



Prevalence and Molecular Characterization of Macrolide Resistance in Clinical Isolates of Beta Hemolytic Streptococci from a Tertiary Care Teaching Hospital in Kerala, India

Ann George, Rosmi Jose *, Chithra Valsan

Department of Microbiology, Jubilee Mission Medical College & Research Institute, Thrissur, Kerala, India.

ARTICLE INFO

Article type:

Research Article

Article history:

Received: 28 May 2023

Revised: 26 Jun 2023

Accepted: 16 Aug 2023

Published: 03 Sep 2023

Keywords:

Beta Hemolytic
Streptococci, Genotypes,
Inducible Clindamycin
Resistance, Macrolide
Resistance.

ABSTRACT

Background: Beta-hemolytic streptococci (BHS) are responsible for both invasive and noninvasive infections, and the preferred treatment for these infections is penicillin due to the distinctive characteristics of these bacteria. However, in patients who cannot tolerate β -lactam antibiotics, macrolides and clindamycin are important alternative options for treating BHS infections. This study aimed to analyse the pattern of macrolide resistance among clinical strains of beta-hemolytic streptococci (BHS).

Methods: Beta hemolytic streptococci isolated from clinical specimens during December 2018 to May 2020 were included in this study. Identification of the isolates were done by conventional and Vitek 2 method. All isolates were subjected to serogrouping. Antibiotic susceptibility testing done by disc diffusion method. Genes encoding macrolide resistance were detected by conventional multiplex polymerase chain reaction.

Results: A total of 129 beta hemolytic streptococcal isolates were obtained which included 27 *S. pyogenes* (20.9%), 77 *S. agalactiae* (59.7%), 23 *S. dysgalactiae* spp equisimilis (17.8%) and one isolate of *S. anginosus* and *S. porcinus* each (0.8%). Erythromycin, clindamycin, quinuprisin and tetracycline resistance were found to be 20.2%, 12.1%, 16.3% and 51.2% respectively. Among the 26 erythromycin resistant isolates, 12(46.2%) were inducible clindamycin resistant phenotype. Out of 26 erythromycin resistant isolates, 7(26.9%) isolates were harbouring *erm(A)* gene, 10(38.5%) *erm(B)* and 9(34.6%) *mef(A)* gene.

Conclusion: Our study highlights the importance of routine antibiotic susceptibility testing for beta-haemolytic streptococci, as well as the detection of inducible resistance to prevent therapeutic failure.

- **Please cite this paper as:** George A, Jose R, Valsan C. Prevalence and Molecular Characterization of Macrolide Resistance in Clinical Isolates of Beta Hemolytic Streptococci from a Tertiary Care Teaching Hospital in Kerala, India. *J Med Bacteriol.* 2023; **11** (3, 4): pp.25-31.

Introduction

Beta-hemolytic streptococci (BHS) are known to cause a diverse array of clinical presentations ranging from mild infections to potentially fatal ones. Group A streptococci (GAS) and Group B streptococci (GBS) constitute the most important and prevalent members of this group. GAS can cause conditions like self-limiting pharyngitis or even life-threatening toxic shock syndrome. Group B streptococci (GBS) remains as a leading cause of infections in neonates and pregnant women and also cause invasive diseases in children and non-pregnant adults. Group C (GCS) and group G (GGS) streptococci are commonly found in the throat, skin, gastrointestinal tract, and female genital tract as harmless inhabitants. Recent studies indicate that GCS and GGS, previously considered weak pathogens, can cause infections similar to those caused by GAS (1, 2).

Penicillin remains the drug of choice of BHS infections but there are reports of rising minimum inhibitory concentration (MIC) or diminished susceptibility to penicillin. Macrolides and clindamycin are important alternatives in the treatment of BHS infections in β -lactam-intolerant patients. But high variable resistance to macrolide and therapeutic failure are increasing which substantiate the need for antimicrobial susceptibility testing of BHS isolates (3, 4).

This study was conducted to analyse the pattern of macrolide resistance among clinical isolates of beta hemolytic streptococci.

Materials and Methods

Study design

The study was conducted in the department of Microbiology, Jubilee Mission Medical College & Research Institute, Thrissur, for a period of 18 months (December 2018 to May 2020) on clinical isolates of beta hemolytic streptococci obtained from various specimens received for routine culture. All consecutive non repetitive beta

haemolytic streptococci obtained during the study period were included in this study.

Identification of the isolates

All isolates were subjected to Gram stain, catalase test, bacitracin susceptibility, CAMP test and bile esculin hydrolysis. Species were identified based on the biochemical characteristics (5) and confirmed by Vitek 2 GP cards (bioMerieux India Pvt Ltd). Serogrouping of the isolates were done using *Streptococcus* grouping kit (Lab21Healthcare, Cameberley).

Antibiotic susceptibility testing

All isolates were subjected to antibiotic susceptibility testing by Kirby Bauer disc diffusion method on Mueller Hinton agar with 0.5% sheep blood. Susceptibility to penicillin, erythromycin, clindamycin, tetracycline, quinupristin, linezolid and vancomycin were tested and the results interpreted based on CLSI guidelines (6). All isolates were subjected to D test by placing erythromycin and clindamycin discs at 12mm apart (edge to edge). Erythromycin resistant isolates were classified into inducible clindamycin resistance (iMLS_B), constitutive clindamycin resistance (cMLS_B) and M phenotype based on the D test phenotype (7).

Genotyping of Macrolide resistance

All macrolide resistant strains were subjected to conventional multiplex polymerase chain reaction to detect the genes responsible for the macrolide resistance. The genes studied were *ermA*, *ermB*, and *mefA*. Forward and reverse primers for the corresponding genes were obtained from a previous published study (8).

The PCR products were analyzed by electrophoresis with 2% agarose gels in TBE buffer. The gels were stained with ethidium bromide (75 μ l in 500ml distilled water) and PCR products were visualized with UV light using gel documentation system.

Result

A total of 129 beta hemolytic streptococcal isolates were obtained from various clinical samples during the study period. BHS isolates included 27 *S. pyogenes* (20.9%), 77 *S. agalactiae* (59.7%), 23 *S. dysgalactiae* spp *equisimilis* (17.8%) and one isolate of *S. anginosus* and *S. porcinus* each (0.8%). Majority of the isolates were obtained from pus swabs or exudates (n=71, 55 %), 32 were from urine (24.8%), 13 from cervical swabs (10.1%), 8 isolates from blood (6.2%), 4 from throat swab (3.1%) and one isolate from cerebrospinal fluid (0.8%) (Table 1).

All isolates were susceptible to penicillin, vancomycin and linezolid. Erythromycin, clindamycin, quinupristin and tetracycline resistance were found to be 20.2%, 12.1%, 16.3% and 51.2% respectively (Table 2). Among the 26 erythromycin resistant isolates, majority were inducible resistant phenotype (46.2%), followed by M phenotype (34.6%) and constitutive resistance (19.2%) (Table 3). Out of the 26 erythromycin resistant isolates, 7(26.9%) isolates harboured *erm(A)* gene, 10(38.5%) *erm(B)* and 9(34.6%) *mef(A)* gene (Table 4). Among the 12 isolates with inducible clindamycin resistance six isolates harboured *erm(A)* and six had *erm(B)* gene. Among the isolates with constitutive resistance four had *erm(B)* and one had *erm(A)*. All nine isolates with M phenotype harboured *mef(A)* gene (Table 5).

Discussion

Beta hemolytic streptococci are important causative agent of various invasive and noninvasive infections. Even though these bacteria remain susceptible to penicillin, variable resistance to other antibiotics have been reported from India and other parts of the world (9-11).

The present study was conducted in an 1800 bedded tertiary care teaching hospital, catering to

patients from three neighboring districts of central Kerala. All clinically significant isolates of beta hemolytic streptococci obtained from different clinical specimens during the study period were included in this study. Although our study was conducted at a single centre, we were able to collect a substantial number of BHS isolates over an 18-month period. In contrast to previous studies where GAS was more common (8), our study found that GBS isolates made up nearly 60% of all isolates. Low isolation rate of GAS may be attributed to variations in the way cultures are sent for treatment of streptococcal pharyngitis.

The susceptibility of all isolates to penicillin was similar to previous studies, but resistance to other antibiotics varied, with the highest resistance observed for tetracyclines. Over half of the isolates were found to be resistant to tetracycline. In comparison to a prior multicentric study in India where 79% of BHS isolates were found to be resistant to erythromycin, the macrolide resistance in our study was low (20.2%) (8). This might be due to the prevalence of non-Group A BHS isolates, particularly GBS, in our study. Among the BHS isolates, GAS isolates exhibited higher macrolide resistance than the others. A study from south India conducted by Tintu et al. reported resistance rates of 53%, 33%, and 58% to erythromycin, clindamycin, and tetracycline, respectively (12). Clindamycin and quinupristin resistance in our isolates were slightly lower than macrolide resistance, which is in accordance with the phenotypes demonstrated in the D test, indicating that 34.6% of macrolide-resistant isolates were of the M phenotype.

Out of the 26 macrolide-resistant isolates in this study, 12 (46.2%) isolates demonstrated an inducible resistance phenotype. A higher percentage of GAS isolates exhibited inducible resistance in comparison to the other BHS isolates.

Table 1. Sample wise distribution of BHS isolates.

BHS(n)	Pus	Cervical Swab	Urine	Throat Swab	Blood	CSF
GAS (27)	23 (85%)	0	0	1(3.7%)	3(11%)	0
GBS (77)	28 (36.4%)	13(16.8%)	31(40%)	1(1.3%)	3(3.9%)	1(1.3%)
GGs (23)	20 (86.9%)	0	0	2(8.7%)	1(4.3%)	0
GFS (1)	0	0	0	0	1(100%)	0
S.porcinus(1)	0	0	1(100%)	0	0	0
Total (129)	71(55.03%)	13(10.07%)	32(24.8%)	4(3.1%)	8(6.2%)	1(0.8%)

Table 2. Antibiotic resistance profile of BHS isolates.

BHS (n)	Erythromycin	Clindamycin	Quinupristin	Tetracycline
GAS (27)	9 (33.3%)	6(22%)	6(22%)	11(40%)
GBS (77)	13(16.8%)	9(12%)	11(14%)	43(56%)
GGs (23)	3(13%)	1(4%)	3(13%)	11(48%)
GFS (1)	0	0	0	0
<i>S.porcinus</i> (1)	1(100%)	1(100%)	1(100%)	1(100%)
Total (129)	26(20.15%)	17(12.14%)	21(16.3%)	66(51.2%)

Table 3. Macrolide resistant phenotypes in BHS isolates.

Table 3 Macrolide resistant phenotypes in BHS isolates				
Macrolide resistant BHS (n=26)	Inducible (n=12)	Constitutive (n=5)	M phenotype(n=9)	
GAS (9)	6 (66.6%)	0	3 (33.3%)	
GBS (13)	5 (38.5%)	4 (30.7%)	4 (30.7%)	
GGG (3)	1 (33.3%)	0	2 (66.6%)	
<i>S. porcinus</i> (1)	0	1 (100%)	0	

Table 4. Distribution of macrolide resistance genes among BHS isolates.

Macrolide Resistant BHS (n)	<i>erm(A)</i> (n=7)	<i>erm(B)</i> (n=10)	<i>mef(A)</i> (n=9)
GAS (9)	0	6 (66.6%)	3 (33.3%)
GBS (13)	6 (46.2%)	3 (23%)	4 (30.8%)
GGG (3)	1 (33.3%)	0	2 (66.6%)
<i>S.porcinus</i> (1)	0	1 (100%)	0

Table 5. Genotype distribution among different phenotypes of macrolide resistant BHS.

Macrolide resistant BHS (n=26)	iMLSb (n=12)		cMLSb (n=5)		Mphenotype (n=9)
	<i>erm(A)</i>	<i>erm(B)</i>	<i>erm(A)</i>	<i>erm(B)</i>	<i>Mef(A)</i>
GAS (9)	0	6 (66.6%)	0	0	3 (33.3%)
GBS (13)	5 (38.46%)	0	1 (7.7%)	3 (23%)	4 (30.8%)
GGG (3)	1 (33.3%)	0	0	0	2 (66.6%)
<i>S.porcinus</i> (1)	0	0	0	1 (100%)	0

Similar to previous studies conducted in India, there was a higher prevalence of inducible resistance (8, 12). The iMLS_B phenotype was also found to be predominant in studies conducted in Norway and Bulgaria (13). These findings highlight the importance of detecting inducible resistance in routine antimicrobial susceptibility testing to prevent treatment failure due to false susceptibility reporting.

Resistance to macrolides in the *Streptococcus* genus is mainly caused by modification of the drug target site through methyltransferases encoded by *erm* genes or active efflux via an efflux pump encoded by the *mef* gene. Among these genes, *erm(A)* is expressed in an inducible manner, while *erm(B)* can be expressed constitutively (cMLS_B) or inducibly (iMLS_B) and leads to high-level resistance (14). This information suggests that different genes and modes of resistance can lead to macrolide resistance in *Streptococcus*, which is important to consider when testing for antibiotic susceptibility.

The macrolide-resistant isolates exhibited a diverse distribution of resistance genes, as revealed by molecular characterization. In GAS isolates, the *erm(B)* gene was the most common genotype detected, followed by *mef(A)*, whereas in GBS isolates, the *erm(A)* gene was the most frequently detected genotype. In our study six GBS isolates and one GGS isolate were found to carry the *erm(A)* gene. This contrasts with previous studies conducted in India where none of the BHS isolates were found to carry the *erm(A)* gene [8,12]. However, Balaji et al. from India and Michos et al. from Greece have reported the predominance of the *erm(A)* gene in GAS isolates [15,16]. Inducible resistance in GAS isolates was exclusively associated with the *erm(B)* gene, while it was linked to the *erm(A)* gene in GBS and GGS isolates. The *erm(B)* gene was the predominant genotype detected among isolates with a constitutive resistance phenotype. In line with previous studies conducted in India and other parts of the world, all isolates with the M phenotype harboured the *mef(A)* gene. These findings suggest a diverse distribution

of macrolide-resistant genes across different *Streptococcus* species.

Conclusion

Our study was one of the first study from Kerala, reporting the antimicrobial resistance in BHS isolates. This study was done just before the beginning of the COVID 19 pandemic and macrolide resistance was found to be relatively low at the time of the study. However, macrolides are commonly misused antibiotics, particularly during the COVID-19 pandemic, and have been identified as a major driver of macrolide resistance in streptococci, both at the population and individual levels (17, 18). In this study we did not investigate the impact of macrolide consumption on resistance rates. Therefore, further research on macrolide resistance and its consumption is necessary to develop policies to regulate the misuse of this agent. Our study highlights the importance of routine antibiotic susceptibility testing for beta-haemolytic streptococci, as well as the detection of inducible resistance to prevent therapeutic failure. Additionally, genotypic analysis will provide insights into the variability of strains in different regions of the country and the world.

Acknowledgements

The authors acknowledge the support of Jubilee Centre for Medical Research for their active support in performing molecular part of the study.

Funding Information

No funding information was reported.

Ethics approval and consent to participate

Letter no. 33/18/IEC/JMMC&RI dated 20/12/2018.

Conflict of interest

The authors notified that there are no conflicts of interest.

References

- Williams GS. Group C and G streptococci infections: emerging challenges. *Clin Lab* 2003;**16**(4):209-13.
- Bramhachari PV, Kaul SY, McMillan DJ, et al. Disease burden due to *Streptococcus dysgalactiae* subsp. *equisimilis* (group G and C streptococcus) is higher than that due to *Streptococcus pyogenes* among Mumbai school children. *J Med Microbiol* 2010;**59**(2):220-3.
- Ibrahim SB, El-Sokkary RH, Elhewala AA, et al. Emerging resistance to erythromycin and penicillin among *Streptococcus pyogenes* isolates in Zagazig, Egypt. *Int J Curr Microbiol Appl Sci* 2014; **3**:750-6.
- Capoor MR, Nair D, Deb M, et al. Resistance to erythromycin and rising penicillin MIC in *Streptococcus pyogenes* in India. *Jpn J Infect Dis* 2006; **59**:334-6.
- Bennett JE, Dolin R, Blaser MJ. Mandell, Douglas, and Bennett's principles and practice of infectious diseases: 2-volume set. Elsevier Health Sciences; 2014; **28**.
- CLSI M100-ED30:2020 Performance Standards for Antimicrobial Susceptibility Testing, 30th Edition (2020).
- Miller JM, Binnicker MJ, Campbell S, et al. A guide to utilization of the microbiology laboratory for diagnosis of infectious diseases: 2018 update by the Infectious Diseases Society of America and the American Society for Microbiology. *Clin Infect Dis* 2018;**67**(6):e1-94.
- Bhardwaj N, Mathur P, Behera B, et al. Antimicrobial resistance in beta-haemolytic streptococci in India: A four-year study. *Indian J Med Res* 2018;**147**(1):81.
- Jain A, Shukla VK, Tiwari V, et al. Antibiotic resistance pattern of group-a beta-hemolytic streptococci isolated from North Indian children. *Indian J Med Sci* 2008; **62**:392-6.
- Mathur P, Kapil A, Das B, et al. Invasive beta-haemolytic streptococcal infections in a tertiary care hospital in Northern India. *J Med Microbiol* 2002; **51**:791-2.
- Dhanda V, Chaudhary P, Toor D, et al. Antimicrobial susceptibility pattern of β -haemolytic group A, C and G streptococci isolated from North India. *J Med Microbiol* 2013; **62**:386-93.
- Abraham T, Sistla S. Trends in antimicrobial resistance patterns of Group A streptococci, molecular basis and implications. *Indian J Med Microbiol* 2018;**36**(2):186-91.
- Littauer P, Caugant DA, Sangvik M, et al. Macrolide-resistant *Streptococcus pyogenes* in Norway: population structure and resistance determinants. *Antimicrob Agents Chemother* 2006;**50**(5):1896-9.
- Berbel D, González-Díaz A, López de Egea G, et al. An Overview of Macrolide Resistance in Streptococci: Prevalence, Mobile Elements and Dynamics. *Microorganisms* 2022;**10**(12):2316.
- Michos A, Koutouzi FI, Tsakris A, et al. Molecular analysis of *Streptococcus pyogenes* macrolide resistance of paediatric isolates during a 7-year period (2007–13). *J Antimicrob Chemother* 2016;**71**(8):2113-7.
- Balaji K, Thenmozhi R, Prajna L, et al. Comparative analysis of emm types, superantigen gene profiles and antibiotic resistance genes among *Streptococcus pyogenes* isolates from ocular infections, pharyngitis and asymptomatic children in South India. *Infect Genet Evol* 2013; **19**(105):12.
- Goossens H, Ferech M, Vander Stichele R, et al. Outpatient antibiotic use in Europe and association with resistance: a cross-national database study. *Lancet* 2005; **365**:579–87.
- Malhotra-Kumar S, Lammens C, Coenen S, et al. Effect of azithromycin and clarithromycin therapy on pharyngeal carriage of macrolide-resistant streptococci in healthy volunteers: a randomised, double-blind, placebo-controlled study. *Lancet* 2007; **369**:482-90.