Case Report

* Corresponding Author:

Cell Biology Group, KIIT

School of Biotechnology,

KIIT University, Odisha,

bnbanerjee@kiitbiotech.ac.

Received: Feb. 18, 2020

Accepted: Jul. 11, 2020

India

in

E-mail:

Birendranath Banerjee, Molecular Stress and Stem

A Novel Balanced Chromosomal Translocation in an Azoospermic Male: A

Abhik Chakraborty¹, Indira Palo², Souvick Roy¹, Shu Wen Koh³, Manoor Prakash Hande³, Birendranath Banerjee^{1,4*}

1- Molecular Stress and Stem Cell Biology Group, School of Biotechnology, KIIT University, Bhubaneswar, Odisha, India

2- Department of Obstetrics and Gynecology, Amit Hospital, Odisha, India

3- Department of Physiology, Yong Loo Lin School of Medicine, National University of Singapore, Kent Ridge, Singapore

4- Division of Cytogenetics, inDNA Life Sciences Private Limited, Bhubaneswar, Odisha, India

Abstract

Background: Balanced translocation and azoospermia as two main reasons for recurrent pregnancy loss are known to be the leading causes of infertility across the world. Balanced translocations in azoospermic males are very rare and extensive studies need to be performed to elucidate the translocation status of the affected individuals.

Case Presentation: The cytogenetic characterization of a 28 year old male and his female partner is reported in this study. The male partner was diagnosed with nonobstructive azoospermia (NOA) and the couple was unable to conceive. Cytogenetic analysis by karyotyping through Giemsa-trypsin-giemsa banding technique (GTG) showed a novel balanced translocation, 46,XY,t(19;22)(19q13.4;22q11.2), 13ps+ in the male and the female karyotype was found to be 46,XX. Multicolor fluorescence in situ hybridization (mFISH) analysis on paternal chromosomal preparations confirmed both the region and origin of balanced translocation. The status of Y chromosome microdeletion (YMD) was analyzed and no notable microdeletion was observed. Furthermore, protein-protein interaction (PPI) network analysis was performed for breakpoint regions to explore the possible functional genetic associations. Conclusion: The azoospermic condition of the male patient along with novel balanced chromosomal translocation was responsible for infertility irrespective of its YMD status. Therefore, cytogenetic screening of azoospermic patients should be performed in addition to routine semen analysis to rule out or to confirm presence of any numerical or structural anomaly in the patient.

Keywords: Azoospermia, Balanced translocation, Infertility, Multicolor fluorescence in situ hybridization.

To cite this article: Chakraborty A, Palo I, Roy S, Koh SW, Hande MP, Banerjee B. A Novel Balanced Chromosomal Translocation in an Azoospermic Male: A Case Report. J Reprod Infertil. 2021;22(2):133-137. http://dx.doi.org/10.18502/jri.v22i2.5802.

Introduction

Infertility among men has become a major problem of the 21st century and 10-15% out of the total population of men have been reported to be infertile during their optimal reproductive age (1). More than 50% of the infertile cases are diagnosed as idiopathic infertility (2). During routine clinical examination, further genetic screening should be provided to individuals diagnosed with azoospermia (2). Earlier studies suggested that 5-10% out of the infertile men are diagnosed with non-obstructive azoospermia (NOA) (2). The occurrence rate of chromosomal anomalies or aberrations is reported to be higher in infertile males as compared to fertile males. Amongst the total number of chromosomal aberrations found in infertile men, Robertsonian trans Downloaded from http://www.jri.ir

J Reprod Infertil. 2021;22(2):133-137

Check for updates

Case Report

locations and numerical sex chromosome anomalies are the most common (3). Normally, carriers of balanced translocations lack any detectable phenotypic characteristics because most of the breakpoints reside in the non-repetitive regions which do not affect the expression of rearranged chromosomes. On the other hand, in 5% of the cases, the breakpoints can affect gene expressions by splitting the haploinsufficient genes which may cause spermatogenic arrest and result in infertility in carrier males of balanced translocations (4). Numerical chromosomal aberrations are usually found in azoospermia and severe oligoazoospermia cases. Klinefelter syndrome is the most common numerical chromosomal anomaly in 5% and 10% of severe oligoazoospermic and azoospermic patients, respectively (5). Rare cases of balanced Robertsonian or non-reciprocal translocation are reported to be involved in azoospermic patients (6). Carriers of balanced alterations have the ability to give birth, but after fertilization the embryos may possess an unbalanced alteration. It may further lead to death of the infant due to errors in segregation during meiosis (7). The present study is a case report of a novel chromosomal balanced translocation and molecular characterization of an azoospermic male patient and his female partner who were unable to attain pregnancy upon repeated attempts. The male had a novel balanced chromosomal translocation involving chromosomes19 and 22 along with an increased satellite region above one of short arms of chromosome 13. This is the first report describing a novel balanced chromosomal translocation involving chromosome 19 and 22, associated with azoospermia leading to male infertility.

Case Presentation

A 28 year old male (Proband) was referred for karyotyping on 14th of December 2018 for general karyotyping. The proband was presented with history of no successful pregnancy after being involved in three years of consanguineous marriage. Semen analysis report of the patient showed low sperm count in the sample provided, and he was diagnosed to have non-obstructive azoospermia (NOA). The proband was presented with no family history of either miscarriage or genetic abnormality among family members for the past two generations as represented with pedigree analysis (Figure 1). Heparinized blood was collected from the proband and his wife for karyotype analysis followed by molecular studies including m-



Figure 1. Pedigree analysis: Pedigree analysis of the proband up to previous three generations. White symbol (Square or circle)-a healthy person, white symbol (Square or circle with diagonal stroke)-death of that individual, individuals connected with double line–involved in consanguineous marriage, downward line with a single line–no issues, downward line with a double line–individual with infertility

FISH and YMD analysis. Further family screening studies were not carried out due to consent denial aimed to resolve the origin of the translocation.

Results

Whole blood was obtained in heparinized tubes from both partners and was processed for karyotyping. In vitro culture was performed using heparinized peripheral blood with RPMI-1640 media complemented with 10% fetal bovine serum, 1% penicillin-streptomycin, and 0.1 µg/ml of colchicine was added at the 67th hour to arrest the cells in metaphase after incubation; next, the cells were harvested, treated with 0.56% potassium chloride, then treated with chilled Carnoy's fixative until a white pellet was obtained. After that GTG-banding was performed. A novel balanced translocation involving long arm of chromosome 19 and long arm of chromosome 22 was observed. According to ISCN 2016, the karyotype of the male and the female was denoted as 46,XY,t(19;22)(19q13.4;22q11.2),13ps+ and 46,XX, respectively (Figure 2). Also, m-FISH was performed as described earlier with minor modifications (8). m-FISH data coincided with the karyotype analysis and m-FISH data confirmed the chromosomes involved in the balanced translocation (Figure 3).

YMD analysis was performed to rule out its association with azoospermia. DNA was isolated by using DNA extraction kit (QIAGEN DNA mini kit, catalogue number-51304) according to the manufacturer's instructions. A PCR reaction was carried out using primers for AZF loci (AZFa, AZFb and AZFc) involved in Y chromosome microdeletion (9-11) and the PCR product was checked in 1.5% agarose gel. No microdeletion in AZF loci was found in the male partner (Figure 4).

Chakraborty A, et al. JR



Figure 2. Karyotype analysis: GTG-banded chromosomal analysis of the proband, which reveals the presence of a balanced translocation involving chromosome 19 and 22 along with chromosome 13 ps+.



Figure 3. Multicolor fluorescence in situ hybridization (mFISH): A stable balanced chromosomal translocation in the proband, detected by mFISH. Chromosomes involved in this translocation are indicated by arrows; a balanced chromosomal translocation is shown involving two chromosomes 19 and 22

In addition, protein-protein interaction of the proteins located in the breakpoint regions was performed using STRING software (Figure 5A and 5B). Minor alteration(s) in one or few genes in the breakpoint region might be responsible for developing infertility or lower fertility in the patient. Few proteins present in the breakpoint region interacted functionally with each other.



Figure 4. Y chromosome microdeletion study: 1.5% agarose gel image of the specimen for Y chromosome microdeletion analysis by PCR. Two different loci for AZFa, AZFb and AZFc along with SRY and ZFY (Control) were used for analysis along with a positive control for AZFa microdeletion. Lane 1 and 7: DNA ladder (250 *bp* plus)

Lane 2: Positive control for YMD (AZFa loci-SY86–318 *bp*)

Lane 3: PCR product for ZFY(495 *bp*) and SY86 (318 *bp*) [AZFa locus]

Lane 4: PCR product for SRY (472 *bp*) and SY134 (301 *bp*) [AZFb locus]

Lane 5: PCR product for SY254 (380 *bp*) [AZFc locus] and SY127 (274 *bp*) [AZFb]

Lane 6: PCR product for SY84 (326 *bp*) [AZFa locus] and SY255 (123 *bp*) [AZFc locus]

Lane 8: No template control



Figure 5. Breakpoint analysis through protein-protein interaction studies. A) STRING generated PPI networks between all the proteins present at the breakpoint location; B) PPI networks between the selected proteins involved in the breakpoint region, which has basic cellular functions

Discussion

Parental chromosomal rearrangements have been reported in 2-5% of the couples with unsuccessful pregnancy (7). These anomalies include reciprocal and Robertsonian translocations, insertions, inver-

JRI A Case of Balanced Translocation in an Azoospermic Male

sions and mosaicism (12). Robertsonian translocations account for 35% of the cases carrying a translocation, while 65% of the cases are of reciprocal type (12). Reciprocal translocations, found in 1 out of 500 people, do not produce any phenotypic effects but usually result in recurrent miscarriages, or offspring with chromosomal abnormalities, or infertility in the carriers (13). During meiosis, chromosomes involved in reciprocal translocation form quadrivalent complexes that segregate in such a way that there remain only two possible balanced translocations in the offspring out of 32 translocations (14). The formation of trivalent or quadrivalent structures in translocation carriers during meiosis may lead to reduced fertility, subfertility or infertility due to lack of time and the process involved and also disjunction of the structures which may further lead to production of genetically unbalanced gametes (15). Molecular characterization of breakpoints involved in translocations would elucidate not only the critical genes responsible for the clinical condition but also the mechanism of inactivation through interruption or by position effect (12). It could be speculated that disruption of critical genes through chromosomal rearrangements and their consequent functional impairment possibly result in infertility (12). The karyotype analysis showed a unique balanced translocation between chromosome 19 and 22 along with a satellite region present in chromosome 13. Our finding was further validated by performing mFISH, which also suggested balanced translocation between chromosome 19 and chromosome 22. The breakpoint locus was found to be 19q13.4 and 22q11.2 through karyotype analysis. Y chromosome microdeletion assay was performed (Figure 4) to rule out Y chromosome microdeletion, which is reported to be the common reason for infertility (9). There were not any notable Y chromosome microdeletions, suggesting that the translocation is playing a crucial role in azoospermia. Protein-protein interactions of the genes located at the breakpoints of chromosome 19 and chromosome 22 were further analyzed. Few proteins were filtered which were known to interact with each other and were located in the breakpoint regions (Figure 5A). The main focus was (According to functionality) on one gene, aurora kinase C (AURKC), which is known to be involved in segregation of the chromosomes during cell division (Cytogenetic location: 19q13.43, which is the long arm of chromosome 19 at position 13.43). AURKC plays a very important role to ensure that two dividing cells separate from each other with a complete set of chromosomes. Aurora kinase C is most abundant in male testes, where sperm are produced and stored. AURKC in the testes regulates the division of sperm cells, ensuring that every new sperm cell divides properly and contains one copy of each chromosome; therefore, expression of AURKC, if altered during the translocation, may have contributed to azoospermia (16). According to our bioinformatics analysis, AURKC was found to interact with BIRC8 and CDC45, and CDC45 in turn reacts with TANGO2, UFD1L, MRPL40, RANBP1, and TRMT2A (Figure 5B). All the interactions with a combined score greater than 0.4 were considered for finding a functional link between the proteins. All these proteins play a crucial role in cell division maintenance, protein transport and regulation, nucleic acid metabolism, and protection against apoptosis (17-21). As these proteins were previously known to be involved in basic functioning of the cells, it could be now speculated that alteration in some of the genes present in the breakpoint may lead to lower fertility or infertility of the proband. Further molecular validation is required to evaluate the role of above mentioned proteins in larger cohort of carriers of balanced translocations involving chromosome 19 and 22 in azoospermic individuals.

Conclusion

Karyotype analysis is mandatory for couples diagnosed with infertility to ascertain the chromosomal makeup which will further help in genetic counselling. Further, prenatal diagnosis in every subsequent pregnancy and oocyte or sperm donation followed by preimplantation genetic diagnosis are recommended when a parent proves to be carrier of chromosomal rearrangement. Alternation of expression of critical genes through chromosomal rearrangements could also underline pregnancy outcome. This is the first case till date with a balanced translocation of chromosome 19 and 22 which can be further contributed to the development of azoospermia in the proband.

Acknowledgement

We would like to acknowledge the contribution of the couple for participating in this study; also, we would like to thank inDNA Life Sciences Pvt. Ltd. for providing the sample for research use.

Conflict of Interest

The authors declare no conflicts of interest.

References

- 1. Esteves SC, Miyaoka R, Agarwal A. An update on the clinical assessment of the infertile male. [corrected]. Clinics (Sao Paulo). 2011;66(4):691-700.
- 2. Esteves SC. Novel concepts in male factor infertility: clinical and laboratory perspectives. J Assist Reprod Genet. 2016;33(10):1319-35.
- 3. Frydman N, Romana S, Le Lorc'h M, Vekemans M, Frydman R, Tachdjian G. Assisting reproduction of infertile men carrying a robertsonian translocation. Hum Reprod. 2001;16(11):2274-7.
- 4. Wang H, Jia Z, Mao A, Xu B, Wang S, Wang L, et al. Analysis of balanced reciprocal translocations in patients with subfertility using single-molecule optical mapping. J Assist Reprod Genet. 2020;37(3): 509-16.
- Foresta C, Garolla A, Bartoloni L, Bettella A, Ferlin A. Genetic abnormalities among severely oligospermic men who are candidates for intracytoplasmic sperm injection. J Clin Endocrinol Metab. 2005;90 (1):152-6.
- 6. Arafa MM, Majzoub A, AlSaid SS, El Ansari W, Al Ansari A, Elbardisi Y, et al. Chromosomal abnormalities in infertile men with azoospermia and severe oligozoospermia in Qatar and their association with sperm retrieval intracytoplasmic sperm injection outcomes. Arab J Urol. 2017;16(1):132-9.
- 7. Sheth FJ, Liehr T, Kumari P, Akinde R, Sheth HJ, Sheth JJ. Chromosomal abnormalities in couples with repeated fetal loss: An Indian retrospective study. Indian J Hum Genet. 2013;19(4):415-22.
- Hande MP, Azizova TV, Burak LE, Khokhryakov VF, Geard CR, Brenner DJ. Complex chromosome aberrations persist in individuals many years after occupational exposure to densely ionizing radiation: an mFISH study. Genes Chromosomes Cancer. 2005;44(1):1-9.
- Agarwal S, Agarwal A, Khanna A, Singh K. Microdeletion of Y chromosome as a cause of recurrent pregnancy loss. J Hum Reprod Sci. 2015;8(3):159-64.
- Krausz C, Hoefsloot L, Simoni M, Tüttelmann F, European Academy of A, European Molecular Genetics Quality N. EAA/EMQN best practice guidelines for molecular diagnosis of Y-chromosomal microdeletions: state-of-the-art 2013. Andrology. 2014;2(1):5-19.

- Simoni M, Bakker E, Krausz C. EAA/EMQN best practice guidelines for molecular diagnosis of ychromosomal microdeletions. State of the art 2004. Int J Androl. 2004;27(4):240-9.
- Arumugam B, Samuel CR, Thyagarajan SS. Balanced autosomal translocations in two women reporting recurrent miscarriage. J Clin Diagn Res. 2016;10(12):Gd01-GD03.
- 13. Dutta UR, Ponnala R, Dalal A. A novel de novo balanced reciprocal translocation t(18;22) associated with recurrent miscarriages: a case report. J Reprod Infertil. 2014;15(2):113-6.
- Ogilvie CM, Scriven PN. Meiotic outcomes in reciprocal translocation carriers ascertained in 3day human embryos. Eur J Hum Genet. 2002;10 (12):801-6.
- 15. Harton GL, Tempest HG. Chromosomal disorders and male infertility. Asian J Androl. 2012;14(1): 32-9.
- 16. Ray PF, Toure A, Metzler-Guillemain C, Mitchell MJ, Arnoult C, Coutton C. Genetic abnormalities leading to qualitative defects of sperm morphology or function. Clin Genet. 2017;91(2):217-32.
- 17. Kfir S, Basavaraja R, Wigoda N, Ben-Dor S, Orr I, Meidan R. Genomic profiling of bovine corpus luteum maturation. PloS One. 2018;13(3):e0194456.
- Zuazua-Villar P, Rodriguez R, Gagou ME, Eyers PA, Meuth M. DNA replication stress in CHK1depleted tumour cells triggers premature (S-phase) mitosis through inappropriate activation of Aurora kinase B. Cell Death Dis. 2014;5(5):e1253.
- 19. Lin M, Pedrosa E, Hrabovsky A, Chen J, Puliafito BR, Gilbert SR, et al. Integrative transcriptome network analysis of iPSC-derived neurons from schizophrenia and schizoaffective disorder patients with 22q11.2 deletion. BMC Syst Biol. 2016;10 (1):105.
- Devaraju P, Yu J, Eddins D, Mellado-Lagarde MM, Earls LR, Westmoreland JJ, et al. Haploinsufficiency of the 22q11.2 microdeletion gene Mrpl40 disrupts short-term synaptic plasticity and working memory through dysregulation of mitochondrial calcium. Mol Psychiatry. 2017;22(9): 1313-26.
- 21. Guna A, Butcher NJ, Bassett AS. Comparative mapping of the 22q11.2 deletion region and the potential of simple model organisms. J Neurodeve Disord. 2015;7(1):18.