:6093-103.