



Exploring the Therapeutic Potential of Sodium Hydrosulfide in Alleviating Oxidative Stress and Ovarian Dysfunction in a Rat Model of Polycystic Ovary Syndrome

Maryam Vaziripour, Mahdiah Faghihi, Mina Ranjbaran, Bahareh Asadi, Arash Abdi, Farzaneh Kianian, Mahdi Hajiaqaee, Behjat Seifi *

- Department of Physiology, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran

Abstract

Background: Oxidative stress is known to play a key role in the occurrence of polycystic ovary syndrome (PCOS) as the most common cause of anovulatory infertility. The purpose of the current study was to investigate whether diminished activity of ovarian enzymes responsible for hydrogen sulfide (H₂S) production, cystathionine β-synthase (CBS), and cystathionine γ-lyase (CSE) contributes to oxidative stress in PCOS. The study also explored whether administration of sodium hydrosulfide (NaSH), an H₂S donor, could ameliorate PCOS symptoms by reducing oxidative stress.

Methods: The total eighteen rats were randomly assigned into three groups (n=6): control, PCOS, and PCOS+NaSH. PCOS was induced by intramuscular injection of estradiol valerate to induce PCOS in the PCOS and PCOS+NaSH groups. The PCOS+NaSH group received 30 μmol/L of NaSH in drinking water for 27 days after PCOS induction. Ovarian tissue samples were analyzed for oxidative stress indices including malondialdehyde (MDA) levels and superoxide dismutase (SOD) activity. Additional analyses measured H₂S levels, CBS, and CSE activity.

Results: PCOS induction led to a significant decrease in SOD activity, H₂S levels, and CBS and CSE activity, accompanied by a significant increase in MDA levels (p<0.0001). Furthermore, PCOS caused severe histological alterations in the ovaries. However, administration of NaSH effectively restored all measured parameters to pre-PCOS induction levels (p<0.0001).

Conclusion: This study showed that the decrease in the activity of H₂S-producing enzymes and H₂S levels may contribute to oxidative stress in PCOS. Therefore, administration of NaSH as a H₂S donor can be considered as a potential therapeutic strategy for PCOS patients.

Keywords: Cystathionine β-synthase, Cystathionine γ-lyase, Hydrogen sulfide, Infertility, Ovary, Oxidative stress, PCOS.

To cite this article: Vaziripour M, Faghihi M, Ranjbaran M, Asadi B, Abdi A, Kianian F, et al. Exploring the Therapeutic Potential of Sodium Hydrosulfide in Alleviating Oxidative Stress and Ovarian Dysfunction in a Rat Model of Polycystic Ovary Syndrome. *J Reprod Infertil.* 2024;25(2):133-139. <https://doi.org/10.18502/jri.v25i2.16007>.

* Corresponding Author:
Behjat Seifi, Department of
Physiology, Tehran
University of Medical
Sciences, Tehran, Iran
E-mail:
b-seifi@tums.ac.ir

Received: 7, Nov. 2023
Accepted: 7, Apr. 2024

Introduction

Polycystic ovary syndrome (PCOS) is a major public health concern that mainly affects the female reproductive system and is characterized by anovulation or oligo-ovulation, hyperandrogenism, and polycystic ovaries. The preva-

lence of PCOS is estimated to be 5-18%, making it the most common cause of anovulatory infertility (1-3).

From a therapeutic standpoint, the treatment options of PCOS depend primarily on the desired

clinical outcome, such as reducing hyperandrogenism symptoms and the treatment of infertility (4). However, currently available treatments are unable to completely resolve the adverse consequences of PCOS, which is attributed to the lack of comprehensive understanding of the pathophysiology of this disease (5).

That is why investigating the pathophysiology of PCOS and identifying new drug targets remains a hot topic of scientific research. Nevertheless, the findings of many studies show that oxidative stress plays a key role in the occurrence and development of PCOS (6-8). Oxidative stress refers to the imbalance between the intracellular production of oxidants and the activity of the cellular antioxidant system. In therefore, the accumulation of oxidants causes protein and deoxyribonucleic acid (DNA) damage and lipid peroxidation, which ultimately leads to cell dysfunction (9). However, understanding the vital role of oxidative stress in PCOS alone is insufficient. To achieve a more effective treatment of the disease, it is crucial to explore the origin of oxidative stress in PCOS patients.

Hydrogen sulfide (H_2S), recently recognized as the third gasotransmitter after nitric oxide (NO) and carbon monoxide (CO), is endogenously produced by cystathionine β -synthase (CBS), cystathionine γ -lyase (CSE), and 3-mercaptopyruvate sulfurtransferase (3-MST), which have different expressions in different mammalian tissues (10, 11). For example, evidence has shown that CBS and CSE play major roles in the female reproductive system (12). The importance of H_2S produced by these two enzymes in female reproduction became more apparent when reports of changes in the H_2S signaling pathway in female reproductive diseases such as PCOS were published (13). Given the potent antioxidant role of H_2S (14), the purpose of this study was to investigate the potential link between decreased activity of H_2S -producing enzymes, CBS and CSE, and oxidative stress in PCOS. The objective was to assess whether administration of sodium hydrosulfide (NaSH) as an H_2S donor can alleviate PCOS symptoms by restoring H_2S levels and mitigating oxidative stress. The second goal was to evaluate the impact of NaSH on enzymatic activity, histological changes in the ovaries, and oxidative stress markers in a rat model of PCOS. The findings may contribute to understanding the role of H_2S in PCOS pathology and offer insights into potential therapeutic strategies.

Methods

Experimental design: For this study, a total of 18 adult female Wistar rats weighing 180-200 g were obtained from the Department of Physiology of Tehran University of Medical Sciences. Animal care and the general protocols for animal use were approved by the Experimental Animal Committee of Tehran University of Medical Sciences.

All animals were kept at a constant temperature of 20-22°C and a 12 hr light-dark cycle with free access to standard laboratory chow and tap water. Six days before the start of the experiment, the animals were checked for regular estrous cycles by vaginal sampling and then randomly divided into three groups (n=6). The control group received a single intramuscular dose of 0.2 ml of sesame oil. The PCOS group was given a single intramuscular dose of 4 mg/kg of estradiol valerate (Aburaihan Pharmaceutical Co, Iran) dissolved in 0.2 ml of sesame oil (15). The PCOS+NaSH group was treated with 30 μ mol/L of NaSH (Sigma Aldrich, USA) in drinking water for 27 days, starting on the third day after PCOS induction (16). At the end of the procedure, the rats were anesthetized with intraperitoneal injections of ketamine (100 mg/kg) and xylazine (10 mg/kg), and their abdomens were opened. Then, the ovaries were resected and washed in cold saline on ice. For biochemical and oxidative stress measurement, the tissues of the left ovary were quickly frozen in liquid nitrogen and the samples were stored at -70°C until further study. The right ovary tissues were fixed in 10% formalin for histological evaluation.

Determination of the estrous cycle: The estrous cycle was determined by collecting a vaginal smear, whereby 100 μ l of sterile saline was gently introduced into the vagina and then aspirated from the tip of the sampler. This allowed for the assessment of vaginal epithelial cells. The procedure was repeated 3 to 4 times. The liquid containing a few drops of cell suspension was poured onto a glass slide, air-dried, and stained using 0.1% crystal violet. Then, the stained slides were observed under a light microscope at 200 \times magnification (17).

Measurement of ovarian oxidative stress indices: MDA levels in the ovarian tissue were determined according to the method of Esterbauer and Cheeseman. MDA reacts with thiobarbituric acid and produces a pink pigment with a maximum absorption of 532 nm (18).

Enzyme-linked immunosorbent assay (ELISA) kit (Nasdox; Navand Salamat, Iran) was used to measure SOD activity. Briefly, 100 mg of the ovarian tissue was homogenized with 500 μ l of lysis buffer and centrifuged at 12,000 rpm for 5 min at 4°C. Next, 50 μ l of the obtained supernatant was added to the wells of the microtiter plate, followed by addition of 200 μ l of reagent 1 and 50 μ l of reagent 2. After 5 min of incubation at room temperature and darkness, the absorbance of the samples was measured using an ELISA reader at 405 nm (19).

Measurement of ovarian H₂S levels: In total, 50 mg of the ovarian tissue were homogenized in 500 μ l of phosphate-buffered saline and incubated with L-cysteine, pyridoxal phosphate, and normal saline for 30 min at 37°C. Then, trichloroacetic acid and zinc acetate were added to them. After 15 min, N, N-dimethyl-p-phenylenediamine sulfate and ferric chloride were introduced, and the light absorption of the samples was read using a microplate reader at a wavelength of 660 nm (20).

Assessment of ovarian CBS and CSE activity: The activity of CBS and CSE in the ovarian tissue was evaluated with a commercially available ELISA kit (Abbexa, United Kingdom). Next, 100 μ l of reagent was added to 100 μ l of ovarian tissue supernatant and incubated for 2 hr. In the next step, 90 μ l of 3,3',5,5'-tetramethylbenzidine (TMB) solution was added to each well, and the optical density of each well was immediately measured by an ELISA reader at a wavelength of 450 nm.

Evaluation of ovarian histopathology: The ovarian tissues fixed in formalin were embedded in paraffin blocks and cut into 5 μ m sections. The tissue sections were then stained with hematoxylin and eosin and observed with a light microscope at 400 \times magnification (21).

Statistical analysis: The data is presented as the mean \pm standard error of the mean (SEM). Comparisons between groups were conducted by one-way analysis of variance (ANOVA) followed by Tukey's post hoc test. The level of significance was set at p<0.05.

Results

Ovarian oxidative stress: PCOS resulted in a significant increase in ovarian MDA levels compared to the control group (4.90 \pm 0.69 vs. 0.81 \pm 0.09) (Figure 1A, p<0.0001). NaSH administration decreased ovarian MDA levels compared to the

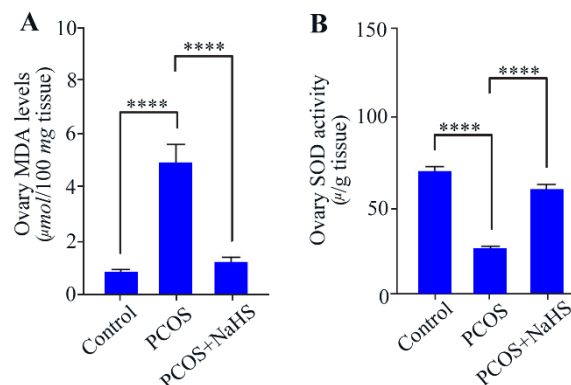


Figure 1. Changes in ovarian malondialdehyde (MDA) levels (A) and superoxide dismutase (SOD) activity (B) in different groups (n=6). Data are expressed as mean \pm SEM. # indicates p<0.0001 compared to the control group. * indicates p<0.0001 compared to the PCOS group. PCOS: polycystic ovary syndrome, NaSH: sodium hydrosulfide

PCOS group (1.17 \pm 0.19 vs. 4.90 \pm 0.69) (Figure 1A, p<0.0001).

PCOS also resulted in a significant decrease in ovarian SOD activity compared to the control group (25.03 \pm 1.63 vs. 68.72 \pm 2.71) (Figure 1B, p<0.0001). NaSH administration increased ovarian SOD activity compared to the PCOS group (58.62 \pm 2.87 vs. 25.03 \pm 1.63) (Figure 1B, p<0.0001).

Ovarian H₂S levels: PCOS resulted in a significant decrease in ovarian H₂S levels compared to the control group (0.62 \pm 0.07 vs. 1.95 \pm 0.06) (Figure 2, p<0.0001). NaSH administration increased

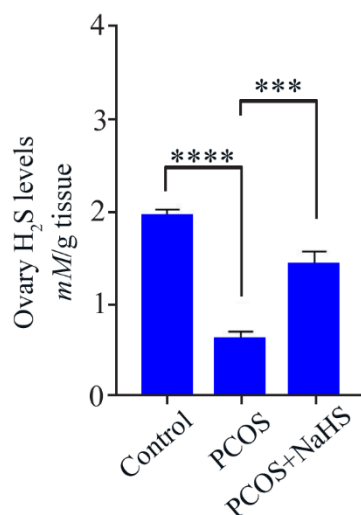


Figure 2. Changes in hydrogen sulfide (H₂S) levels in different groups (n=6). Data are expressed as mean \pm SEM. # Indicates p<0.0001 compared to the control group. * Indicates p<0.0001 compared to the PCOS group. PCOS: polycystic ovary syndrome, NaSH: sodium hydrosulfide

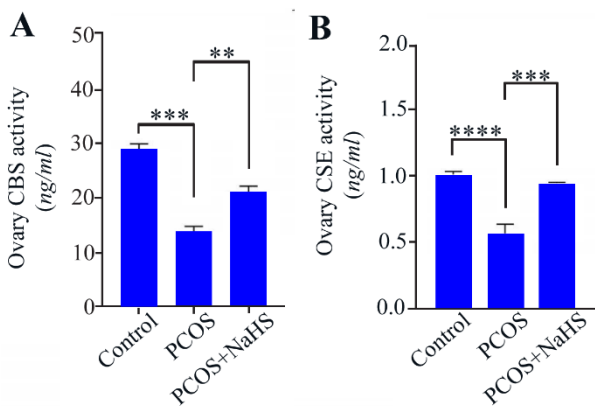


Figure 3. Changes in cystathionine β-synthase (CBS) (A) and cystathionine γ-lyase (CSE) (B) activity in different groups (n=6). Data are expressed as mean±SEM. # Indicates p<0.0001 compared to the control group. * Indicates p<0.001 compared to the PCOS group. PCOS: polycystic ovary syndrome, NaSH: sodium hydrosulfide

ovarian H₂S levels compared to the PCOS group (1.42±0.14 vs. 0.62±0.07) (Figure 2, p<0.0001).

Ovarian CBS and CSE activity: PCOS resulted in a significant decrease in ovarian CBS activity compared to the control group (13.68±0.95 vs. 28.30±1.51) (Figure 3A, p<0.0001). NaSH administration increased ovarian CBS activity compared to the PCOS group (20.85±1.16 vs. 13.68±0.95) (Figure 3A, p<0.001).

PCOS also resulted in a significant decrease in ovarian CSE activity compared to the control group (0.54±0.07 vs. 0.98±0.03) (Figure 3B, p<0.0001). NaSH administration increased ovarian CSE activity compared to the PCOS group (0.92±0.01 vs. 0.54±0.07) (Figure 3B, p<0.001).

Ovarian Histological changes: There were no detectable cysts in the ovarian tissue of the control group (Figure 4A). In contrast, the ovarian tissue of the PCOS group exhibited significant histological alterations, characterized by the presence of cystic follicles (CF), corpus luteum (CL), preantral follicles (PA), and antral follicles (AF), when compared to the control group (Figure 4B). NaSH administration significantly reduced these histological changes (Figure 4C).

Discussion

In the present study, the induction of PCOS in the rats resulted in oxidative stress in the ovary, decreased ovarian H₂S levels, decreased ovarian CBS and CSE activity, and led to significant histological changes in the ovary. However, the ad-

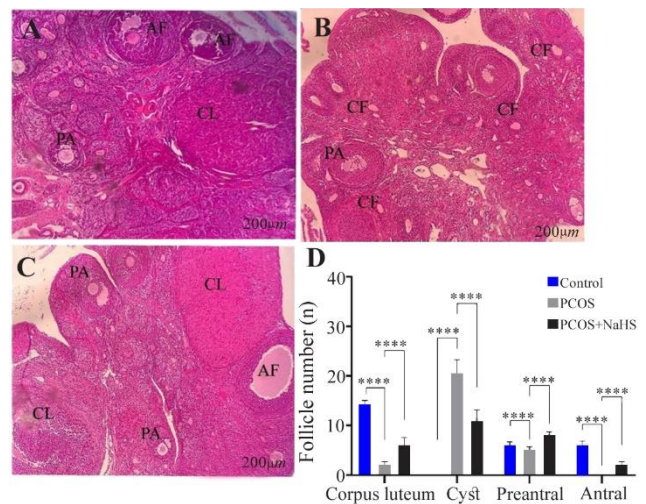


Figure 4. Changes in ovarian histology in different groups depicted with hematoxylin and eosin staining using light microscopy (n=3). A) control, B) PCOS, C) PCOS+ NaSH group. In the control group (A), there was no detectable damage to the ovarian tissue. The PCOS group (B) exhibited significant histological changes, including the presence of cystic follicles (CF), corpus luteum (CL), preantral follicles (PA), and antral follicles (AF). Administration of NaSH reduced the severity of these histopathological damages (C). Scale bar: 100 μm. D) The number of preantral follicles, antral follicles, corpus luteum, and cysts in different groups. Data are expressed as mean±SEM. # Indicates p<0.0001 compared to the control group. * Indicates p<0.0001 compared to the PCOS group. PCOS: polycystic ovary syndrome, NaSH: sodium hydrosulfide

ministration of NaSH significantly reversed all these parameters, restoring them towards normal levels.

An important consequence following various pathological insults is the overproduction of free radicals, which exceeds the body's antioxidant capacity, resulting in oxidative stress. In this condition, protein and DNA degradation, as well as membrane lipid peroxidation leads to extensive tissue damage (9). Considering the important role of oxidative stress in the pathogenesis of PCOS, MDA levels were assessed as a valuable parameter of lipid peroxidation in the first step, and the activity of SOD as a major endogenous antioxidant in the ovarian tissues was evaluated in the second step (18, 22). In this study, MDA levels in the ovarian tissues were significantly higher in the PCOS group compared to the control group. The presence of oxidative stress in PCOS negatively affects oocyte maturation and the development of embryos (23). Previous studies have also suggested a direct relationship between MDA levels and

the prevalence of PCOS (24). Also, SOD activity in rats with PCOS decreased significantly compared to the control group. SOD plays an essential role in protecting ovarian cells against reactive oxygen species (ROS), while SOD depletion leads to corpus luteum apoptosis and deterioration. In addition, our histological results showed a decrease in the number of corpora lutea in the PCOS group. One of the hallmarks of PCOS is the occurrence of follicular atresia, which arises as a result of abnormal ovulation caused by increased ROS levels (25).

In the next step, ovarian H₂S levels were examined in PCOS rats as a potential source of oxidative stress. H₂S is recognized as a potent antioxidant due to its ability to scavenge free radicals and enhance the activity of other free radical scavengers, such as glutathione. Moreover, changes in the signaling pathway of H₂S have been reported in female reproductive diseases such as PCOS (13, 26). Our finding showed a significant reduction in ovarian H₂S levels in PCOS rats.

Subsequently, the activity of two enzymes, CBS and CSE, were evaluated which are key players in the production of ovarian H₂S. Moreover, the underlying cause of the decreased ovarian H₂S levels was assessed in our study. The results indicated decreased activity of ovarian CBS and CSE in PCOS rats. Similarly, Bries et al. stated that letrozole-induced PCOS decreased CBS expression levels and thus its activity in rat ovaries (27). Furthermore, CBS plays an important role in the production of cystathionine from homocysteine, and therefore, decreased CBS activity leads to hyperhomocysteinemia, which has been reported to be a major risk factor for reproductive disorders such as PCOS (28). In addition, while both CBS and CSE enzymes produce H₂S, CSE uniquely converts cystathionine to cysteine, which subsequently produces H₂S. Importantly, cysteine deficiency has been shown to induce oxidative stress, mitochondrial dysfunction, and apoptosis in ovarian cells (29).

PCOS is a common endocrine and metabolic disorder associated with adipose dysfunction, chronic low-grade inflammation, high prevalence of obesity, and insulin resistance (30, 31). Common treatments for PCOS include a range of approaches including administration of metformin, combined oral contraceptive pills (COCPs), spironolactone, and topical treatments for managing hirsutism and acne (32). Because PCOS is a chronic disease and long-term use of these treat-

ments is associated with various complications such as gastrointestinal disorders and hyperkalemia, research continues to find appropriate treatment strategies with few side effects (33, 34). Considering the significant role of oxidative stress in the development of PCOS, and the findings indicating the involvement of H₂S and its producing enzymes in the induction of oxidative stress during PCOS, a final investigation was conducted to assess whether NaSH, as a H₂S donor, could mitigate this disease and its underlying pathological mechanisms in rats. It is important to note that the induction of PCOS led to the arrest of follicles in the pre-ovulatory stage, which is considered a diagnostic hallmark of PCOS. In contrast, the administration of NaSH caused normal follicular and oocyte maturation, a decrease in the number of cysts, and an increase in the number of pre-antral and antral follicles. Based on the results of this study, it seems that NaSH has exerted these positive effects through increasing ovarian CBS and CSE activity and increasing ovarian H₂S levels and thus improving oxidative stress status in the ovary.

Conclusion

The findings of the present study suggest that the decrease in ovarian H₂S levels, attributed to decreased activity of the enzymes CBS and CSE, may contribute to oxidative stress in PCOS. NaSH administration appears to ameliorate the ovarian activity of CBS and CSE, leading to increased H₂S levels. Therefore, this intervention may be considered as a promising therapeutic strategy for PCOS patients.

Acknowledgement

The authors would like to express their gratitude to Tehran University of Medical Sciences for financial support of the thesis from which this article originated.

Funding: This research was supported by a grant (no=1400-2-372-53181) from Tehran University of Medical Sciences.

Conflict of Interest

The authors declare that they have no conflict of interest.

References

1. Singh S, Pal N, Shubham S, Sarma DK, Verma V, Marotta F, et al. Polycystic ovary syndrome: etiology, current management, and future therapeutics. *J Clin Med.* 2023;12(4):1454.

2. Joham AE, Norman RJ, Stener-Victorin E, Legro RS, Franks S, Moran LJ, et al. Polycystic ovary syndrome. *Lancet Diabetes Endocrinol.* 2022;10(9):668-80.
3. Kim CH, Lee SH. Effectiveness of lifestyle modification in polycystic ovary syndrome patients with obesity: a systematic review and meta-analysis. *Life (Basel).* 2022;12(2):308.
4. Bednarska S, Siejka A. The pathogenesis and treatment of polycystic ovary syndrome: what's new? *Adv Clin Exp Med.* 2017;26(2):359-67.
5. Bargiota A, Diamanti-Kandarakis E. The effects of old, new and emerging medicines on metabolic aberrations in PCOS. *Ther Adv Endocrinol Metab.* 2012;3(1):27-47.
6. Dubey P, Reddy S, Boyd S, Bracamontes C, Sanchez S, Chattopadhyay M, et al. Effect of nutritional supplementation on oxidative stress and hormonal and lipid profiles in PCOS-affected females. *Nutrients.* 2021;13(9):2938.
7. Mancini A, Bruno C, Vergani E, d'Abate C, Giacchi E, Silvestrini A. Oxidative stress and low-grade inflammation in polycystic ovary syndrome: controversies and new insights. *Int J Mol Sci.* 2021;22(4):1667.
8. Shan H, Luo R, Guo X, Li R, Ye Z, Peng T, et al. Abnormal endometrial receptivity and oxidative stress in polycystic ovary syndrome. *Front Pharmacol.* 2022;13:904942.
9. Liu X, Hussain R, Mehmood K, Tang Z, Zhang H, Li Y. Mitochondrial-Endoplasmic Reticulum Communication-Mediated Oxidative Stress and Autophagy. *Biomed Res Int.* 2022;2022:6459585.
10. Hao Y, Wang H, Fang L, Bian J, Gao Y, Li C. H₂S donor and bone metabolism. *Front Pharmacol.* 2021;12:661601.
11. Cirino G, Szabo C, Papapetropoulos A. Physiological roles of hydrogen sulfide in mammalian cells, tissues, and organs. *Physiol Rev.* 2023;103(1):31-276.
12. Gao L, Wu TW, Ni X. Gas transmitters in female reproductive system. *Sheng Li Xue Bao.* 2016;68(5):611-20.
13. Mitidieri E, Vanacore D, Turnaturi C, Sorrentino R, d'Emmanuele di Villa Bianca R. Uterine dysfunction in diabetic mice: the role of hydrogen sulfide. *Antioxidants (Basel).* 2020;9(10):917.
14. Murphy B, Bhattacharya R, Mukherjee P. Hydrogen sulfide signaling in mitochondria and disease. *FASEB J.* 2019;33(12):13098-125.
15. Daneasa A, Cucolas C, Lenghel LM, Olteanu D, Orasan R, Filip GA. Letrozole vs estradiol valerate induced PCOS in rats: glycemic, oxidative and inflammatory status assessment. *Reproduction.* 2016;151(4):401-9.
16. Lorian K, Kadkhodae M, Kianian F, Abdi A, Ranjbaran M, Ashabi G, et al. Long-term NaHS administration reduces oxidative stress and apoptosis in a rat model of left-side varicocele. *Andrologia.* 2020;52(2):e13496.
17. Ramadoss M, Vijayaraman M, Jessica A. Estradiol valerate dose determination for PCOS induction. *Int J Pharm Biol Sci.* 2019;9(2):1131-6.
18. Kianian F, Seifi B, Kadkhodae M, Sadeghipour HR, Ranjbaran M. Nephroprotection through modifying the apoptotic *tnf- α /erk1/2/bax* signaling pathway and oxidative stress by long-term sodium hydrosulfide administration in ovalbumin-induced chronic asthma. *Immunol Invest.* 2022;51(3):602-18.
19. Pourmirzaei F, Ranjbaran M, Kadkhodae M, Kianian F, Lorian K, Abdi A, et al. Sperm and testicular dysfunction during cecal ligation and puncture-induced sepsis in male rats and effects of tannic acid through reducing testicular oxidative stress and inflammation. *Iran J Basic Med Sci.* 2021;24(11):1554-60.
20. Shafie A, Kianian F, Ashabi G, Kadkhodae M, Ranjbaran M, Hajiaqaei M, et al. Beneficial effects of combination therapy with testosterone and hydrogen sulfide by reducing oxidative stress and apoptosis: Rat experimental varicocele model. *Int J Reprod Biomed.* 2022;20(11):941-54.
21. Chinwe GS, Azuka OI, Aadaeze NC. Resveratrol supplementation rescues pool of growing follicles and ovarian stroma from Cisplatin-induced toxicity on the ovary in sprague-dawley rats: an experimental study. *Int J Reprod Biomed.* 2018;16(1):19-30.
22. Ighodaro O, Akinloye O. First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): their fundamental role in the entire antioxidant defence grid. *Alexandria J Med.* 2018;54(4):287-93.
23. Bandariyan E, Moghiseh A, Ahmadi A. The effect of lutein and Urtica dioica extract on in vitro production of embryo and oxidative status in polycystic ovary syndrome in a model of mice. *BMC Complement Med Ther.* 2021;21(1):55.
24. Nawrocka-Rutkowska J, Szydłowska I, Jakubowska K, Olszewska M, Chlubek D, Rył A, et al. Assessment of the parameters of oxidative stress depending on the metabolic and anthropometric status indicators in women with PCOS. *Life (Basel).* 2022;12(2):225.
25. Zeber-Lubecka N, Ciebiera M, Hennig EE. Polycystic ovary syndrome and oxidative stress-from

- bench to bedside. *Int J Mol Sci.* 2023;24(18):14126.
26. Scammahorn JJ, Nguyen IT, Bos EM, Van Goor H, Joles JA. Fighting oxidative stress with sulfur: Hydrogen sulfide in the renal and cardiovascular systems. *Antioxidants (Basel).* 2021;10(3):373.
 27. Bries AE, Webb JL, Vogel B, Carrillo C, Keating AF, Pritchard SK, et al. Letrozole-induced polycystic ovary syndrome attenuates cystathionine- β synthase mRNA and protein abundance in the ovaries of female sprague dawley rats. *J Nutr.* 2021;151(6):1407-15.
 28. Li D, Liu HX, Fang YY, Huo JN, Wu QJ, Wang TR, et al. Hyperhomocysteinemia in polycystic ovary syndrome: decreased betaine-homocysteine methyltransferase and cystathionine β -synthase-mediated homocysteine metabolism. *Reprod Biomed Online.* 2018;37(2):234-41.
 29. Novera W, Lee ZW, Nin DS, Dai MZY, Binte Idres S, Wu H, et al. Cysteine deprivation targets ovarian clear cell carcinoma via oxidative stress and iron-sulfur cluster biogenesis deficit. *Antioxid Redox Signal.* 2020;33(17):1191-208.
 30. Areloegbe SE, Peter MU, Oyeleke MB, Olaniyi KS. Low-dose spironolactone ameliorates adipose tissue inflammation and apoptosis in letrozole-induced PCOS rat model. *BMC Endocr Disord.* 2022;22(1):224.
 31. Pruet JE, Everman SJ, Hoang NH, Salau F, Taylor LC, Edwards KS, et al. Mitochondrial function and oxidative stress in white adipose tissue in a rat model of PCOS: effect of SGLT2 inhibition. *Biol Sex Differ.* 2022;13(1):45.
 32. Witchel SF, Oberfield SE, Peña AS. Polycystic ovary syndrome: pathophysiology, presentation, and treatment with emphasis on adolescent girls. *J Endocr Soc.* 2019;3(8):1545-73.
 33. Greff D, Juhász AE, Vánca S, Váradi A, Sipos Z, Szinte J, et al. Inositol is an effective and safe treatment in polycystic ovary syndrome: a systematic review and meta-analysis of randomized controlled trials. *Reprod Biol Endocrinol.* 2023;21(1):10.
 34. deOliveira TA, Marchesan LB, Spritzer PM. Potassium levels in women with polycystic ovary syndrome using spironolactone for long-term. *Clin Endocrinol (Oxf).* 2024;100(3):278-83.