Original Article

Ecology of Malaria Vectors in an Endemic Area, Southeast of Iran

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Abstract

Background: Malaria has long been regarded as one of the most important public health issues in Iran. Although the country is now in the elimination phase, some endemic foci of malaria are still present in the southeastern areas of the country. In some endemic foci, there are no data on the malaria vectors. To fill this gap, the present study was designed to provide basic entomological data on malaria vectors in the southeastern areas of Iran.

Methods: Adult and larval stages of *Anopheles* mosquitoes were collected by using different catch methods. Resistance of the main malaria vector in the study area to selected insecticides was evaluated using diagnostic doses advised by the World Health Organization in 2013–2014.

Results: A total of 3288 larvae and 1055 adult *Anopheles* mosquitoes were collected, and identified as: *Anopheles stephensi* (32.1%), *Anopheles culicifacies* s.l. (23.4%), *Anopheles dthali* (23.2%), *Anopheles superpictus* s.l. (12.7%), and *Anopheles fluviatilis* s.l. (8.6%). *Anopheles stephensi* was the most predominant mosquito species collected indoors at the study area, with two peaks of activity in May and November. This species was found to be resistant to DDT 4%, tolerant to malathion 5% and susceptible to other tested insecticides.

Conclusion: All the five malaria vectors endemic to the south of Iran were collected and identified in the study area. Our findings on the ecology and resting/feeding habitats of these malaria vectors provide information useful for planning vector control program in this malarious area.

Keywords: Malaria; Anopheles; Malaria vectors; Bio-ecology; Iran

Introduction

Malaria has long been regarded as one of the most important public health issues in Iran. The disease caused irreparable financial and fatality losses in the country, which made initiation of elimination necessary. The most important endemic foci in the country are Sistan and Baluchistan, Hormozgan and Kerman Provinces, which in total, account for 96% of all cases. At present, Iran is in the process of eliminating malaria, and under this condition, even low number of reported cases is very important (1).

Adequate understanding of the association between the behavioral characteristics of the disease vectors and their ecology is important in the planning and determination of strategies to fight against the disease. *Anopheles* species has been considered an important part in malaria transmission cycle after its role in the transmission of the disease was discovered. The ability of *Anopheles* mosquitoes to transmit *Plasmodium* infections is attributed to the physiology and biochemistry of their bodies, which are different according to species characteristics. Other factors such as frequency of blood feeding, longevity, ecological and environmental conditions are important in this respect (2). According to the last checklist of mosquitoes of Iran, there are 30 *Anopheles* species (3). Kerman Province has a long history of ma-

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laria, and even though the disease has been well controlled in other endemic areas of the country, malaria is still considered a major health problem in the province. Previous studies have identified nine *Anopheles* species in the province, and among them, *Anopheles fluviatilis* s.l., *Anopheles dthali*, *Anopheles culicifacies* s.l., *Anopheles stephensi* and *Anopheles superpictus* s.l. have been reported as vectors of malaria (4-6).

Residual spraying and use of insecticidetreated bed nets have recently been implemented in Qaleh-Ganj County in Kerman Province. Due to the higher potential of malaria transmission by *Anopheles* vectors, it was necessary to carry out studies on the fauna and ecology of *Anopheles* species in the area and their susceptibility to some of the conventional pesticides. The aim of this study was to collect information about the fauna and bioecology of *Anopheles* mosquitoes in the area, and to determine their susceptibility to some selected insecticides. Our study intended to provide data that can be useful for future vector control programs in the area.

Materials and Methods

Study area

Kerman Province is located in the southeast of the central plateau of Iran (Fig. 1). The province has a mean annual rainfall of 152.9mm, and according to the current national Counties distribution, the province is composed of 16 counties. Qaleh-Ganj is a county located in the south of the province (27.5277° N, 57.8651° E), with a population of about 70,000 people, 17% of the people live in urban and 83% in rural areas. According to the Koppen-Geiger climate classification, Oale-Ganj County is classified under hot desert climate (BWh); however, in recent years due to reduced rainfall, drought has gripped the area. The maximum and minimum recorded temperatures in the district are 52 and 2 °C, respectively. Monthly maximum and minimum relative humidity in the district is 70% and 35%, respectively, and rainfall in the area is between 0–125mm (Statistical Yearbook of Kerman Province, 2015).

Entomological survey

In this study, sample collection was carried out in three villages including: Shah-Kahan (27.519126°N, 57868245°E), Marz (27.537 180°N, 57.852005°E), and Rameshk (27.521 183°N, 57.857334°E), during 2015. Study sites were chosen according to the WHO standard techniques (7). Mosquitoes were collected using the simple sampling method, before the activity season of the vectors. Specimen collection was carried out over a period of 12 months using total catch, pit shelter, light trap, window trap and dipping methods (7) and by using night catch method during the month of May in 2015.

Total catch method was used to determine species richness of each location and monthly changes in species population and diversity, and to determine the physiological status of endophagic and endophilic species. Artificial pit shelters were created for the collection of outdoor adult mosquitoes. A pit was drilled in each study location (village). Cavities with a dimension of 30×30cm were dug in the walls of the pit shelters about 0.5 meters from the floor of the shelter. These attractive cavities served as resting-sites for mosquitoes entering the pit. Mosquitoes were aspirated from the pit shelters using an aspirator, before sunrise (6-9am) in each day of collection. Another method used for mosquito sampling in our study was the light trap method. During the study, a CDC light trap was used, before sunset and until sunrise in the next day, to collect mosquitoes in Ramsehk Village.

To study the feeding and resting behaviors of adult mosquitoes, window exit traps were installed on the outside of window frames in selected locations. Collected females were identified at species level and their abdominal condition was recorded. Night collection method using human and animal (cow) baits was conducted to identify the host preference of *Anoph*- *eles* mosquitoes in the study area. In this method, mosquitoes were collected as soon as they landed on the host using torch and aspirator, from sunset to sunrise. To determine the peak of host seeking and blood feeding activity, samples collected in each hour were kept in individual cups covered by a fine net and labeled based on the place and time of capture.

After adult specimen collection, the mosquito species were mounted on entomological pins and were identified with a morphological key (8). Female mosquitoes collected by the different methods were classified according to their abdominal condition as: gravid (G), semigravid (SG), unfed (U) and/or freshly blood-fed (F) (7).

In the present study, anopheline mosquito larvae were collected using the dipping method (7). Physical characteristics of the larval habitats were recorded during larval collection. Collected larval specimens were preserved in lacto-phenol for least 24h prior to specimen preparation for microscopic identification. The specimens were mounted on glass slides using Chloral-Gum mounting media and were covered with coverslips. The glass slides were then placed in an incubator at 37 °C to dry before observing under a microscope. Identification key was used to identify the mounted samples based on morphological characteristics (8).

Susceptibility test studies

Susceptibility tests were performed using insecticide-impregnated papers on 2 to 3 day old dominant *Anopheles* species fed with 5% sugar. Mosquitoes were exposed to insecticide-impregnated papers at diagnostic doses, as described by the World Health Organization pesticide scheme guidelines, for one hour, and mortality after 24-hour recovery period was recorded. The tests were carried out at a temperature between 22–26 °C and relative humidity of 60%. In the present study, DDT 4%, Malathion 5%, Propoxur 0.1% and Deltamethrin 0.05% were used for susceptibility testing. Mortality rate of 98–100% was considered as susceptible, 90–97.99% as tolerant, and < 90% as resistant (9). For each insecticide, four replicates of 25 2 to 3day old sugarfed female mosquitoes were tested for susceptibility, whilst two replicates were used as controls. In this study: if control mortality was less than five percent, the results of tests were considered acceptable; if control mortality was between 5–20%, the test results were corrected using Abbotts' formula; and if control mortality was > 20%, the tests were repeated (9).

Results

Mosquitoes Collection

A total of 3288 larvae and 1055 adult Anopheles mosquitoes were collected and identified as An. stephensi (32.1%), An. culicifacies s.l. (23.4%), An. dthali (23.2%), An. superpictus s.l. (12.7%) and An. fluviatilis s.l. (8.6%). Results are described, according to the method of collection, as follows:

Total catch

A total of 541 *Anopheles* species were collected indoors using this method and *An. stephensi* (47.31%) was the most predominant species. The abundance of other species collected by this method is as follows: 114 (21.07%) belonged to *An. culicifacies* s.l., 123 (22.73%) belonged to *An. dthali*, 7 (1.32%) belonged to *An. fluviatilis* s.l., and 40 (7.57%) belonged to *An. superpictus* s.l. (Fig. 2). *Anopheles stephensi* was collected throughout the sampling period except in January and February.

Pit shelter

A total of 137 *Anopheles* species were collected from shelter pits, and *An. fluviatilis* s.l. was the most predominant species (35.76%). *Anopheles superpictus* s.l. was also identified by this method (Fig. 3).

Light trap

Three *Anopheles* species were collected by this method, and *An. superpictus* s.l. was the most numerically dominant. Other species col-

lected by this method include *An. culicifacies* s.l. and *An. stephensi* (Table 1).

Window trap

A total of 57 female *Anopheles* mosquitoes representing four species were trapped by window traps. *Anopheles dthali* (33.3%), *An. culicifacies* s.l. (31.6%), *An. stephensi* (26.3%), and *An. superpictus* s.l. (8.8%) were collected and identified using this method.

Night catch using human/animal baits

Table 2 shows the abundance of adult mosquitoes collected by night catch method using human and animal baits during May 2015. Using human baits, the highest collection occurred in the first third of the night, and the species were identified as *An. superpictus* s.l. and *An. fluviatilis* s.l. *An. culicifacies* s.l. and *An. fluviatilis* s.l. were also collected using animal baits, with the highest collection also occurring at the first third of the night. Blood feeding peak of *An. fluviatilis* s.l. and *An. culicifacies* s.l. were 8–9pm and 10–11pm, respectively.

Abdominal status of collected female mosquitoes

Abdominal condition of the female *Anopheles* mosquitoes collected by three different methods was examined. We classified the females as unfed (U), blood fed (F), and gravid/ semigravid (G/SG), according to their abdominal condition (Tables 3–5).

Total catch

The number of gravid and semi gravid *An. stephensi* collected by this method was higher than the number of unfed and blood-fed *An. stephensi.* Unlike *An. stephensi*, the number of gravid and semi gravid *An. culicifacies, An. fluviatilis* s.l., *An. superpictus* s.l. and *An. dthali* was lower than the number of unfed and blooded ones (Table 3).

Pit shelter

The number of gravid and semi gravid *An. culicifacies* s.l. species was higher than the number of unfed and blood-fed *An. culicifacies* s.l. In contrast, the number of gravid and semi gravid *An. stephensi*, *An. fluviatilis* s.l., *An. superpictus* s.l. and *An. dthali* was lower than that of unfed and blood fed ones (Table 4).

Window trap

The number of gravid and semi gravid *An. culicifacies* s.l. and *An. stephensi* was higher than the number of unfed and blood-fed ones; in contrast, the number of gravid and semi gravid *An. fluviatilis* s.l., *An. superpictus* s.l. and *An. dthali* was lower than the number of unfed and blood-fed species (Table 5).

Larval collection

Anopheles stephensi (31.59%) and An. fluviatilis s.l. (7.29%) were respectively the most abundant and least abundant larval species collected in the study area. The results of species abundance per 10 dips have been summarized in Fig. 4. Characteristics of larval habitats in the study area have been detailed in Table 6.

Susceptibility tests

Mortality rate of *An. stephensi* against DDT 4%, malathion, propoxur, and deltamethrin was 25%, 97%, 99% and 98%, respectively. Mortality in the control against all insecticides tested were zero except for malathion and propoxur. *Anopheles stephensi* was quite resistant to DDT, but tolerant to Malathion and sensitive to the remaining insecticides.

Species	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec	Total	%
An. stephensi	0	2	0	0	0	0	0	0	3	1	0	0	6	27.27
An. culicifacies	0	0	0	0	0	0	0	0	0	2	2	0	4	18.18
An. superpictus	0	2	3	1	0	0	0	2	0	1	3	0	12	54.55

Table 2 . The abundance of mosquitoes collected on human/animal baits by night catch method, Qaleh-Ganj County,
Kerman Province of Iran, May 2013

Dait	Species	Cat	Total		
Bait	Species	1 st third	2 nd third	3 rd third	Total
	An. stephensi	7	2	1	10
	An. culicifacies	5	2	2	9
	An. superpictus	9	5	4	18
Human	An. dthali	6	3	1	10
	An. fluviatilis	11	6	2	19
	Total	38	18	10	66
	An. stephensi	4	3	2	9
	An. culicifacies.	15	3	4	22
A	An. superpictus	9	5	4	18
Animal	An. dthali	12	2	1	15
	An. fluviatilis	19	6	4	29
	Total	59	19	15	93

Table 3. Abdominal status of collected female mosquitoes by total catch, in the study area, Qaleh-Ganj County, Ker-
man Province of Iran, 2015

Ti								Spe	ecies/	Abdo	mina	l Stat	us							
Time		An. st	ephen	si	A	n. cul	licifac	ies	A	ı. sup	erpict	us.		An.	dthali		A	n. flı	ıviatil	lis
	n	ы	G+SG	Total	Ŋ	Ы	G+SG	Total	n	Ы	G+SG	Total	n	Ы	G+SG	Total	n	H	G+SG	Total
Mar	6	15	28	49	3	9	7	19	1	3	2	6	6	14	12	32	0	0	0	0
Apr	7	17	44	68	2	14	25	41	2	5	7	14	0	21	15	36	0	3	2	5
May	5	22	14	41	1	8	6	15	0	2	5	7	3	6	2	11	0	2	0	2
Jun	2	6	3	11	0	2	2	4	0	3	0	3	0	2	1	3	0	0	0	0
Jul	0	2	3	5	0	2	0	2	0	1	0	1	1	2	0	3	0	0	0	0
Aug	0	0	2	2	0	0	1	1	0	0	0	0	0	0	1	1	0	0	0	0
Sep	2	2	5	9	1	1	2	4	0	1	2	3	1	4	4	9	0	0	0	0
Oct	9	15	24	48	2	5	8	15	0	0	0	0	3	5	4	12	0	0	0	0
Nov	2	7	5	14	2	6	2	10	0	3	1	4	2	7	3	12	0	0	0	0
Dec	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Jan	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Feb	3	3	3	9	0	2	1	3	0	2	1	3	0	3	1	4	0	0	0	0
Total	36	89	131	256	11	49	54	114	3	20	18	41	16	64	43	123	0	5	2	7

Time								Sp	ecies	/Abdo	omina	l Stat	tus							
	A	n. ste	phen	si	Ar	ı. culi	cifaci	es.	A	n. sup	erpict	us		An. c	lthali		A	n. flu	viatil	is
	U	Ч	G+SG	Total	U	Ч	G+SG	Total	U	F	G+SG	Total	U	F	G+SG	Total	U	F	G+SG	Total
Mar	0	3	1	4	0	1	1	2	1	1	3	5	0	0	0	0	1	4	2	7
Apr	1	2	2	5	0	1	2	3	1	4	4	9	0	2	1	3	2	7	3	12
May	0	1	1	2	0	1	2	3	1	2	1	4	1	2	0	3	0	3	3	6
Jun	0	1	2	3	0	1	0	1	1	1	0	2	1	0	1	2	1	1	1	3
Jul	0	1	0	1	0	2	1	3	0	0	2	2	0	1	0	1	0	0	0	0
Aug	1	1	0	2	0	0	1	1	0	0	0	0	0	0	0	0	0	0	0	0
Sep	1	1	1	3	0	0	1	1	0	2	1	3	0	2	1	3	0	2	1	3
Oct	0	3	2	5	0	0	0	0	0	0	1	1	0	0	1	1	1	2	3	6
Nov	2	0	1	3	0	1	1	2	0	0	0	0	0	0	0	0	2	2	2	6
Dec	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	1	1	2
Jan	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Feb	0	2	0	2	0	1	0	1	0	0	0	0	0	1	1	2	0	2	2	4
Total	5	15	10	30	0	8	9	17	4	10	12	26	2	8	5	15	7	24	18	49

Table 4. Abdominal states of collected female mosquitoes by shelter pit, in the study area, Qaleh-Ganj County, Ker-
man Province of Iran, 2015

 Table 5. Abdominal status of collected female mosquitoes by outdoor window trap, in the study area, Qaleh-Ganj

 County, Kerman Province of Iran, 2015

Time						Sp	ecies/	Abdo	mina	l Stat	us					
	A	n. ste	phens	ri 🛛	An	ı. culi	cifaci	es	An	. supe	erpicti	us		An. d	thali	
	Ŋ	ί τ ι	G+SG	Total	Ŋ	ί Ξ ι	G+SG	Total	Ŋ	Ĩ	G+SG	Total	Ŋ	ы	G+SG	Total
Mar	0	0	2	2	0	0	1	1	0	0	0	0	0	1	1	2
Apr	0	0	1	1	0	1	2	3	0	0	1	1	0	0	2	2
May	0	0	1	1	0	0	1	1	0	0	0	0	0	0	1	1
Jun	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Jul	0	0	0	0	0	0	1	1	0	0	0	0	0	0	0	0
Aug	0	1	1	2	0	0	0	0	0	0	0	0	0	0	0	0
Sep	0	0	0	0	0	1	1	2	0	0	0	0	0	0	0	0
Oct	0	1	1	2	0	1	3	4	1	1	0	2	0	2	1	3
Nov	1	1	2	4	0	1	1	2	0	0	0	0	0	3	1	4
Dec	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0	0
Jan	0	0	0	0	0	0	0	0	0	0	0	0	0	2	1	3
Feb	0	1	2	3	1	1	2	4	0	1	1	2	0	3	1	4
Total	1	4	10	15	1	5	12	18	1	2	2	5	0	11	8	19

Table 6. Characteristics of larval habitats in the study area, Qaleh-Ganj County, Kerman Province of Iran, 2015

	Categories	An. stephensi	An. culic- ifacies	An. super- pictus	An. dthali	An. flu- viatilis
	Constant	64	75	67	64	72
TT 1. (C) ()	Temporary	36	25	33	36	28
Habitat Situation	Running Water	100	100	100	100	100
	Resident Water	0	0	0	0	0
Vegetation Situation	Without Vegetation	64	75	67	64	72
Vegetation Situation	With Vegetation	36	25	33	36	28

	Muddy	36.4	25	33	36	28
Туре	Sand	27.6	38	23	26	30
	Stone or Cement	36	37	46	38	42
	Turbid	64	75	67	64	72
Water Quality	Fresh	36	25	33	36	28
	Sunny	27.5	38	23	26	30
Sunlight Situation	Semi-Shade	72.5	62	77	74	70
-	Shade	0	0	0	0	0
Habitat	Natural	64	75	67	64	72
парна	Artificial	36	25	33	36	28
Temperature (°C)	Mean	29.5	28	26.5	31.5	22
рН	Mean	7.6	7.6	7.6	7.6	7.6

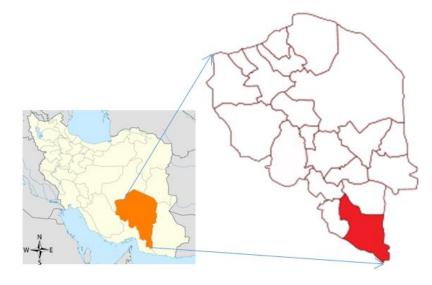


Fig. 1. Study area in Kerman Province of Iran

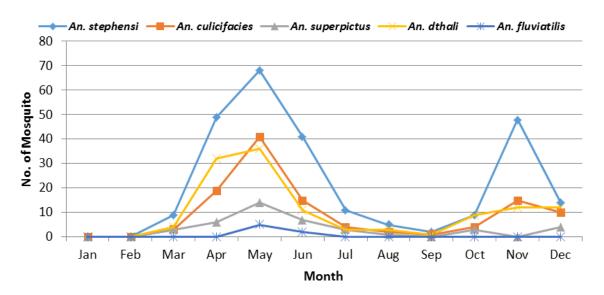


Fig. 2. The abundance of mosquitoes collected from indoors by total catch, in the study area, Qaleh-Ganj County, Kerman Province of Iran, 2015

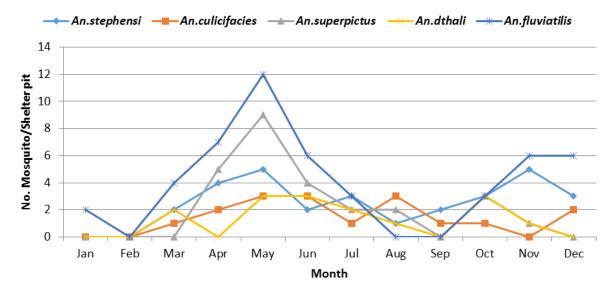


Fig. 3. The abundance of mosquitoes collected by shelter pit method in the study area, Qaleh-Ganj County, Kerman Province of Iran, 2015

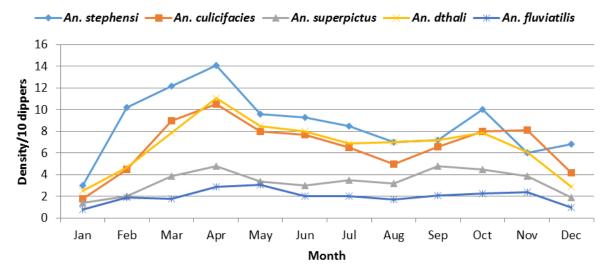


Fig. 4. Larval abundance per 10 dips in the study area, Qaleh-Ganj County, Kerman Province of Iran, 2015

Discussion

Anopheles stephensi

Among the 541 Anopheles species collected indoors by the total catch method, An. stephensi was the most predominant species. It was highly abundant in May. In contrast to our study, An. stephensi was sampled throughout the year in another study in southeastern Iran, with the peak of activity occurring in February and September–October (10). In another study, the peak of *An. stephensi* activity was found in May and November in Bandar Abbas in southern Iran (11). However, a recent study conducted in Jask County, south of our study area, reported two peaks of activity for *An. stephensi* in March–April and October (12). The peak of activity of *An. stephensi* is directly affected by weather variables especially temperature, which varies between different study areas.

The density of Anopheles stephensi collected in shelter pits was low. A similar result was reported in the southern Iran (10). With the night catch method, the highest number of An. stephensi mosquitoes was collected in the first third of the night between 9–10pm. In a study conducted at Khesht area in Fars Province in Iran, similar results were obtained (13). It should be mentioned that this method was only performed during the month of May. We recommend this method to be used at least in all months of the malaria transmission season in future studies in the study area. In this study, the number of gravid and semi gravid An. stephensi collected by shelter pit method was lower than the number of unfed and blood-fed ones, but in total catch method, the gravid and semi gravid ratio was more than 1. This shows that An. stephensi has a higher endophilic tendency. In a similar study conducted in Sistan and Baluchistan and Hormozgan Provinces of Iran, An. stephensi was found to be the most numerically dominant among the sampled mosquito species. Moreover, the number of gravid and semi gravid An. stephensi collected in the study, both in shelter pits and indoors, was higher than the number of unfed and bloodfed species. The investigators indicated that An. stephensi is more endophilic compared with the other species sampled in the study areas: with G+SG/F+UF ratio lower than one both outdoors and indoors (18)

Anopheles culicifacies s.l.

In our study, the number of gravid and semi gravid *An. culicifacies* s.l. collected by total catch method was lower than unfed and bloodfed ones, but in the pit shelters and outdoor window trap methods, the gravid and semi gravid ratio was more than 1. Moreover, the high number of unfed mosquitoes compared with gravid and blood-fed mosquitoes collected by indoor window trap method shows a high endophagic tendency of this species. In contrast

to our findings, in a study conducted in Sistan and Baluchistan, the number of gravid and semi gravid An. culicifacies s.l. collected indoors was lower than the number of unfed and bloodfed mosquitoes. The researchers also reported the same abdominal state findings for An. culicifacies s.l. mosquitoes collected from pit shelters (14). Another study conducted in Sistan and Baluchistan indicated that this Anopheles species is more endophilic (15, 16). Consistent with our study, based on ventral aspect ratio (G+SG/U+F), another study stated that An. culicifacies s.l. prefers both indoors and outdoors as their resting places (10). Similar results have also been reported in different regions of India (17).

Although this species had two peaks of activity during March-April and October-November, a study conducted in Sistan and Baluchistan in Iran, larvae of An. culicifacies s.l. were more abundant in April to December in rice fields with palm trees (18). In terms of breeding sites, this species was mostly collected from sites which have turbid water. semi-shade and have no vegetation. The mean temperature and pH of the study area were 28 °C and 7.6, respectively. An earlier study conducted in the south of our study area reported clear, no vegetation, sunny, and natural breeding sites with average temperature and pH of 25-30 °C and 7.14-8.90, respectively as climate preferences of this species (19).

Anopheles dthali

This species was most abundant in May, June and October during the study period. A previous study conducted in a relatively warmer area in Iran reported that the peak of activity of this species occurs in April and September–October, which is one month earlier than observed in our study (20). It seems that weather conditions play crucial role in the period of activity of this species, making it necessary to take into consideration the weather condition of the area before planning any vector control measures. The highest abundance of this species, using night catch method with human and animal baits, occurred in the first third of the night between 8–9pm. We thus recommend that people be encouraged to use bed nets or avoid outdoors at these times.

Larvae of An. dthali were mostly found in stagnant, turbid waters, natural breeding sites in river banks which have no vegetation, and semi-shade. In agreement with our results, this Anopheles species was sampled from breeding sites without vegetation (19), but unlike our findings, it was mostly collected from clear and sunny sites. Although a previous study reported that breeding places with temperature ranging between 13 °C and 28 °C and pH between 6.9 and 8.0 are preferred by this species (21), the mean temperature and pH of our study sites were 31.5% and 7.6, respectively. Another study also reported that about 50% of An. dthali larvae were collected from breeding sites with temperature ranging between 25.1-30 °C and pH between 7.14 and 8.20 (19).

Anopheles fluviatilis s.l.

This Anopheles species is also considered as a secondary vector of malaria in most of its' distribution areas in Iran (22). In some studies, it was captured in outdoor habitats and on animal baits (23). We found An. fluviatilis s.l. was the most predominant species (35.76%) caught by pit shelter method, confirms its exophilic habit. It was also collected by night catch method using human and animal baits in May and November, mostly in the first third of the night between 9–10pm. The number of unfed and blood-fed An. fluviatilis s.l. was higher than the number of gravid and semi gravid An. fluviatilis s.l. captured by total catch method, which shows high exophilic tendency of this species.

Breeding sites for this species in our study area were natural water bodies, stagnant, turbid waters, and semi-arid areas without vegetation. Mean temperature and pH of the study sites were 22 °C and 7.6, respectively. This species usually breeds in fresh, slow flowing or even stagnant waters (22).

Anopheles superpictus s.l.

The highest collection of this species occurred in the first third of the night between 8 to 9pm. In our study, the number of unfed and blood-fed *An. superpictus* s.l. was lower than the number of gravid and semi gravid *An. superpictus* s.l. captured by total catch method, which shows high exophilic tendency of this species. Contrary to our results, a study conducted in south west of Iran, reported endophilic habit of this species in agreement with earlier studies (24).

Studies on the larval habitats of *An. superpictus* s.l. in Iran revealed that this species is frequently abundant in river banks, both in natural breeding sites and artificial habitats created by human activities like mining pools (22). We collected this species from natural breeding sites, stagnant, turbid waters and semi-arid areas with no vegetation. Mean temperature and pH of the collection sites of this species were 26.5 °C and 7.6, respectively. An earlier study reported clear, sandy bed, full sunlight and natural water bodies as the main breeding sites for *An. superpictus* s.l. (19).

Insecticide resistance

Our results show that Anopheles stephensi is quite resistant to DDT, tolerant to malathion and susceptible to deltamethrin and Propoxur. This result is in accordance with that of a study conducted in Chabahar (25), in which An. stephensi was found to be resistant to DDT but tolerant to malathion. Resistance of this species to DDT in Iran has been reported in the last two decades (11, 13, 26-30). Fortunately, in our study, this species was found to be susceptible to both deltamethrin and propoxur insecticides. Thus, these insecticides can be used in indoor residual spraying for vector control against this main endophilic malaria vector. Furthermore, it is recommended that more susceptibility tests be conducted on other species with high endophilic and endophagic tendencies, especially An. culicifacies s.l. and An. dthali, in the area.

Conclusion

In conclusion, five malaria vectors are active in the study area. Regular entomological studies are recommended to monitor their monthly activity and susceptibility status to insecticides. The peak of host seeking activity of the indoor mosquitoes occur in the first 3rd of the night, therefore, community-based training programs should be designed and implemented to enforce proper use of mosquito nets and personal protective measures against mosquitoes.

Country has a long history of work on malaria and publication of several papers on different aspects of malaria including insecticide resistance monitoring, sibling species, molecular study, new record, novel methods for vector control, faunestic study, use of plants for larval control, using bednets and long lasting impregnated nets, morphological studies, malaria epidemiology, ecology of malaria vectors, biodiversity, community participation, vector control, repellent evaluation, anthropophilic index of malaria vectors, training is designated as malaria training center by WHO. There are several reports on different aspects of malaria vectors recently (31-118).

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References

- 1. World Health Organization (2016) World Malaria Repot. Geneva, Switzerland.
- 2. Renshaw M, Silver JB (2001) Malaria, human. Encyclopedia of arthropod-trans-

mitted infections of man and domesticated animals. CABI Publishing. New York, pp. 314–327.

- Azari-Hamidian S, Norouzia B, Harbach RE (2019) A detailed review of the mosquitoes (Diptera: Culicidae) of Iran and their medical and veterinary importance. Acta Trop. 194: 106–122
- Saebi ME (1987) Morphological study on anopheline larvae and their distribution in Iran [PhD dissertation]. Tehran, Iran: School of Public Health, Tehran University of Medical Sciences (In Persian).
- 5. Mehravaran A, Vatandoost H, Abai MR, Javadian E, Edalat H, Grouhi A, Mohtarami F, Oshaghi MA (2011) Species identification of the *Anopheles fluviatilis* complex using PCR-sequencing and phylogenetic analysis in Jiroft district, Kerman province. Zahedan J Res Med Sci. 13(1): 4–9.
- Baseri HR, Moussakazemi SH, Yosafi S, Mohebali M, Hajaran H, Jedari M (2005) Anthropophily of malaria vectors in Kahnouj District, south of Kerman, Iran. Iranian J Publ Health. 34(2): 27–35.
- 7. World Health Organization (2013) Malaria entomology and vector control. WHO, Geneva, Switzerland.
- Azari-Hamidian S, Harbach RE (2009) Keys to the adult females and fourth-instar larvae of the mosquitoes of Iran (Diptera: Culicidae). Zootaxa. 2078(1): 1–33.
- World Health Organization (2016) Test procedures for insecticide resistance monitoring in malaria vector mosquitoes. WHO, Geneva, Switzerland.
- Basseri HR, Raeisi A, Ranjbar Khakha M, Pakarai A, Hassanzehi A (2010) Seasonal abundance and host-feeding patterns of Anopheline vectors in malaria endemic area of Iran. J Parasitol Res. 2010: 1– 8.
- 11. Vatandoost H, Oshaghi MA, Abai MR, Shahi

M, Yaghoobi F, Baghaii M, Hanafi-Bojd AA, Zamani G, Townson H (2006) Bionomics of *Anopheles stephensi* Liston in the malarious area of Hormozgan Province, southern Iran. Acta Trop. 97: 196– 205.

- Yeryan M, Basseri HR, Hanafi-Bojd AA, Raeisi A, Edalat H, Safari R (2016) Bioecology of malaria vectors in an endemic area, Southeast of Iran. Asian Pac J Trop Med. 9(1): 32–38.
- Manouchehri AV, Javadian E, Eshghy N, Motabar M (1976) Ecology of *Anopheles stephensi* Liston in southern Iran. Trop Geog Med. 28(3): 228–232.
- 14. Vatandoost H, Emami SN, Oshaghi MA, Abai MR, Raeisi A, Piazzak N, Akbarzadeh K, Sartipi M (2011) Ecology of malaria vector *Anopheles culicifacies* in a malarious area of Sistan va Baluchestan Province, south-east Islamic Republic of Iran. East Mediterr Health J. 17(5): 439–445.
- 15. Zaim M, Javaherian Z (1991) Occurrence of *Anopheles culicifacies* species A in Iran. J Am Mosq Control Assoc. 7(2): 324–326.
- 16. Zaim M, Zahirnia AH, Manouchehri AV (1993) Survival rates of Anopheles culicifacies s.l. and Anopheles pulcherrimus in sprayed and unsprayed villages in Ghassreghand district, Baluchistan, Iran, 1991. J Am Mosq Control Assoc. 9: 421–425.
- 17. Dash AP, Adak T, Raghavendra K, Singh OP (2007) The biology and control of malaria vectors in India. Current Sci. 92 (11): 1571–1578.
- Zahirnia AH, Taherkhani H, Vatandoost H (2001) Observation of malaria sporozoite in *Anopheles culicifacies* (Diptera: Culicidae) in Ghasreghand District, Sistan and Baluchistan Province. Hakim Res J. 4: 149–153.
- 19. Hanafi-Bojd AA, Vatandoost H, Oshaghi MA, Charrahy Z, Haghdoost AA, Seda-

ghat MM, Abedi F, Soltani M, Raeisi A (2012) Larval habitats and biodiversity of Anopheline mosquitoes (Diptera Culicidae) in a malarious area of southern Iran. J Vector-Borne Dis. 49: 91–100.

- Vatandoost H, Shahi M, Hanafi-Bojd AA, Abai MR, Oshaghi MA, Rafii F (2007) Ecology of *Anopheles dthali* Patton in Bandar Abbas. Iran J Arthropod Borne Dis. 1: 21–27.
- 21. Vatandoost H, Shahi H, Abai MR, Hanafi-Bojd AA, Oshaghi MA, Zamani G (2004) Larval habitats of main malaria vectors in Hormozgan Province and their susceptibility to different larvicides. Southeast Asian J Trop Med Publ Hlth. 35: 22–25.
- Hanafi-Bojd AA, Azari-Hamidian S, Vatandoost H, Charrahy Z (2011) Spatio-temporal distribution of malaria vectors (Diptera: Culicidae) across different climatic zones of Iran. Asian Pac J Trop Med. 4(6): 498–504.
- Naddaf SR, Oshaghi MA, Vatandoost H, Asmar M (2003) Molecular characterization of the *Anopheles fluviatilis* species complex in Iran. East Mediterr Health J. 9: 257–265.
- 24. Maghsoodi N, Ladonni H, Basseri HR (2015) Species composition and seasonal activities of malaria vectors in an area at reintroduction prevention stage, Khuzestan, south-western Iran. J Arthropod Borne Dis. 9(1):60–70.
- 25. Fathian M, Vatandoost H, Moosa-Kazemi SH, Raeisi A, Yaghoobi-Ershadi MR, Oshaghi MA, Sedaghat MM (2015) Susceptibility of Culicidae mosquitoes to some insecticides recommended by WHO in southeastern Iran. J Arthropod Borne Dis. 9(1): 22–34.
- 26. Davari B, Vatandoost H, Oshaghi MA, Ladonni H, Enayati AA, Shayeghi M, Basseri HR, Rassi Y, Hanafi-Bojd AA (2007) Selection of *Anopheles* stephensi with DDT and dieldrin and cross-re-

sistance spectrum to pyrethroids and fipronil. Pest Biochem Physiol. 89: 97–103.

- 27. Gorouhi MA, Vatandoost H, Oshaghi MA, Raeisi A, Enayati AA, Mirhendi H (2016) Current susceptibility status of *Anopheles stephensi* (Diptera: Culicidae) to different imagicides in a malarious area, Southeastern of Iran. J Arthropod Borne Dis. 10(4): 493–500.
- 28. Hanafi-Bojd AA, Vatandoost H, Oshaghi MA, Haghdoost AA, Shahi M, Sedaghat MM, Abedi F, Yeryan M, Pakari A (2012) Entomological and epidemiological attributes for malaria transmission and implementation of vector control in southern Iran. Acta Trop. 121(2): 85–92.
- Vatandoost H, Hanafi-Bojd AA (2012) Indication of pyrethroid resistance in the main malaria vector, *Anopheles stephensi* from Iran. Asian Pac J Trop Med. 5(9): 722–726.
- 30. Abbasi M, Hanafi-Bojd AA, Yaghoobi-Ershadi MR, Vatandoost H, Oshaghi MA, Hazratian T, Sedaghat MM, Fekri S, Safari R, Mojahedi AR, Salari Y (2019) Resistance status of main malaria vector, *Anopheles stephensi* Liston (Diptera: Culicidae) to insecticides in a malaria Endemic Area, Southern Iran. Asian Pac J Trop Med. 12(1): 43–48.
- 31. Vatandoost H, Shahi H, Abai MR, Hanafi-Bojd AA, Oshaghi MA, Zamani G (2004) Larval habitats of main malaria vectors in Hormozgan Province and their susceptibility to different larvicides. Southeast Asian J Trop Med Public Health. 35: 22–25.
- 32. Vatandoost H, Mashayekhi M, Abai MR, Aflatoonian MR, Hanafi-Bojd AA, Sharifi I (2005) Monitoring of insecticides resistance in main malaria vectors in a malarious area of Kahnooj District, Kerman Province, southeastern Iran. J Vector Borne Dis. 42(3): 100–108.
- 33. Davari B, Vatandoost H, Ladonni H, Shaeghi

M, Oshaghi M, Basseri H (2006) Comparative efficacy of different imagicides against different strains of *Anopheles stephensi* in the malarious areas of Iran, 2004–2005. Pak J Biol Sci. 9(5): 885– 892.

- Hanafi-Bojd AA, Vatandoost H, Jafari R (2006) Susceptibility status of *Anopheles dthali* and *An. fluviatilis* to commonly used larvicides in an endemic focus of malaria, southern Iran. J Vector Borne Dis. 43(1): 34–38.
- 35. Abai MR, Mehravaran A, Vatandoost H, Oshaghi MA, Javadian E, Mashayekhi M (2008) Comparative performance of imagicides on *Anopheles stephensi*, main malaria vector in a malarious area, southern Iran. J Vector Borne Dis. 45(4): 307– 312.
- Vatandoost H, Zahirnia AH (2010) Responsiveness of *Anopheles maculipennis* to different imagicides during resurgent malaria. Asian Pac J Trop Med. 3(5): 360–363.
- Vatandoost H, Hanafi-Bojd AA (2012) Indication of pyrethroid resistance in the main malaria vector, *Anopheles stephensi* from Iran. Asian Pac J Trop Med. 5 (9): 722–726.
- Soltani A, Vatandoost H, Oshaghi MA, Enayati AA, Raeisi A, Eshraghian MR (2013) Baseline susceptibility of different geographical strains of *Anopheles stephensi* (Diptera: Culicidae) to Temephos in malarious Areas of Iran. J Arthropod Borne Dis. 7(1): 56–65.
- 39. Lak SS, Vatandoost H, Entezarmahdi M, Ashraf H, Abai M, Nazari M (2002) Monitoring of insecticide resistance in Anopheles sacharovi (Favre, 1903) in borderline of Iran, Armenia, Naxcivan and Turkey, 2001. Iran J Public Health. 31 (3–4): 96–99.
- 40. Enayati AA, Vatandoost H, Ladonni H, Townson H, Hemingway J (2003) Molecular evidence for a kdr-like pyrethroid

resistance mechanism in the malaria vector mosquito *Anopheles stephensi*. Med Vet Entomol. 17(2): 138–144.

- Naddaf SR, Oshaghi MA, Vatandoost H, Assmar M (2003) Molecular characterization of *Anopheles fluviatilis* species complex in the Islamic Republic of Iran. Eastern Mediterr Health J. 9(3): 257– 265.
- 42. Oshaghi MA, Sedaghat MM, Vatandoost H (2003) Molecular characterization of the *Anopheles maculipennis* complex in the Islamic Republic of Iran. Eastern Mediterr Health J. 9(4): 659–666.
- 43. Sedaghat MM, Linton YM, Oshaghi MA, Vatandoost H, Harbach RE (2003) The *Anopheles maculipennis* complex (Diptera: Culicidae) in Iran: molecular characterization and recognition of a new species. Bull Entomol Res. 93(6): 527–535.
- 44. Azari-Hamidian S, Abai MR, Ladonni H, Vatandoost H, Akbarzadeh (2006) *Anopheles peditaeniatus* (Leicester) new to the Iranian mosquito fauna with notes on *Anopheles hyrcanus* group in Iran. J Am Mosq Control Assoc. 22(1): 144–146.
- 45. Oshaghi MA, Shemshad K, Yaghobi-Ershadi MR, Pedram M, Vatandoost H, Abai MR (2007) Genetic structure of the malaria vector *Anopheles superpictus* in Iran using mitochondrial cytochrome oxidase (COI and COII) and morphologic markers: a new species complex? Acta Trop. 101(3): 241–248.
- 46. Naddaf SR, Oshaghi MA, Vatandoost H (2012) Confirmation of two sibling species among *Anopheles fluviatilis* mosquitoes in south and southeastern iran by analysis of Cytochrome Oxidase I Gene. J Arthropod Borne Dis. 6(2): 144–150.
- 47. Dezfouli SR, Oshaghi MA, Vatandoost H, Assmar M (2003) rDNA-ITS2 based species diagnostic polymerase chain reaction assay for identification of sibling species of *Anopheles fluviatilis* in Iran. Southeast Asian J Trop Med Public Health.

34(2): 56–60.

- 48. Soltani A, Vatandoost H, Jabbari H, Mesdaghinia A, Mahvi A, Younesian M (2008) Use of expanded polystyrene (EPS) and shredded waste polystyrene (SWAP) beads for control of mosquitoes. J Arthropod Borne Dis. 2(2): 12–20.
- Omrani SM, Vatandoost H, Oshaghi MA, Shokri F, Guerin PM, Yaghoobi-Ershadi MR (2010) Fabrication of an olfactometer for mosquito behavioral studies. J Vector Borne Dis. 47(1): 17–25.
- 50. Omrani SM, Vatandoost H, Oshaghi M, Shokri F, Yaghoobi-Ershadi M, Rassi Y (2010) Differential responses of *Anopheles stephensi* (Diptera: Culicidae) to skin emanations of a man, a cow, and a guinea pig in the olfactometer. Iran J Arthropod Borne Dis. 4(1): 1–16.
- 51. Omrani SM, Vatandoost H, Oshaghi MA, Rahimi A (2012) Upwind responses of *Anopheles stephensi* to carbon dioxide and L-lactic acid: an olfactometer study. Eastern Mediterr Health J. 18(11): 1134– 1142.
- 52. Chavshin AR, Oshaghi MA, Vatandoost H, Pourmand MR, Raeisi A, Enayati AA (2012) Identification of bacterial microflora in the midgut of the larvae and adult of wild caught *Anopheles stephensi*: a step toward finding suitable paratransgenesis candidates. Acta Trop. 121 (2): 129–134.
- 53. Soltani A, Vatandoost H, Jabbari H, Mesdaghinia AR, Mahvi AH, Younesian M (2012) Field efficacy of expanded polystyrene and shredded waste polystyrene beads for mosquito control in artificial pools and field trials, Islamic Republic of Iran. Eastern Mediterr Health J. 18(10): 1042–1048.
- 54. Moosa-Kazemi S, Vatandoost H, Nikookar H, Fathian M (2009) Culicinae (Diptera: culicidae) mosquitoes in Chabahar County, Sistan and Baluchistan Province, south-

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eastern Iran. Iran J Arthropod Borne Dis. 3(1): 29–35.

- 55. Oshaghi MA, Vatandoost H, Gorouhi A, Abai MR, Madjidpour A, Arshi S (2011) Anopheline species composition in borderline of Iran-Azerbaijan. Acta Trop. 119(1): 44–49.
- 56. Hadjiakhoondi A, Aghel N, Zamanizadeh-Nadgar N, Vatandoost H (2008) Chemical and biological study of *Mentha spicatal* essential oil from Iran. DARU J Pharmaceut Sci. 8(1–2): 19–21.
- 57. Hadjiakhoondi A, Vatandoost H, Jamshidi A, Amiri EB (2003) Chemical Constituents of Efficacy of *Cymbopogon Olivieri* (Boiss) bar essential oil against malaria vector, *Anopheles stepensi*. DARU J Pharmaceut Sci. 11(3): 125–128.
- 58. Oshaghi M, Ghalandari R, Vatandoost H, Shayeghi M, Kamali-Nejad M, Tourabi-Khaledi H (2003) Repellent effect of extracts and essential oils of *Citrus limon* (Rutaceae) and *Melissa officinalis* (Labiatae) against main malaria vector, *Anopheles stephensi* (Diptera: Culicidae). Iran J Public Health. 32(4): 47–52.
- 59. Vatandoost H, Vaziri VM (2004) Larvicidal activity of a neem tree extract (Neemarin) against mosquito larvae in the Islamic Republic of Iran. Eastern Mediterr Health J. 10(4–5): 573–581.
- 60. Hadjiakhoondi A, Vatandoost H, Khanavi M, Abai MR, Karami M (2005) Biochemical investigation of different extracts and larvicidal activity of *Tagetes minuta* L. on *Anopheles stephensi* larvae. Iran J Pharmaceut Sci. 1(2): 81–84.
- 61. Hadjiakhoondi A, Vatandoost H, Khanavi M, Sadeghipour Roodsari HR, Vosoughi M, Kazemi M (2006) Fatty acid composition and toxicity of *Melia azedarach* L. fruits against malaria vector *Anopheles stephensi*. Iran J Pharmaceut Sci. 2 (2): 97–102.
- 62. Sadat Ebrahimi S, Hadjiakhoondi A, Rezazadeh S, Fereidunian N, Vatandoost

H, Abai M (2005) The components of *Tagetes minuta* L. and its biological activities against malaria vector, *Anopheles stephensi* in Iran. J Med Plants Res. 4(16): 43–47.

- 63. Shahi M, Hanafi-Bojd AA, Iranshahi M, Vatandoost H, Hanafi-Bojd MY (2010) Larvicidal efficacy of latex and extract of *Calotropis procera* (Gentianales: Asclepiadaceae) against *Culex quinquefasciatus* and *Anopheles stephensi* (Diptera: Culicidae). J Vector Borne Dis. 47(3): 185–188.
- 64. Khanavi M, Toulabi PB, Abai MR, Sadati N, Hadjiakhoondi F, Hadjiakhoondi A (2011) Larvicidal activity of marine algae, *Sargassum swartzii* and *Chondria dasyphylla*, against malaria vector *Anopheles stephensi*. J Vector Borne Dis. 48(4): 241–244.
- 65. Sedaghat MM, Dehkordi AS, Khanavi M, Abai MR, Mohtarami F, Vatandoost H (2011) Chemical composition and larvicidal activity of essential oil of *Cupressus arizonica* E.L. Greene against malaria vector *Anopheles stephensi* Liston (Diptera: Culicidae). J Pharmacogn Res. 3(2): 135–139.
- 66. Khanavi M, Vatandoost H, Khosravi Dehaghi N, Sanei Dehkordi A, Sedaghat MM, Hadjiakhoondi A (2013) Larvicidal activities of some Iranian native plants against the main malaria vector, Anopheles *stephensi*. Acta Medica Iranica. 51(3): 141–147.
- 67. Vatandoost H, Sanei Dehkordi A, Sadeghi SM, Davari B, Karimian F, Abai MR (2012) Identification of chemical constituents and larvicidal activity of *Kelussia odoratissima* Mozaffarian essential oil against two mosquito vectors *Anopheles stephensi* and *Culex pipiens* (Diptera: Culicidae). Exp Parasitol. 132(4): 470–474.
- 68. Vatandoost H, Dehakia M, Javadian E, Abai MR, Duchson S (2006) Compara-

tive study on the efficacy of lambdacyhalothrin and bifenthrin on torn nets against the malaria vector, *Anopheles stephensi* as assessed by tunnel test method. J Vector Borne Dis. 43(3): 133–135.

- 69. Moosa-Kazemi SH, Vatandoost H, Raeisi A, Akbarzadeh K (2007) Deltamethrin impregnated bed nets in a malaria control program in Chabahar, Southeast Baluchistan, Iran. Iran J Arthropod Borne Dis. 1(1): 43–51.
- 70. Rafinejad J, Vatandoost H, Nikpoor F, Abai MR, Shaeghi M, Duchen S (2008) Effect of washing on the bioefficacy of insecticide-treated nets (ITNs) and longlasting insecticidal nets (LLINs) against main malaria vector *Anopheles stephensi* by three bioassay methods. J Vector Borne Dis. 45(2): 143–150.
- 71. Soleimani Ahmadi M, Vatandoost H, Shaeghi M, Raeisi A, Abedi F, Eshraghian MR (2012) Effects of educational intervention on long-lasting insecticidal nets use in a malarious area, southeast Iran. Acta Medica Iranica. 4: 279–287.
- 72. Soleimani-Ahmadi M, Vatandoost H, Shaeghi M, Raeisi A, Abedi F, Eshraghian MR (2012) Field evaluation of permethrin long-lasting insecticide treated nets (Olyset^{((R)))} for malaria control in an endemic area, southeast of Iran. Acta Trop. 123(3): 146–153.
- 73. Vatandoost H, Mamivandpoor H, Abai MR, Shayeghi M, Rafi F, Raeisi A (2013) Wash Resistance and Bioefficacy of Alpha-cypermethrin Long Lasting Impregnated Nets (LLIN-Interceptor^{((R)))} against *Anopheles stephensi* using Tunnel Test. J Arthropod Borne Dis. 7(1): 31–45.
- 74. Vatandoost H, Ramin E, Rassi Y, Abai M (2009) Stability and wash resistance of local made mosquito bednets and detergents treated with pyrethroids against susceptible strain of malaria vector *Anopheles stephensi*. Iran J Arthropod Borne

Dis. 3(1): 19–28.

- 75. Emami SN, Vatandoost H, Oshaghi MA, Mohtarami F, Javadian E, Raeisi A (2007) Morphological method for sexing anopheline larvae. J Vector Borne Dis. 44(4): 245–249.
- 76. Doosti S, Azari-Hamidian S, Vatandoost H, Hosseini MOM (2006) Taxonomic differentiation of *Anopheles sacharovi* and *An. maculipennis* sl (Diptera: Culicidae) larvae by seta 2 (antepalmate hair). Acta Medica Iranica. 44(1): 21–27.
- 77. Doosti S, Vatandoost H, Oshaghi M, Hosseini M, Sedaghat M (2007) Applying morphometric variation of seta 2 (Antepalmate Hair) among the larvae of the members of the Maculipennis Subgroup (Diptera: Culicidae) in Iran. J Arthropod Borne Dis. 1(1): 28–37.
- 78. Vatandoost H, Ashraf H, Lak SH, Mahdi RE, Abai MR, Nazari M (2003) Factors involved in the re-emergence of malaria in borderline of Iran, Armenia, Azerbaijan and Turkey. The Southeast Asian J Trop Med Public Health. 34(2): 6–14.
- 79. Hanafi-Bojd A, Vatandoost H, Philip E, Stepanova E, Abdi A, Safari R (2010) Malaria situation analysis and stratification in Bandar Abbas County, southern Iran, 2004–2008. Iran J Arthropod Borne Dis. 4(1): 31–41.
- 80. Hanafi-Bojd AA, Vatandoost H, Oshaghi MA, Eshraghian MR, Haghdoost AA, Abedi F (2011) Knowledge, attitudes and practices regarding malaria control in an endemic area of southern Iran. Southeast Asian J Trop Med Public Health. 42(3): 491–501.
- Hemami MR, Sari AA, Raeisi A, Vatandoost H, Majdzadeh R (2013) Malaria elimination Iran, importance and challenges. Int J Prevent Med. 4(1): 88–94.
- 82. Vatandoost H, Shahi M, Hanafi-Bojd A, Abai M, Oshaghi M, Rafii F (2007) Ecology of *Anopheles dthali* Patton in Bandar

- Hanafi-Bojd AA, Azari-Hamidian S, Vatandoost H, Charrahy Z (2011) Spatiotemporal distribution of malaria vectors (Diptera: Culicidae) across different climatic zones of Iran. Asian Pacific J Trop Med. 4(6): 498–504.
- 84. Hanafi-Bojd AA, Vatandoost H, Oshaghi MA, Haghdoost AA, Shahi M, Sedaghat MM (2012) Entomological and epidemiological attributes for malaria transmission and implementation of vector control in southern Iran. Acta Trop. 121(2): 85–92.
- Mehravaran A, Vatandoost H, Oshaghi MA, Abai MR, Edalat H, Javadian E (2012) Ecology of *Anopheles stephensi* in a malarious area, southeast of Iran. Acta Medica Iranica. 50(1): 61–65.
- 86. Soleimani-Ahmadi M, Vatandoost H, Hanafi-Bojd AA, Zare M, Safari R, Mojahedi A (2013) Environmental characteristics of anopheline mosquito larval habitats in a malaria endemic area in Iran. Asian Pacific J Trop Med. 6(7): 510–515.
- 87. Soleimani-Ahmadi M, Vatandoost H, Shaeghi M, Raeisi A, Abedi F, Eshraghian MR (2012) Vector ecology and susceptibility in a malaria-endemic focus in southern Islamic Republic of Iran. Eastern Mediterr Health J. 18(10): 1034– 1041.
- 88. Oshaghi MA, Chavshin AR, Vatandoost H, Yaaghoobi F, Mohtarami F, Noorjah N (2006) Effects of post-ingestion and physical conditions on PCR amplification of host blood meal DNA in mosquitoes. Exp Parasitol. 112(4): 232–236.
- 89. Nikookar S, Moosa-Kazemi SH, Oshaghi M, Yaghoobi-Ershadi M, Vatandoost H, Kianinasab A (2010) Species composition and diversity of mosquitoes in Neka County, Mazandaran Province, north-

ern Iran. Iran J Arthropod Borne Dis. 4 (2): 26–34.

H Edalat et al.: Ecology of ...

- 90. Vatandoost H, Abai MR, Abbasi M, Shaeghi M, Abtahi M, Rafie F (2009) Designing of a laboratory model for evaluation of the residual effects of deltamethrin (K-othrine WP 5%) on different surfaces against malaria vector, *Anopheles stephensi* (Diptera: Culicidae). J Vector Borne Dis. 46(4): 261–267.
- 91. Vatandoost H, Hanafi-Bojd AA (2008) Laboratory evaluation of 3 repellents against *Anopheles stephensi* in the Islamic Republic of Iran. Eastern Mediterr Health J. 14(2): 260–267.
- 92. Oshaghi MA, Chavshin AR, Vatandoost H (2006) Analysis of mosquito bloodmeals using RFLP markers. Exp Parasitol 2006. 114(4): 259–262.
- 93. Vatandoost H, Mesdaghinia AR, Zamani G, Madjdzadeh R, Holakouie K, Sadrizadeh B (2004) Development of Training the Regional Malaria Centre in Bandar Abbas, Islamic Republic of Iran. Eastern Mediterr Health J. 10(1–2): 215– 224.
- 94. Khoshdel-Nezamiha F, Vatandoost H, Azari-Hamidian S, Bavani MM, Dabiri F, Entezar-Mahdi R (2014) Fauna and larval habitats of mosquitoes (diptera: culicidae) of west Azerbaijan Province, Northwestern Iran. J Arthropod Borne Dis. 8(2): 163–173.
- 95. Chavshin AR, Oshaghi MA, Vatandoost H, Hanafi-Bojd AA, Raeisi A, Nikpoor F (2014) Molecular characterization, biological forms and sporozoite rate of *Anopheles stephensi* in southern Iran. Asian Pacific J Trop Biomed. 4(1): 47–51.
- 96. Chavshin AR, Oshaghi MA, Vatandoost H, Pourmand MR, Raeisi A, Terenius O (2014) Isolation and identification of culturable bacteria from wild *Anopheles culicifacies*, a first step in a paratransgenesis approach. Parasit Vectors. 7: 419–424.

- 97. Karimian F, Oshaghi MA, Sedaghat MM, Waterhouse RM, Vatandoost H, Hanafi-Bojd AA (2014) Phylogenetic analysis of the oriental-Palearctic-Afrotropical members of Anopheles (Culicidae: Diptera) based on nuclear rDNA and mitochondrial DNA characteristics. Jpn J Infect Dis. 67(5): 361–367.
- 98. Chavshin AR, Oshaghi MA, Vatandoost H, Yakhchali B, Zarenejad F, Terenius O (2015) Malpighian tubules are important determinants of Pseudomonas transstadial transmission and longtime persistence in *Anopheles stephensi*. Parasit Vectors. 8: 36–42.
- 99. Khoshdel-Nezamiha F, Vatandoost H, Oshaghi MA, Azari-Hamidian S, Mianroodi RA, Dabiri F (2016) Molecular Characterization of Mosquitoes (Diptera: Culicidae) in Northwestern Iran by Using rDNA-ITS2. Jpn J Infect Dis. 69 (4): 319–322.
- 100. Shayeghi M, Vatandoost H, Gorouhi A, Sanei-Dehkordi AR, Salim-Abadi Y, Karami M (2014) Biodiversity of aquatic insects of Zayandeh Roud River and its branches, Isfahan Province, Iran. J Arthropod Borne Dis. 8(2): 197–204.
- 101. Gezelbash Z, Vatandoost H, Abai MR, Raeisi A, Rassi Y, Hanafi-Bojd AA (2014) Laboratory and field evaluation of two formulations of *Bacillus thuringiensis* M-H-14 against mosquito larvae in the Islamic Republic of Iran, 2012. Eastern Mediterr Health J. 20(4): 229– 235.
- 102. Anjomruz M, Oshaghi MA, Pourfatollah AA, Sedaghat MM, Raeisi A, Vatandoost H (2014) Preferential feeding success of laboratory reared *Anopheles stephensi* mosquitoes according to ABO blood group status. Acta Trop. 140: 118–123.
- 103. Anjomruz M, Oshaghi MA, Sedaghat MM, Pourfatollah AA, Raeisi A, Vatandoost H (2014) ABO blood groups of resi-

dents and the ABO host choice of malaria vectors in southern Iran. Exp Parasitol. 136: 63–67.

- 104. Soleimani-Ahmadi M, Vatandoost H, Zare M (2014) Characterization of larval habitats for anopheline mosquitoes in a malarious area under elimination program in the southeast of Iran. Asian Pacific J Trop Biomed. 4(Suppl 1): 573–580.
- 105. Soleimani-Ahmadi M, Vatandoost H, Zare M, Alizadeh A, Salehi M (2014) Community knowledge and practices regarding malaria and long-lasting insecticidal nets during malaria elimination programme in an endemic area in Iran. Malar J. 13: 511–518.
- 106. Soleimani-Ahmadi M, Vatandoost H, Zare M, Turki H, Alizadeh A (2015) Topographical distribution of anopheline mosquitoes in an area under elimination programme in the south of Iran. Malar J. 14: 262.
- 107. Soleimani-Ahmadi M, Vatandoost H, Zare M, Turki H, Alizadeh A (2015) Topographical distribution of anopheline mosquitoes in an area under elimination programme in the south of Iran. Malar J. 14(1): 1.
- 108. Ataie A, Moosa-Kazemi SH, Vatandoost H, Yaghoobi-Ershadi MR, Bakhshi H, Anjomruz M (2015) Assessing the susceptibility status of mosquitoes (Diptera: Culicidae) in a Dirofilariasis focus, northwestern Iran. J Arthropod Borne Dis. 9(1): 7–21.
- 109. Soltani A, Vatandoost H, Oshaghi MA, Ravasan NM, Enayati AA, Asgarian F (2015) Resistance mechanisms of *Anopheles stephensi* (Diptera: Culicidae) to Temephos. J Arthropod Borne Dis. 9 (1): 71–83.
- 110. Golfakhrabadi F, Khanavi M, Ostad SN, Saeidnia S, Vatandoost H, Abai MR (2015) Biological activities and composition of *Ferulago carduchorum* Essen-

tial Oil. J Arthropod Borne Dis. 9(1): 104–115.

- 111. Nikookar SH, Moosa-Kazemi SH, Oshaghi MA, Vatandoost H, Yaghoobi-Ershadi MR, Enayati AA (2015) Biodiversity of culicid mosquitoes in rural Neka township of Mazandaran Province, northern Iran. J Vector Borne Dis. 52(1): 63–72.
- 112. Nikookar SH, Moosa-Kazemi SH, Yaghoobi-Ershadi MR, Vatandoost H, Oshaghi MA, Ataei A (2015) Fauna and larval habitat characteristics of mosquitoes in Neka County, Northern Iran. J Arthropod Borne Dis. 9(2): 253–266.
- 113. Chavshin AR, Dabiri F, Vatandoost H, Bavani MM (2015) Susceptibility of Anopheles maculipennis to different classes of insecticides in West Azarbaijan Province, Northwestern Iran. Asian Pacific J Trop Biomed. 5(5): 403–406.
- 114. Shayeghi M, Nejati J, Shirani-Bidabadi L, Koosha M, Badakhshan M, Bavani MM (2015) Assessing the fauna of aquatic insects for possible use for malaria vector control in large river, central iran. Acta Medica Iranica. 53(9): 523–532.
- 115. Pirmohammadi M, Shayeghi M, Vatandoost H, Abai MR, Mohammadi A, Bagheri A (2016) Chemical composition and repellent activity of *Achillea vermiculata* and *Satureja hortensis* against *Anopheles stephensi*. J Arthropod Borne Dis. 10(2): 201–210.
- 116. Gorouhi MA, Vatandoost H, Oshaghi MA, Raeisi A, Enayati AA, Mirhendi H (2015) Current susceptibility status of *Anopheles stephensi* (Diptera: Culicidae) to different imagicides in a malarious area, southeastern Iran. J Arthropod Borne Dis. 10(4): 493–500.
- 117. Abai MR, Hanafi-Bojd AA, Vatandoost H (2015) Laboratory Evaluation of Temephos against *Anopheles stephensi* and *Culex pipiens* larvae in Iran. J Arthropod Borne Dis. 10(4): 510–518.

118. Sanei-Dehkordi A, Vatandoost H, Abai MR, Davari B, Sedaghat MM (2016) Chemical Composition and larvicidal activity of *Bunium persicum* essential oil against two important mosquitoes vectors. J Essent Oil Bear Pl. 19(2): 349– 357.