Original Article

First Comprehensive List of Phlebotomine Sand Fly Species (Diptera: Psychodidae) in a Leishmaniasis Focus (Djelfa, Algeria), Including the First Record of *Phlebotomus langeroni* Females in Algeria

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Abstract

Background: The wide distribution of phlebotomine vectors complicates the leishmaniasis situation in the world, with the risk of spreading from rural to urban areas. Our study investigates for the first time the ecology and distribution of sand fly populations in leishmaniasis focus (Djelfa, Algeria).

Methods: Sampling is performed using light traps from August 2021 to July 2022 at ten sites with different biotopes: two peri-urban stations (Ain Oussera and Hassi Bahbah), one urban station (Djelfa), and three rural stations (Ain El-Bel, Haniet Ouled Salem and Mlaga).

Results: A total of 2,866 specimens with 12 species belonging to *Phlebotomus* and *Sergentomyia* were identified with the first record of *Phlebotomus langeroni* female nationally, revealing a good diversity of up to 1.812 bits. The greatest richness of species was found in rural sites at Ain El Bel station, while the lowest richness was observed in the first periurban site of Ain Oussera station. Interestingly, the surveyed species were evenly distributed across all the study sites, in particular *Phlebotomus papatasi* with 57.29%. Statistical treatment revealed a strong positive relationship between temperature and species (p < 0.001, r = 0.91). Considering the other factors, there was a moderate link between species richness and altitude ($p = 0.01 R^2 = 0.519$). However, neither rainfall nor wind speed (p > 0.05) were correlated with sand fly fauna. Moreover, humidity showed a negative correlation (p < 0.001, r = -0.96).

Conclusion: The findings of this study on phlebotomine fauna in this endemic region for leishmaniasis provide valuable data to support the success of a vector control program.

Keywords: Sand fly; Environmental factors; Leishmania; Djelfa; Algeria

Introduction

Phlebotomine sand flies (Diptera: Psychodidae) are tiny haematophagous insects that take blood meals on a wide range of vertebrate hosts including humans (1–3). These arthropods can potentially transmit important pathogens such as the protozoan parasites of *Leishmania* spp. (4–6). Female sand flies infected with these parasites are known to carry pathogens that cause a range of diseases known as leishmaniasis, which affects humans and dogs causing public and veterinary health issues. The most notable forms including cutaneous leishmaniasis (CL) and visceral leishmaniasis (VL) (7, 8).

In Algeria, as in several Mediterranean and African countries, leishmaniasis is an endemic disease and the risk of important outbreaks is real. This disease has considerably increased over the last years (9). In Algeria, five species of *Phlebotomus* are proven vectors, while three

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species of Leishmania are declared as causative agents of the disease (10). In the Sahara, Phlebotomus papatasi (Scopoli, 1780) is the known vector of Leishmania major (11). In the northern regions of Algeria, Ph. perniciosus (Newstead, 1911), Ph. perfiliewi (Parrot, 1930), and probably Ph. longicuspis (Nitzulescu, 1930) are vectors of L. infantum, causing two forms of the disease: CL and VL (12-15). In several regions of the country such as Ghardaïa in the South, Tipaza in the North-Center, and Annaba in the North-East, the vector Ph. sergenti is responsible for the transmission of L. killicki, which causes CL (16-19). The Djelfa region is endemic for leishmaniasis, primarily due to its agro-pastoral vocation, arid climate, significant population, and close geographical proximity to highly infected regions including M'sila to the East, Biskra to the South-East and Ghardaia to the South (20-24).

Entomological data specific to this unique geographical region are particularly scarce, highlighting a major lack of epidemiological information on phlebotomine sand flies in the Djelfa region. Therefore, the purpose of this research initiative is to respond to this lack by carrying out an in-depth survey for the first time, by establishing a rigorous and comprehensive taxonomic inventory of the sand fly species present in the area, with particular emphasis on their spatio-temporal dynamics. Consequently, this survey will make a significant contribution to advancing ecological and epidemiological knowledge of leishmaniasis transmission in the Djelfa region and, more widely, throughout central Algeria. Implications of our results for monitoring sand fly populations and, at the same time, the occurrence of leishmaniasis are discussed.

Materials and Methods

Study area

Djelfa region stretches over an area of $66,415 \text{ km}^2$, with a population of 1,538,476 (according to the updated census of 2018), As one of the largest steppe regions in Algeria,

Djelfa is located on the high plateaus of central Algeria between the Tellian Atlas in the North and the Saharan Atlas in the South. with coordinates stretching from 2° and 5° East longitude to 33° and 35° North latitude. It comprises 36 communes and is recognized for its arid climate, marked by cold winters and hot summers. This climatic setting has propelled the region into eminence for its dominant pastoral vocation, serving a significant center for the sheep sector, as well as for arboriculture, cereal cultivation, market gardening, and fodder crops.

Sand fly collection and identification

The study sites were selected based on the most infected areas with leishmaniasis, according to epidemiological data from the Department of Health and Population (20). The surveys were carried out across 181 km transect linking the six study stations, in which ten sampling sites were selected (Fig. 1). The sampling took place from August 2021 to July 2022, over a year (i.e., 12 months) with one outing each month. The sites were chosen according to the type of area as follows: peri-urban (Ain Oussera and Hassi Bahbah), urban (center of Djelfa), rural (Ain El-Bel, Haniet Ouled Salem and Mlaga) (Table 1).

To ensure a high capture rate, CDC light traps were used. The selection of this trap was predicated on its effectiveness in attracting phototropic species and its ability to capture active sand flies (25, 26). Four light traps were set up from 7 p.m. to 7 a.m. for one night per month from August 2021 to July 2022. It was strategically placed within and in the vicinity of human and animal habitats, as well as in proximity to vegetation.

Specimens captured using light traps were stored in 70% ethanol. This was followed by dissection, in which we separated the head and genitalia from the body, and then these were clarified with 20% Potassium Hydroxide for 4 hours and Marc-André liquid for 2 hours and then fixed in chloral gum (25). Taxonomic identification was carried out individually with a focus on morphological characteristics of male genitalia and female structures such as spermatheca, pharynx, and cibarium, as described by Abonnenc (25), Dedet et al. (27), El Sawaf and Said (28) and Depaquit et al. (29). For some specimens that require further validations for identification, we used the Phleb Key Tool identification software (30).

Collection of climatic data

The climatic parameters analysed in this study included the monthly data on maximum, average or minimum temperatures, rainfall, and wind speed obtained from the meteorological office of Djelfa province.

Data analysis

The data regarding the phlebotomine fauna was subjected to ecological analysis, employing various ecological indexes related to both composition and structure. The measures of composition that were included are relative abundance $(RA\% = ni/N \times 100)$, where ("*ni*"= number of specimens of each sand fly species and "N" = number of total specimens in the surveyed station); Species richness (S), where ("S"= total number of species identified in each survey); and frequency of occurrence $(FO = Ri/R \times 100)$, where ("Ri"= number of surveys containing the surveyed species and"*R*"= total number of surveys carried out). For the structure index, we used the Shannon-Weaver index $(H' = -\Sigma pi \log_2 pi)$, which is expressed in bit units ("pi"= the relative frequency of the species considered and "log₂"= the natural logarithm to base-2) and equipartition index (E = H'/H'max), where ("*H'max*"= maximum diversity that is represented as follows: $H'max = log_2 S$). Simpson's reciprocal index (SRI = 1/D) and dominance index $(SDI = \Sigma Ni (Ni - 1)/N (N - 1))$, where ("Ni"= number of species specimens considered and "N" = total number of species specimens) (31-34). Statistical analyses were conducted using the software R, v. 4.3.0. The graphs of multilines of ecological indexes, of points of species richness, and the altitude regression were performed using the packages Tidyverse, ggplot2 and reshape as well as species phenology (35–37). Correlation matrix between all of these ecological parameters was plotted using Corrplot (38). Furthermore, Metan package was applied to plot the Pearson correlation between all the environmental factors and surveyed species (39).

Results

Diversity of the sand fly fauna

Investigation of the sand fly population in Djelfa during the survey period from August 2021 to July 2022 in four different biotopes, urban, peri-urban, and rural, revealed a total of 2,866 specimens spread over 12 species belonging to two genera: *Phlebotomus* and *Sergentomyia* (Table 2).

The analysis of the indices of diversity and homogeneity have revealed variations from one site to another (Fig. 2). The highest richness was found in sites 7 and 8 of Ain El Bel station (S=9) with an abundance of 315 in site 7 and 273 in site 8, while the lowest richness was observed in site 1 at Ain Oussera station. Shannon Weaver index values exhibited a range between 0.543 bits and 1.812 bits, indicative of a commendable diversity within the surveyed area. The equitability index revealed relatively even species distribution across most sites, with values spanning from E = 0.572 to E = 0.825. In contrast, the sites 1 (in the animal habitats) and 3 (surrounding human habitats) deviated from this pattern, displaying lower equitability values (E = 0.494and E = 0.404, respectively). Examining Simpson's indices, these latter sites featured the highest values for Simpson's dominance index (SDI = 0.721 and SDI = 0.713, respectively)and the lowest values for Simpson's reciprocal index (SRI = 1.386 and SRI = 1.402, respectively), indicating a lower degree of species dominance. In contrast, site 8 (in the animal habitats) exhibited a substantially lowest Simpson's dominance index (SDI = 0.215) and a notably highest Simpson's reciprocal index (SRI = 4.647), reflecting a higher level of species dominance within this particular site.

Composition of the sand fly fauna

The comprehensive list of sand flies collected is presented in Table 2. The genus Phlebotomus is represented by seven species of which Ph. papatasi was the most abundant constituting 57.29% of the total specimens. This species exhibited a presence in various seasons, with 8.37% in the summer of 2021, 28.30% in autumn, 1.78% in spring, and 18.84% in the summer of 2022. It is followed by Ph. alexandri with 10.19% of the total population. Its distribution across the seasons was as follows: 2.38% in summer 2021, 13.44 % in summer 2022, 8.73% in autumn, and 0.24% in spring. The lowest reported species was Ph. longiscuspis that only appeared in summer 2022 with (2.09%). Concerning the second genus Sergentomyia, five species were collected. Among them, Se. minuta was particularly abundant, representing 9.42% of the overall population. Its distribution encompassed various seasons with 1.19% in summer 2021, 2.23% in autumn, 0.03% in spring, and 12.85% in summer 2022. It is followed by Se. fallax with a very low prevalence, constituting 3.52% of the total, respectively detailed in summer 2021 with (0.14%), in autumn with (1.05%) and in summer 2022 with (2.23%). The least represented species is Se. dreyfussi. Only one single specimen, with a rate of 0.03%, represents this species. In addition, most of the surveyed species were captured with both sexes male and female. Exception for Se. dreyfussi and Se. minuta where only females were captured and for Se. antennata and Se. schwetzi where only males were collected.

Seasonal phenology and occurrence of phlebotomine species

The seasonal occurrence and phenology of

phlebotomine sand flies in the Djelfa region from August 2021 to July 2022 revealed that Ph. papatasi is ubiquitous with the occurrence of 100% throughout summer 2021, autumn, and summer 2022 and occasional in spring with 33% at the study area. On the other hand, Ph. alexandri is presented only in autumn and summer 2022 in sites 1 and 2 and is considered as an accidental species (FO%≤25 %). Interestingly, the same species transitioned to the occasional ubiquitous category in sites 5, 6, and 10 in summer 2021 (100%), autumn (67%), spring (33%), and summer 2022 (100 %). A similar occurrence pattern was noted for Ph. sergenti at site 6 and Ph. perniciosus at sites 5, 6, and 7. Furthermore, Se. schwetzi displayed an occurrence of 67% in site 7 and 33% in site 8 during autumn, while reaching 100% in summer 2022 in both sites. During the two summers, this species was only observed in one site out of the ten surveyed. The occurrence of Ph. ariasi in site 7 was occasional in autumn and ubiquitous in the summer of 2022. However, at sites 2, and 8, this species exhibited occurrences ranging from 33% to 100% during both summer and autumn.

The accidental occurrence of *Se. antenna*ta during all seasons ($F0\% \le 25$) was observed in some sites as follows: in site 3 during summer 2022 and site 5 during summer 2021, with the exception that this species was recorded during summer 2021, autumn, and summer 2022 in sites 4, 7 and 8. In addition, *Ph. longiscuspis* was only observed during summer 2022 in site 10 with 100%.

Likewise, *Ph. langeroni* occurred occasionally in sites 7 with 42% and site 8 with 33%, whereas this species occurred accidentally in site 3 with 8%. It is important to point out that this species is being reported for the first time in Algeria and that no studies are revealing its presence in endemic outbreaks of leishmaniasis. *Phlebotomus langeroni* females can be distinguished by several distinctive characteristics, including shorter wings (0.03 mm), a lengthier ascoid antennular segment IV (0.46 times the segment length), and a distinctive feature in the structure of the spermatheca. The spermatheca exhibits a reservoir composed of nine to ten annuli that extends into a duct culminating in a characteristic fusiform dilatation. This structural differentiation sets it apart from other *Larroussius* species (Fig. 4).

Statistical treatments

Correlation matrix based on ecological indices for phlebotomine species

The correlation matrix unveiled positive correlations among all the diversity parameters, except for the *SDI* parameter. Specifically, the latter parameter (*SDI*) exhibited a weak negative correlation with the number of species specimens but demonstrated strong negative correlations with the other indices. Notably, parameters *J*, *H*, *SRI* and *S* displayed strong positive correlations with each other. Conversely, the number of species specimens displayed weak positive correlations with the latter four parameters (Fig. 5).

Altitude distribution

The linear regression analysis revealed a moderate and average relationship between sand fly species richness and altitude $(P = 0.01; R^2 = 0.519)$. It was observed that species richness increased moderately and significantly with higher altitude. However, there was no significant relationship between spe-

cies abundance and altitude (P = 0.731; $R^2 = 0.016$), the results are shown in Fig. 6.

Pearson correlation between environmental factors and sand fly population

The correlation analyses carried out on the data obtained for environmental factors (maximum, mean and minimum temperatures, relative humidity, rainfall, and wind speed) and sand fly species are shown in Fig. 7.

The results unveiled robust and statistically significant positive correlations between maximum, mean, and minimum temperatures and all species belonging to the genus Phlebotomus (except for Ph. longiscuspis) and to the genus Sergentomyia (excluding Se. drevfussi). The correlation values ranged from (p < 0.05, r = 0.58 to p < 0.001, r = 0.91).In contrast, concerning humidity, a pronounced and statistically significant negative correlation was observed with the sand fly population, except for two species, Ph. longiscuspis, and Se. dreyfussi. The correlation coefficients for humidity ranged from (p < 0.05, r = 0.59)to p < 0.001, r = 0.89). As for the correlations between rainfall and wind speed, no statistically significant relationships were detected with any of the sand fly species.

Station	Site	Coordinates	Altitude (m asl)	Biotope	Site descrip- tion	Vegetation	Water source
Ain	1	35°28'18.815'	661	Periurban	Animal habi-	++	Irrigation+
Oussera		'N			tats		Oued
	2	2°54'40.299'' E		Periurban	Surrounding human habi- tats	+	Irrigation+ Oued
Hassi Bahbah	3	35°3'4.63''N 3°2'47.264''E	850	Periurban	Surrounding human habi- tats	+	Irrigation
	4			Periurban	Animal habi- tats	++	Irrigation

Table 1. Characteristics of sam	pling sites for sand flies in st	udy region Djelfa, A	lgeria, 2021–2022

http://jad.tums.ac.ir Published Online: June 30, 2024

Djelfa	5	34°39'32.693'	1186	Urban	Surrounding	+	Irrigation+
-		'N			human habi-		Oued
		3°13'12.636''			tats		
	6	Е		Urban	Animal habi-	++	Irrigation+
					tats		Oued
Ain El-	7	34°20'47.347'	1043	Rural	A junk room	++	Irrigation
bel	8	'N		Rural	Animal habi-	+++	Irrigation
		3°13'39.093''			tats		U U
		Е					
Mlaga	9	34°12'58.832'	811	Rural	Animal habi-	+++	Irrigation+
U		'N			tats		Oued
		3°23'49.377'E					
Haniet	10	34°10'14.849'	790	Rural	Animal habi-	+++	Irrigation
Ouled		'N			tats		8
Salem		3°27'22.755''					
		Е					

Table 1. Continued ...

(+): low; (++) medium; (+++) dense, asl: above sea level

Table 2. Inventory and abundance of phlebotomine species sampled in Djelfa region, Algeria (August 2021–July
2022)

			Total		Summer 2021		Autumn		Winter		Spring 2022		Summer 2022	
	Sex	Ν	Ν	RA%	Ν	RA%	Ν	RA%	Ν	RA%	Ν	RA%	Ν	RA%
Phlebotomus	М	484	1642	57.29	240	8.37	811	28.30	_	_	51	1.78	540	18.84
papatasi (Scopoli, 1780)	F	1158												
Ph. langeroni	Μ	23	89	3.10	6	0.21	27	0.94	_	_	0	0	56	1.95
(Nitzulescu, 1930)	F	66												
Ph. sergenti	Μ	10	63	2.20	3	0.10	18	0.63	_	_	2	0.07	40	1.40
(Parrot, 1917)	F	53												
Ph. perniciosus	Μ	18	142	4.95	3	0.10	64	2.23	_	_	0	0	75	2.62
(Newstead, 1911)	F	124												
Ph. longiscuspis	Μ	4	60	2.09	0	0	0	0	_	_	0	0	60	2.09
(Nitzulescu, 1930)	F	56												
Ph. alexandri	Μ	16	292	10.19	8	0.28	104	3.63	_	_	7	0.24	173	6.04
(Sinton, 1928)	F	276												
Ph. ariasi	Μ	2	107	3.73	24	0.84	52	1.81	_	_	0	0	31	1.08
(Tonnoir, 1921)	F	105												
Sergentomyia	Μ	56	56	1.95	3	0.10	9	0.31	_	_	0	0	44	1.54
schwetzi	F	0												
(Theodor&Parrot, 1929)	-	0												
Se. fallax	Μ	12	101	3.52	4	0.14	30	1.05	_	_	3	0.10	64	2.23
(Parrot, 1921)	F	89												
Se. antennata	Μ	43	43	1.50	11	0.38	12	0.42	_	_	0	0	20	0.70
(Newstead, 1921)	F	0												
Se. dreyfussi	М	0	1	0.03	0	0	0	0	_	_	0	0	1	0.03
(Parrot, 1933)	F	1												
Se. minuta	М	0	270	9.42	34	1.19	64	2.23	_	_	1	0.03	171	5.97
(Rondani, 1843)	F	270												
Total			2866	100	336	11.71	1191	41.55	_	_	64	2.22	1275	44.49

-: Absence; M: male; F: female; RA: relative abundance

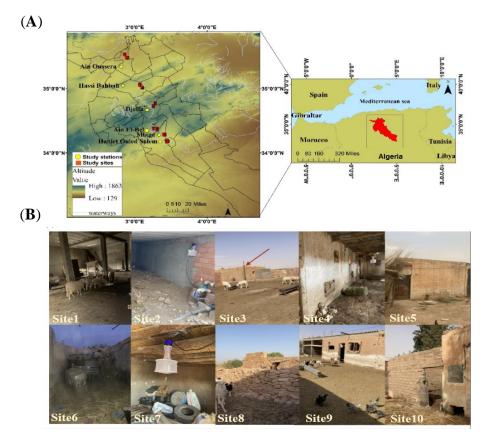


Fig. 1. (A) Map showing the geographical position of the Djelfa region, highlighting the study sites. (B) Photographs showing the habitats of the study sites: in the animal habitats (sites 1, 4, 6, 8, 9, and 10); Surrounding human habitats (sites 2, 3, and 5); and site 7, representing a junk room

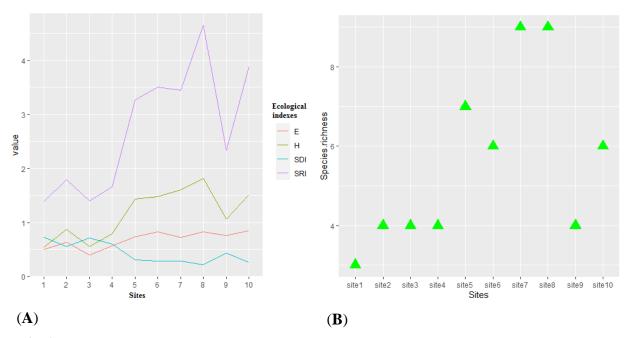


Fig. 2. Ecological analysis of the phlebotomine species in the study sites (A) Ecological indices: Shannon-Weaver index (*H*[']), Equipartition (*E*), Simpson's reciprocal index (*SRI*) Simpson's dominance index (*SDI*) and (B) Species richness

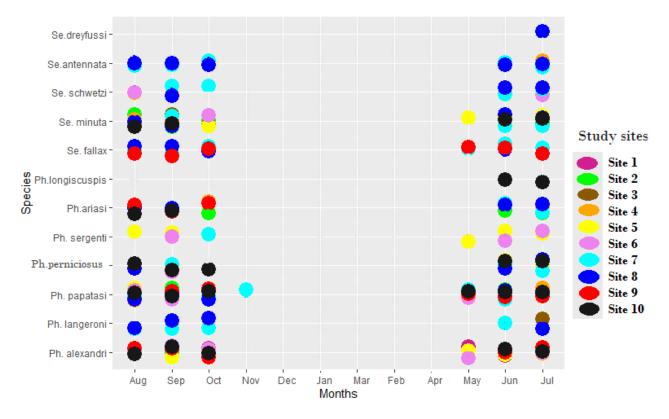


Fig. 3. Phenology of phlebotomine species surveyed in Djelfa region, Algeria (August 2021–July 2022)

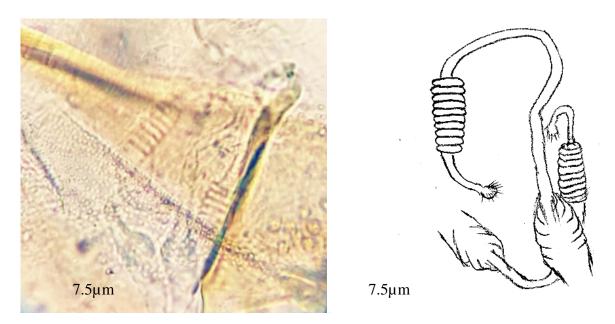


Fig. 4. Spermatheca of Phlebotomus langeroni female captured in Djelfa region (Algeria), 2021–2022

SDI						-0.8
-0.41	N					0.6
-0.95	0.44	E				0.2
-0.99	0.38	0.90	Н			-0.2
-0.96	0.36	0.88	0.98	SRI		-0.4 -0.6
-0.84	0.27	0.65	0.91	0.88	S	-0.8

Fig. 5. Corrplot of the correlation matrix between ecological parameters of phlebotomine species from August 2021 to July 2022 in the Djelfa region. The parameters are SDI (Simpson's Dominance Index), E (Equipartition Index), N (Total Number of Species Specimens), H (Shannon-Weaver Index), SRI (Simpson's Reciprocal Index), and S (Species Richness); blue circles represent positive correlations, while red circles represent negative correlations and color degradations represent the intense of correlation

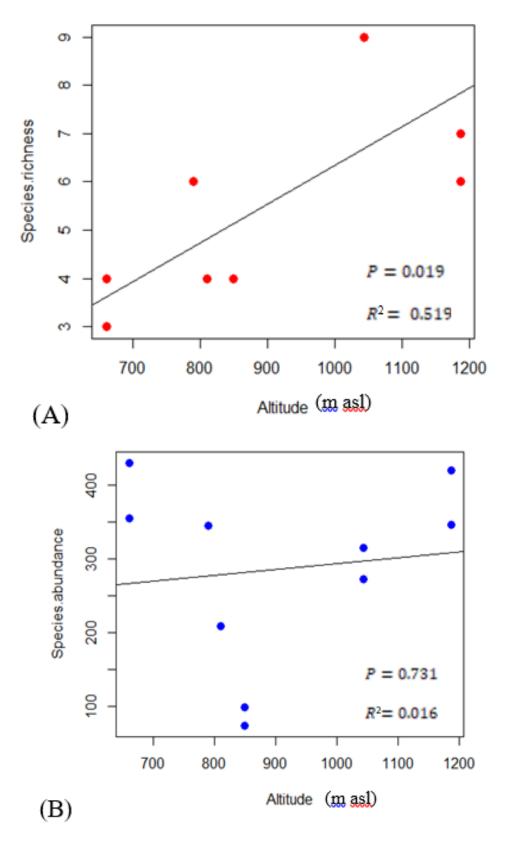


Fig. 6. Plot showing the relationship between altitude (m asl) and sand fly species richness (A); abundance (B) in Djelfa region, Algeria, 2021–2022

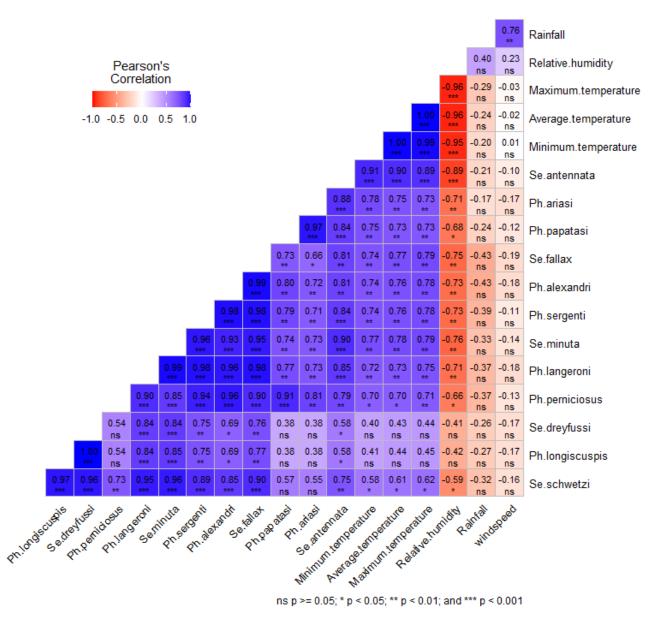


Fig. 7. Corrplot of the Pearson correlation between all the environmental factors and the phlebotomine species in the Djelfa region, Algeria, 2021–2022

Discussion

In Algeria, the risk of leishmaniasis has increased significantly in recent years, according to the Algerian National Institute of Health and Population. In the current study, the first entomological survey was carried out in Djelfa, one of the regions mostly affected by this disease (20). Our investigations led to the identification of 12 species of both genera *Phlebotomus* and *Sergentomyia*, out of the 26 species known in Algeria (41). All species that are proven or suspected vectors of *Leishmania* have been found in the current survey. Starting with the genus *Phlebotomus*, *Ph. papatasi* emerges as the most prevalent species, constituting 57.29% of the observed specimens across all surveyed biotopes. The known distribution of this species is in steppe areas and the northern Sahara, and it is a proven vector of *L. ma*- *jor* in Algeria (42). Although it can be found in the Tell region and the Highlands (27). For Ph. alexandri which represents 10.19% of collected sand flies in the peri-urban and urban biotopes. This species is known for its ability to transmit several Leishmania species such as L. major, L. infantum, L. donovani, and L. tropica, and it inhabits a wide range of environments, extending from natural habitats to periurban and urban areas (43-47). In China, this species is present in restricted areas such as the stony desert (48, 49). Phlebotomus perniciosus, found in rural and urban areas at a rate of 4.95%, is a proven vector of L. infantum MON 1, which causes visceral leishmaniasis in rural villages (50). This species is mostly distributed in the northern part of Algeria (27). Phlebotomus ariasi, collected in rural and periurban settings with a prevalence of 3.73%, has been proved as a vector of L. infantum in Morocco and Tunisia, causing VL (51, 52).

Phlebotomus langeroni, comprises 3.1% of the specimens in peri-urban and rural areas. We achieved the interesting presence of female specimens of this species in two rural sites at the Ain El Bel station, with a total of 66 specimens, although this is a significant number, it is certainly an underestimate of reality. Furthermore, a unique specimen was collected in peri-urban site 3. This species was found infected with L. infantum, responsible for CL in northern Tunisia (53) as well as L. donovani which causes VL in Egypt (54, 55). This sand fly occurs particularly in dry, rocky biotopes, where rodent and reptile breeding grounds are found (27). In Algeria, only male specimens of the Ph. langeroni species have been documented (10, 13, 27) and this is the first survey that highlights the presence of females of this species.

Phlebotomus sergenti is distributed in rural and urban (2.2%) and is the proven vector of *L. tropica* in Morocco (56) and Iran (57) and *L. killicki* in Algeria (16) and in Tunisia (58), where it is mostly found in the South of Algeria in the Saharan Atlas mountains, the pre-Saharan steppes and the Tell areas (16, 27, 59). *Phlebotomus longicuspis*, representing 2.09% in rural areas, has epidemiologically been linked to *L. infantum* transmission in bioclimatic arid zones of northern Africa (14, 60, 61).

With regard to sand flies belonging to the genus Sergentomyia, several studies have raised the potential involvement of species belonging to this genus as vectors of Leishmania (62, 63). In the present survey, six species were identified in a variety of biotopes. Previous studies in northern Africa prove that all these species were found to be infected by L. major in rocky areas, the natural biotope of the rodents (64, 65). This is in line with the features of the study region Djelfa, which is characterized by its great diversity of rodents, in particular Meriones shawii, which is considered the secondary L. major reservoir after the sand rodent Psammomys obesus and plays an important role in the transmission of zoonotic cutaneous leishmaniasis in pastoral areas (66, 67).

Investigations on the potential of *Sergentomyia* species as vectors of the parasite to mammal hosts are actively pursued in several countries such as Tunisia, Kenya, Iran, and Portugal (65, 68, 69, 47). In Algeria, there exists a scarcity of experimental research focusing on the genus *Sergentomyia*, emphasizing the need for greater attention in future studies (41).

To understand the epidemiology and ecology of parasite activity, environmental and demographic factors must be taken into account jointly (70). This study marks the first investigation into the ecology and spatio-temporal distribution of sand fly populations within the Djelfa region. Initially, the effect of climatic and environmental factors on the dynamics of these species, which are distinguished by various ecological and biological traits, was assessed. Next, the findings were compared with other data to discern alterations in the seasonal dynamics and abundance of sand flies.

Throughout the winter months, all sand fly species were absent due to their diapause response, an adaptation to survive the cold season (71,72). However, during the summer of 2021 and subsequently, in the autumn, spring, and summer of 2022, most species reappeared, primarily due to the increasing temperature. It is noteworthy that the increase in temperature, associated with the socio-ecological changes transpiring in the region (humidity, rainfall, and altitude), could potentially influence the dynamics of Leishmania infection. Additionally, it must be highlighted that in this study, the CDC light trap was used exclusively for the faunistic study of sand flies. This trap choice imposes a considerable limitation on capturing sand fly species as effectively as other trapping techniques, such as sticky traps.

The correlation of Pearson has shown a positive link ranging from (p < 0.05, r = 0.58 to p < 0.001, r = 0.91). As a result of the opposite relationship between relative humidity and temperature, a negative correlation has been noted with sand flies to humidity ranging between (p < 0.05, r = 0.59 to p < 0.001 r = 0.96). The findings of this correlation are in line with those reported in central Morocco (73). Although there was no correlation between relative humidity and temperature between the two species Se. drevfussi and Ph. longiscuspis, which could be related to the disappearance of these two species during most sampling months and their appearance only during the summer of 2022. Neither rainfall nor wind speed (p > 0.05) were correlated with phlebotomine fauna, which is consistent with the previous results from central and southern Morocco, where both factors were determined not to be decisive for sand fly abundance (73, 74). The knowledge that bio-ecological factors play a significant role in the geographical and seasonal distribution of disease vectors (75, 76).

The relationship between altitude and the incidence of leishmaniasis is contingent on a myriad of environmental factors, most notably temperature, which plays a pivotal role in the

development of parasites within sand flies (77). In the current study, the linear regression analysis indicated a moderate and intermediate relationship between sand fly species richness and altitude. This aligns with observations in Morocco, Libya, and Turkey, where the distribution of sand fly species is notably influenced by altitude (78-81). In Golestan Province, Iran, an increase in the Shannon-Wiener and Evenness indices at higher altitudes, coupled with a decrease in the Simpson index, suggests that the sand fly fauna is more stable in areas with higher altitude compared to lower altitude areas (82). Inversely, regarding sand fly abundance, there is no discernible relationship with altitude, consistent with research conducted in plains of North-Eastern Italy (83) but contradictory to findings in South-Central Turkey, where altitude was found to significantly impact population abundance (84).

Conclusion

The widespread distribution of sand flies in the endemic focus of Djelfa along with the extended activity of 12 sand fly species highlights the potentially high risk of the spread of leishmaniasis throughout the Algerian steppes. These findings provide the foundation for subsequent research in the region. Furthermore, it is highly recommended that continuous monitoring efforts be implemented to mitigate the risk of spreading leishmaniasis disease. Additionally, the risk of spreading needs to be elucidated in terms of the climatic conditions that favour sand fly proliferation and to understand host-blood feeding preferences. These precious findings are essential for the development of effective control strategies, the optimization of operational measures, and the evaluation of their effectiveness.

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Ethical consideration

The ethics committee of the Laboratory of Exploration and Valorization of Steppe Ecosystems at the University of Djelfa (Algeria) has approved this entomological investigation.

Conflict of interest statement

The authors declare there is no conflict of interest.

References

- 1. Rogers ME (2012) The role of *Leishmania* proteophosphoglycans in sand fly transmission and infection of the mammalian host. Front Microbiol. 3(223): 1–13.
- Bongiorno G, Habluetzel A, Khoury C, Maroli M (2003) Host preferences of phlebotomine sand flies at a hypoendemic focus of canine leishmaniasis in central Italy. Acta Trop. 88(2): 109–116.
- Yared S, Gebresilassie A, Abbasi I, Aklilu E, Kirstein OD, Balkew M, Brown AS, Clouse RM, Warburg A, Hailu A, Teshome GM (2019) A molecular analysis of sand fly blood meals in a visceral leishmaniasis endemic region of northwestern Ethiopia reveals a complex hostvector system. Heliyon. 5(7): 1–10.
- 4. Dolmatova AV, Demina NA (1971) Les phlébotomes (Phlebotominae) et les maladies qu'ils transmettent. Vol. 18. Office de la recherche scientifique et technique outre-mer (ORSTOM). Paris.
- Addadi K, Dedet JP (1976) Epidemiology of leishmaniasis in Algeria. 6. Survey of clinical cases of infantile visceral leishmaniasis from 1965 to 1974. Bull Soc Pathol Exot. 69(1): 68–75.
- 6. Killick-Kendrick R (1990) The life-cycle of *leishmania* in the sandfly with special reference to the form infective to the vertebrate host. Ann Parasitol Hum Compa-

rée. 15(65): 37-42.

- Dostálová A, Volf P (2012) *Leishmania* development in sand flies: parasite-vector interactions overview. Parasit Vectors. 5(276): 1–12.
- Desjeux P (2004) Leishmaniasis: current situation and new perspectives. Comp Immunol Microbiol Infect Dis. 27(5): 305– 318.
- Alvar J, Vélez ID, Bern C, Herrero M, Desjeux P, Cano J, Jannin J, Boer MD, The WHO Leishmaniasis Control Team (2012) Leishmaniasis worldwide and global estimates of its incidence. PLoS One. 7(5): 1–12.
- Bennai K, Tahir D, Lafri I, Bendjaballah-Laliam A, Bitam I, Parola P (2018) Molecular detection of *Leishmania infantum* DNA and host blood meal identification in *Phlebotomus* in a hypoendemic focus of human leishmaniasis in northern Algeria. PLoS Negl Trop Dis. 12(6): 1–12.
- Boudrissa A, Cherif K, Kherrachi I, Benbetka S, Bouiba L, Boubidi SC, Benikhlef. R, Arrar L, Hamrioui B, Harrat Z (2012) Extension de *Leishmania major* au nord de l'Algérie. Bull la Société Pathol Exot. 105(1): 30–35.
- Izri MA, Belazzoug S (1993) Phlebotomus (Larroussius) perfiliewi naturally infected with dermotropic Leishmania infantum at Tenes, Algeria. Trans R Soc Trop Med Hyg. 87(4): 399 – 399.
- Berdjane-Brouk Z, Charrel RN, Hamrioui B, Izri A (2012) First detection of *Leishmania infantum* DNA in *Phlebotomus longicuspis* Nitzulescu, 1930 from visceral leishmaniasis endemic focus in Algeria. Parasitol Res. 111(1): 419–422.
- 14. Harrat Z, Pratlong F, Belazzoug S, Dereure J, Deniau M, Rioux JA, Belkaid M, Dedet JP (1996) *Leishmania infantum* and *L. major* in Algeria. Trans R Soc Trop Med Hyg. 90(6): 625–629.
- 15. Zait H, Ferhani Y, Achir I, Hamrioui B

(2012) Étude de 71 cas de leishmaniose viscérale diagnostiqués au CHU Mustapha d'Alger entre 1998 et 2009. Médecine Mal Infect. (3): 119–125.

- 16. Boubidi S, Benallal K, Boudrissa A, Bouiba L, Bouchareb B, Garni R, Bouratbine A, Ravel C, Dvorak V, Votypka J, Volf P, Harrat Z (2011) *Phlebotomus sergenti* (Parrot, 1917) identified as *Leishmania killicki* host in Ghardaïa, south Algeria. Microbes Infect. 13(7): 691–696.
- Mansouri R, Pratlong F, Bachi F, Hamrioui B, Dedet JP (2012) The first isoenzymatic characterizations of the *Leishmania* strains responsible for cutaneous leishmaniasis in the Area of Annaba (Eastern Algeria). The Open Conference Proceedings Journal. 3(suppl 2-M2): 6–11.
- Izri A, Bendjaballah A, Andriantsoanirina V, Durand R (2014) Cutaneous leishmaniasis caused by *Leishmania killicki*, Algeria. Emerg Infect Dis. 20(3): 502– 504.
- 19. Harrat Z, Boubidi SC, Pratlong F, Benikhlef R, Selt B, Dedet JP, Ravel C, Belkaid M (2009) Description of a dermatropic *Leishmania* close to L. killicki (Rioux, Lanotte and amp; Pratlong 1986) in Algeria. Trans R Soc Trop Med Hyg. 103(7): 716–720.
- Messaoudene F, Boukraa S, Boubidi SC, Guerzou A, Ouahabi A (2023) Human cutaneous leishmaniasis in North Africa and its threats to public health: A statistical study focused on Djelfa (Algeria). Microorganisms. 11(10): 1–14.
- Benikhlef R, Aoun K, Boudrissa A, Ben Abid M, Cherif K, Aissi W, Benrekta S, Boubidi SC, Späth GF, Bouratbine A, Sereno D, Harrat Z (2021) Cutaneous Leishmaniasis in Algeria; Highlight on the Focus of M'Sila. Microorganisms. 9(5): 1–22.
- 22. Randa G, Samir Z, Hamid B (2017) Association between climatic changes and leishmaniasis incidence in Biskra District, Algeria. J Entomol Zool Stud. 5:

43–49.

- Saadene Y, Salhi A, Mliki F, Bouslama Z (2023) Climate change and cutaneous leishmaniasis in the province of Ghardaïa in Algeria: A model-based approach to predict disease outbreaks. Ann Saudi Med. 43(5): 263–276.
- 24. Boukraa S, Boubidi S, Zimmer JY, Francis F, Haubruge E, Alibenali-Lounaci Z, Doumandji S (2011) Surveillance des populations de phlébotomes (Diptera: Psychodidae), vecteurs des agents responsables des leishmanioses dans la région du M'Zab-Ghardaïa (Algérie). Entomol Faun Faun Entomol. 63(3): 97–101.
- 25. Abonnenc E (1972) Les phlébotomes de la région éthiopienne (Diptera, Psychodidae).Vol.2. Office de la recherche scientifique et technique outre-mer (ORSTOM). Paris.
- 26. Alten B, Ozbel Y, Ergunay K, Kasap OE, Cull B, Antoniou M, Velo E, Prudhomme J, Molina R, Bañuls AL, Schaffner F, Hendrickx G, Bortel WV, Medlock JM (2015) Sampling strategies for phlebotomine sand flies (Diptera: Psychodidae) in Europe. Bull Entomol Res. 105(6): 664–678.
- 27. Dedet JP, Addadi K, Belazzoug S (1984) Les phlébotomes (Diptera, Psychodidae) d'Algérie. Cah Entomol médicale Parasitol. 22(2): 99–127.
- 28. El Sawaf B, Kassem HA, El Said S (1985) Description of the hitherto unknown female of *Phlebotomus langeroni* (Diptera: Psychodidae). J Med Entomol. 22 (3): 312–314.
- Depaquit J, Léger N, Ferté H (1998) The taxonomic status of *Phlebotomus sergenti* Parrot, 1917, vector of *Leishmania tropica* (Wright, 1903) and *Phlebotomus similis* Perfiliev, 1963 (Diptera-Psychodidae). Morphologic and morphometric approaches. Biogeographical and epidemiological corollaries. Bull Soc Pathol Exot. 91(4): 346–352.
- 30. Erisoz-Kasap O, Depaquit J, Rahola N,

Haddad N, Gunay F, Akhoundi M, Dvorak V I V, Ozbel Y, Sawalha S, Picard M, Robert V AB (2021) PhlebKeyTool, European Union. Available at: https://www.medilabsecure.com/phlebk eytool.html.

- Dajoz R (1985) Précis d'écologie. Vol. 6. Dunod. Paris.
- 32. Blondel J (1979) Biogéographie et écologie. Vol. 15. Masson. Paris.
- Daget J (1979) Les modèles mathématiques en écologie. Vol. 8. Masson. Paris.
- Southwood TRE (1978) Ecological Methods. Vol. 9. Springer. Dordrecht.
- 35. Wickham H (2007) Reshaping Data with the reshape Package. J Stat Softw. 21 (12): 1–20.
- 36. Wickham H, Averick M, Bryan J, Chang W, McGowan L, François R, Grolemund G, Hayes A, Henry L, Hester J, Kuhn M, Pedersen TL, Miller E, Bache SM, Müller K, Ooms J, Robinson D, Seidel DP, Spinu V, Takahashi K, Vaughan D,Wilke C, Woo K, Yutani H (2019) Welcome to the Tidyverse. J Open Source Softw. 4(43): 1–6.
- 37. Wickham H, Chang W, Wickham MH (2016) Package 'ggplot2.': Create elegant data Vis ualisation Grammar of Graphics, Github, California, USA. Available at: https://github.com/tiduuerse/ggplot2
 - https://github.com/tidyverse/ggplot2.
- 38. Wei T, Simko VR (2021) corrplot: Visualization of a Correlation Matrix (R package), Github, California, USA. Available at:
 - https://github.com/taiyun/corrplot.
- Olivoto T, Lúcio AD (2020) metan: An R package for multi-environment trial analysis. Methods Ecol Evol. 11(6): 783–789.
- Hamiroune M, Selt F, Senni Z, Saidani K, Djemal M (2019) Situation épidémiologique de la leishmaniose cutanée humaine dans la région steppique de Djelfa en Algérie: Incidence et facteurs de va-

riation. Int J Innov Appl Stud. 26(1): 253–261.

- Benallal KE, Garni R, Harrat Z, Volf P, Dvorak V (2022) Phlebotomine sand flies (Diptera: Psychodidae) of the Maghreb region: A systematic review of distribution, morphology, and role in the transmission of the pathogens. PLoS Negl Trop Dis. 16(1): 1–51.
- Izri MA, Belazzoug S, Pratlong F, Rioux JA (1992) Isolation of *Leishmania major* from *Phlebotomus papatasi* in Biskra. Complition of an epidemiological saga. Ann Parasitol Hum Comparée. 67 (1): 31–32.
- Li-Ren G, Yong-Xiang X, Bao-Shan L, Jiang D (1986) The role of *Phlebotomus alexandri* Sinton, 1928 in the transmission of kala-azar. Bull World Health Organ. 64(1): 107–112.
- 44. Kiplagat S, Villinger J, Kigen CK, Kidambasi KO, Muema JM, Mwangi SM, Wangaria M, Matoke-Muhiaa D, Masigaa DK, Bargula JL (2023) Discovery of the vector of visceral leishmaniasis, *Phlebotomus (Artemievus) alexandri* Sinton, 1928, in Kenya suggests complex transmission dynamics. Curr Res Parasitol Vector-Borne Dis. 4(1): 1–10.
- 45. Azizi K, Rassi Y, Javadian E, Motazedian MH, Rafizadeh S, Yaghoobi Ershadi MR, Mohebali M (2006) *Phlebotomus (Paraphlebotomus) alexandri*: a probable vector of *Leishmania infantum* in Iran. Ann Trop Med Parasitol. 100(1): 63–68.
- 46. Naghian A, Oshaghi MA, Moein-Vaziri V, Rassi Y, Sedaghat MM, Mostafavi E, Veysi A, Soleimani H, Dehghan H, Zahraei-Ramazani A, Mirhendi H, Amini MH, Yaghoobi-Ershadi MR, Akhavan AA (2020) Molecular identification of *Leishmania* species in *Phlebotomus alexandri* (Diptera: Psychodidae) in Western Iran. J Arthropod Borne Dis. 14(1): 8–16.
- 47. Bakhshi H, Oshaghi MA, Abai MR, Rassi

Y, Akhavan AA, Sheikh Z, Mohtarami F, Saidi Z, Mirzajani H, Anjomruz M (2013) Molecular detection of *Leishmania* infection in sand flies in borderline of Iran-Turkmenistan: restricted and permissive vectors. Exp Parasitol. 135(2): 382–387.

- 48. Guan LR, Xu YX, Wang G, Jia JX (1990) Observation on the infectivity of *Leishmania donovani* from different areas to *Phlebotomus alexandri* from Xinjiang. Zhongguo ji Sheng Chong xue yu ji Sheng Chong Bing za zhi= Chinese J Parasitol Parasit Dis. 8(2): 104–107.
- 49. Guan LR, Zhou ZB, Jin CF, Fu Q, Chai JJ (2016) Phlebotomine sand flies (Diptera: Psychodidae) transmitting visceral leishmaniasis and their geographical distribution in China: a review. Infect Dis Poverty. 5(15): 1–13.
- 50. Izri MA, Belazzoug S, Boudjebla Y, Dereure J, Pratlong S, Delalbre-Belmonte A, Rioux JA (1990) *Leishmania infantum* Mon-1 isole de *Phlebotomus perniciosus*, en Kabylie (Algerie). Ann Parasitol Hum Comparée. 65(3): 150–150.
- 51. Guessous-Idrissi N, Hamdani A, Rhalem A, Riyad M, Sahibi H, Dehbi F, Bichichi M, Essari A, Berrag B (1997) Epidemiology of human visceral leishmaniasis in Taounate, a northern province of Morocco. Parasite. 4(2): 181–185.
- 52. Rioux JA, Lanotte G, Petter F, Dereure J, Akalay O, Pratlong F, Velez ID, Fikri NB, Maazoun R, Denial M, Jarry DM, Zahaf A, Ashford RW, Cadi-Soussi M, Killick-Kendrick R, Benmansour N, Moreno G, Perieres J, Guilvard E, Zribi M, Kennou MF, Rispail P, Knechli R, Serres E (1986) Les leishmanioses cutanées du bassin Méditerranéen occidental. De l'identification enzymatique à l'analyse éco-épidémiologique. L'exemple de trois foyers, tunisien, marocain et français. IMEEE. 1986(1) : 365–395.
- 53. Guerbouj S, Chemkhi J, Kaabi B, Rahali

A, Ismail R Ben, Guizani I (2007) Natural infection of *Phlebotomus (Larroussius) langeroni* (Diptera: Psychodidae) with *Leishmania infantum* in Tunisia. Trans R Soc Trop Med Hyg. 101(4): 372–377.

- 54. Doha S, Shehata MG (1992) Leishmania infantum MON-98 isolated from naturally infected Phlebotomus langeroni (Diptera: Psychodidae) in El Agamy, Egypt. J Med Entomol. 29(5): 891–893.
- 55. El-Sawaf BM, Beier JC, Hussein SM, Kassem HA, Satter SA (1984) *Phlebotomus langeroni*: a potential vector of kala-azar in the Arab Republic of Egypt. Trans R Soc Trop Med Hyg. 78(3): 421.
- 56. Ajaoud M, Es-sette N, Hamdi S, El-Idrissi AL, Riyad M, Lemrani M (2013) Detection and molecular typing of *Leishmania tropica* from *Phlebotomus sergenti* and lesions of cutaneous leishmaniasis in an emerging focus of Morocco. Parasit Vectors. 6(1): 1–9.
- 57. Oshaghi MA, Rasolian M, Shirzadi MR, Mohtarami F, Doosti S (2010) First report on isolation of *Leishmania tropica* from sandflies of a classical urban cutaneous leishmaniasis focus in southern Iran. Exp Parasitol. 126(4): 445–450.
- 58. Tabbabi A, Bousslimi N, Rhim A, Aoun K, Bouratbine A (2011) First report on natural infection of *Phlebotomus sergenti* with *Leishmania* promastigotes in the cutaneous leishmaniasis focus in southeastern Tunisia. Am J Trop Med Hyg. 85(4): 646–646.
- 59. Tabbabi A (2019) Review of Leishmaniasis in the Middle East and North Africa. Afr Health Sci. 19(1): 1329–1337.
- 60. Dereure J, Velez ID, Pratlong F, Denial M, Lardi M, Moreno G, Serres E, Lanotte G, Rioux JP (1986) La leishmaniose viscérale autochtone au Maroc méridional. Présence de *Leishmania infantum* MON-1 chez le Chien en zone présaharienne. Leishmania Taxonomie et Phylogenese

Applications éco- épidémiologiques. IMEEE, Montpellier.

- 61. Zhioua E, Kaabi B, Chelbi I (2007) Entomological investigations following the spread of visceral leishmaniasis in Tunisia. J Vector Ecol. 32(2): 371–374.
- Kanjanopas K, Siripattanapipong S, Ninsaeng U, Hitakarun A, Jitkaew S, Kaewtaphaya P, Tan-ariya P, Mungthin M, Charoenwong C, Leelayoova S (2013) Sergentomyia (Neophlebotomus) gemmea, a potential vector of Leishmania siamensis in southern Thailand. BMC Infect Dis. 13(333): 1–4.
- 63. Senghor MW, Niang AA, Depaquit J, Ferté H, Faye MN, Elguero E, Gaye O, Alten B, Perktas U, Cassan C, Faye B, Bañuls AL (2016) Transmission of *Leishmania infantum* in the canine leishmaniasis focus of Mont-Rolland, Senegal: Ecological, parasitological and molecular evidence for a possible role of *Sergentomyia* sand flies. Kamhawi S, editor. PLoS Negl Trop Dis. 10(11): 1–17.
- 64. Abbas MAS, Lachheb J, Chelbi I, Louati D, Dachraoui K, Ben Miled S, Zhioua E (2022) Independent circulation of *Leishmania major* and *Leishmania tropica* in their respective sandfly vectors for transmission of zoonotic and chronic cutaneous leishmaniasis co-existing in a mixed focus of central Tunisia. Pathogens. 11 (8): 1–16.
- 65. Remadi L, Farjallah D, Chargui N, Belgacem S, Baba H, Zrieq R, Alzain MA, Babba H, Haouas N (2023) Blood meal analysis and molecular detection of mammalian *Leishmania* DNA in wild-caught *Sergentomyia* spp. from Tunisia and Saudi Arabia. Parasitol Res.122(9): 2181– 2191.
- 66. Souttou K, Sekour M, Gouissem K, Hadjoudj M, Guezoul O, Doumandji S, Denys C (2012) Paramètres écologiques des rongeurs recensés dans un milieu semi aride à Djelfa (Algérie). Alger J Arid

Environ "AJAE.". 2(2): 28-41.

- Guerzou A, Derdoukh W, Guerzou M, Souttou K, Doumandji S (2019) Prédation du hérisson d'Algérie Atelerix algirus (Insectivora, Erinaceidae) sur quelques organismes nuisibles aux végétaux dans les milieux agricoles à Djelfa (Algérie). EPPO Bull. 49(1): 152–158.
- 68. Maia C, Depaquit J (2016) Can *Sergentomyia* (Diptera, Psychodidae) play a role in the transmission of mammal-infecting *Leishmania*?. Parasite. 23(55): 1–8.
- 69. Ticha L, Volfova V, Mendoza-Roldan JA, Bezerra-Santos MA, Maia C, Sadlova J, Otranto D, Volf P (2023) Experimental feeding of *Sergentomyia minuta* on reptiles and mammals: comparison with *Phlebotomus papatasi*. Parasit Vectors. 16(126): 1–9.
- Giraudoux P, Raoul F, Pleydell D, Craig PS (2008) Multidisciplinary studies, systems approaches and parasite eco-epidemiology: something old, something new. Parasite. 15(3): 469–476.
- 71. Lawyer P, Young D (1991) Diapause and quiescence in *Lutzomyia diabolica* (Diptera: Psychodidae). Parassitologia. 33: 353–360.
- 72. Oshaghi MA, Ravasan NM, Javadian E, Rassi Y, Sadraei J, Enayati AA, Vatandoost H, Zare Z, Emami SN (2009) Application of predictive degree day model for field development of sandfly vectors of visceral leishmaniasis in northwest of Iran. J Vector Borne Dis. 46(4): 247–255.
- 73. Talbi FZ, El Ouali Lalami A, Fadil M, Najy M, Ech-Chafay H, Lachhab M, Lotfi S, Nouayti N, Lahouiti K, Faraj C, Idrissi AJ (2020) Entomological investigations, seasonal fluctuations and impact of bioclimate factors of Phlebotomines sand flies (Diptera: Psychodidae) of an emerging focus of cutaneous leishmaniasis in Aichoun, Central Morocco. J Parasitol Res. 2020(1): 1–10.
- 74. Kahime K, Boussaa S, Idrissi ALE, Nham-

mi H, Boumezzough A (2016) Epidemiological study on acute cutaneous leishmaniasis in Morocco. J Acute Dis. 5(1): 41-45.

- 75. Alayat MS, Bendali-Saoudi F, Mahmoudi K, Soltani N (2023) Diversity and spatio-temporal distribution of mosquitoes (Diptera: Culicidae) in the Laghouat arid region (Algerian northern Sahara). Orient Insects. 57(4): 1–26.
- 76. Mozaffari E, Vatandoost H, Rassi Y, Mohebali M, Akhavan AA, Moradi-Asl E, Zarei Z, Zahrai-Ramazani A, Ghorbani E (2020) Epidemiology of visceral leishmaniasis with emphasis on the dynamic activity of sand flies in an important endemic focus of disease in Northwestern Iran. J Arthropod Borne Dis. 14 (1): 97–105.
- 77. Rioux JA, Aboulker JP, Lanotte G, Killick-Kendrick R, Martini-Dumas A (1985) Écologie des leishmanioses dans le sud de la France. Ann Parasitol Hum Comparée. 60(3): 221–229.
- 78. Guernaoui S, Boumezzough A, Laamrani A (2006) Altitudinal structuring of sand flies (Diptera: Psychodidae) in the High-Atlas mountains (Morocco) and its relation to the risk of leishmaniasis transmission. Acta Trop. 97(3): 346–351.
- 79. Abdel-Dayem MS, Annajar BB, Hanafi HA, Obenauer PJ (2012) The potential distribution of *Phlebotomus papatasi* (Diptera: Psychodidae) in Libya based on ecological niche model. J Med Entomol. 49(3): 739–745.
- Salah AB, Kamarianakis Y, Chlif S, Alaya N Ben, Prastacos P (2007) Zoonotic cutaneous leishmaniasis in central Tunisia: spatio temporal dynamics. Int J Epidemiol. 36(5): 991–1000.
- Alten B, Maia C, Afonso MO, Campino L, Jiménez M, González E, Molina R, Bañuls AL, Prudhomme J, Vergnes B, Toty C, Cassan C, Rahola N, Thierry M, Sereno D, Bongiorno G, Bianchi R,

Khoury C, Tsirigotakis N, Dokianakis E, Antoniou M, Christodoulou V, Mazeris A, Karakus M, Ozbel Y, Arserim SK, Kasap OE, Gunay F, Oguz G, Kaynas S, Tsertsvadze N, Tskhvaradze L, Giorgobiani E, Gramiccia M, Volf P, Gradoni L (2016) Seasonal dynamics of Phlebotomine sand fly species proven vectors of Mediterranean leishmaniasis caused by *Leishmania infantum*. PLoS Negl Trop Dis. 10(2): 1–22.

- 82. Sofizadeh A, Akbarzadeh K, Allah Kalteh E, Karimi F (2020) Relationship between the distribution and biodiversity of sand flies (Diptera: Psychodidae) with the incidence of zoonotic cutaneous leishmaniasis in endemic foci of Golestan Province, Iran. J Med Entomol. 57(6): 1768–1774.
- 83. Michelutti A, Toniolo F, Bertola M, Grillini M, Simonato G, Ravagnan S, Montarsi F (2021) Occurrence of *Phlebotomine* sand flies (Diptera: Psychodidae) in the northeastern plain of Italy. Parasit Vectors. 14(164): 1–7.
- 84. Belen A, Alten B (2011) Seasonal dynamics and altitudinal distributions of sand fly (Diptera: Psychodidae) populations in a cutaneous leishmaniasis endemic area of the Cukurova region of Turkey. J Vector Ecol. 36(1): S87–S94.