Journal of Advanced Biomedical Sciences
Homepage: http://jabs.fums.ac.ir
Online ISSN: 2783-1523

Phytochemical and Cytotoxic Properties of Gum and Leaf Extracts of *Ferula assafoetida*

Moulazadeh Alireza[1, 2], Ranjbar Razieh[1], Dakhili Ardestani Amin[2], Ranjbar Kamran[2], Kouhpayeh Seyed Amin[1], Maghbool Maryam[1], Najafipour Sohrab[3, 4] ✉, Najafipour Fatemeh[5]

1. Noncommunicable Disease Research Center, Fasa University of Medical Sciences, Fasa, Iran
2. Student Research Committee, Fasa University of Medical Sciences, Fasa, Iran
3. Medicinal Plants Research Center, Fasa University of Medical Sciences, Fasa, Iran
4. Department of Microbiology, Fasa University of Medical Sciences, Fasa, Iran
5. Ghutbuddin Clinic, Shiraz University of Medical Sciences, Shiraz, Iran

Article Info

Article Type:
Original Article

Article History:
Received
05 Sep 2023
Received in revised form
04 Nov 2023
Accepted
15 Nov 2023
Published online
20 Nov 2023

Publisher:
Fasa University of Medical Sciences

Abstract

Background & Objectives: Breast cancer, a significant threat to women's health, is associated with a high annual mortality rate. Oxidative stress, characterized by the accumulation of reactive oxygen species, plays a pivotal role in cancer development. *Ferula assafoetida*, a traditional medicinal plant, has been widely used in cancer treatment. This study aims to compare the cytotoxic effects, phytochemical profiles, and antioxidant activities of *Ferula assafoetida* gum (FAGE) and leaf (FALE) extracts.

Materials & Methods: The herbal extracts from FAGE and FALE were prepared using the maceration method in 70% ethanol. The phenolic content and radical scavenging activity of the extracts were assessed using the Folin-Ciocalteu and diphenylpicrylhydrazyl (DPPH) assays, respectively. The cytotoxic effects of the extracts were examined on the MDA-MB-231 and MCF-7 cell lines through the MTT assay. Additionally, the morphological changes of the cell lines were observed using an inverted microscope.

Results: The phenolic content in FAGE (142.25± 19.35 µgGAE/mg) was significantly higher (P=0.01) than that in FALE (48.15± 3.01 µgGAE/mg). Additionally, the radical scavenging activity and cytotoxic effects of FAGE were also significantly stronger than those of FALE (P< 0.05). Regarding the cytotoxic effects of the extracts, FAGE exhibited IC50 values of 80.59± 10.98 and 59.39± 15.89 µg/mL for MCF-7 and MDA-MB-231 cells, respectively, after 72 hours of incubation.

Conclusions: FAGE exhibited a greater cytotoxic effect on the MDA-MB-231 cell line than on the MCF-7, possibly following an estrogen-independent pattern. It appears that FAGE holds significant potential in refining oxidative stress, thereby inhibiting cell proliferation.

Keywords: Phenol, Flavonoid, Oxidative stress, Cytotoxicity, Cancer

Cite this article: : Moulazadeh A, Ranjbar R, Dakhili Ardestani A, Ranjbar K, Kouhpayeh SA, Maghbool M, Najafipour S, Najafipour F. Phytochemical and Cytotoxic Properties of Gum and Leaf Extracts of *Ferula assafoetida*. JABS.2023; 13(4): 329-339.
DOI: 10.18502/jabs.v13i4.13904

Introduction

Mutations in critical molecules, such as proteins and DNA, drive cancer development. These alterations disrupt the normal proliferation and growth patterns of cells, leading to uncontrolled cell division and cancer formation. The antioxidant system plays a pivotal role in cancer prevention. Induction of oxidative stress and accumulation of reactive oxygen species (ROS) in the microenvironment of normal cells are crucial factors in cancer initiation. Dysfunction of the antioxidant system can exacerbate the risk of cancer by disrupting physiological balance (1).

Breast cancer arises from the uncontrolled proliferation of cancerous epithelial cells. According to the latest World Health Organization (WHO) data, breast cancer is the most prevalent cancer

✉ **Corresponding Authors: Najafipour Sohrab,** Fasa University of Medical Sciences, Ibn-Sina square, Fasa city, Fars province, Iran
Email: resimmuno@gmail.com

among women, accounting for approximately 25% of all cancer cases globally. In 2020, an estimated 2.3 million new cases of breast cancer were diagnosed worldwide (2). In Iran, breast cancer is the most common cancer among women, with an estimated 11,000 new cases annually. The incidence rate of breast cancer in Iran is also higher than the global average, with an age-standardized rate of 32.1 per 100,000 women compared to the global rate of 25.4 per 100,000 women (3). Consequently, effective treatment of breast cancer is a fundamental priority for healthcare systems, particularly in Iran.

Conventional breast cancer treatments often have various clinical side effects, including therapeutic resistance, damage to healthy organs, cancer recurrence, and high costs, especially for triple-negative tumors. Triple-negative tumors lack expression of the human epidermal growth factor receptor (HER2), the progesterone receptor, and the estrogen receptor on their cell surfaces. These tumors are highly resistant to conventional treatments and can lead to the most aggressive form of breast cancer (4). Therefore, the development of effective herbal drugs with fewer side effects has become a priority for researchers in recent years (5). Certain natural compounds have demonstrated promising anti-cancer properties in laboratory studies and animal models.

Secondary metabolites in plants possess various pharmacological and biochemical properties, including antioxidant and cytotoxic activities (6). Phenols and flavonoids are two major groups of secondary compounds with well-established antioxidant and cytotoxic effects. Ferula assafoetida is a rich source of secondary metabolites, including phenols and flavonoids. Pharmacological studies have demonstrated its antioxidant and cytotoxic properties (7). The use of Ferula assafoetida leaves and gum is widespread among the general public. While some studies have explored the potential anti-cancer effects of certain compounds found in Ferula assafoetida, the evidence remains limited and inconclusive. The present study aimed to compare the cytotoxic effects, phytochemical profiles, and antioxidant activities of Ferula assafoetida leaf and gum extracts.

Materials and Methods

Herbal extraction

Following identification by experts at the Fasa Medicinal Plants Research Center (FMPRC), *Ferula assafoetida* gum and leaf were assigned voucher numbers FMPRC-100-18 and FMPRC-100-19, respectively. The herbal extraction was performed using the maceration method (8). Dried hydroethanolic extracts (500 mg) were dissolved in dimethyl sulfoxide (5 mL). Sterilization of the stock solutions (100 mg/mL) of Ferula assafoetida gum extract (FAGE) and leaf extract (FALE) was accomplished using a 0.22 µm filter. Preparation of a working solution (1 mg/mL) for cell culture and phytochemical evaluations involved 1:100 (v/v) dilution of the sterilized solutions with DMEM and distilled water, respectively (9). Ethical approval for the study was obtained from the Medical Ethics Committee of Fasa University of Medical Sciences (Code: IR.FUMS.REC.1400.166).

The Phenolic content assays

The phenolic content of FAGE and FALE was assessed using the Folin-Ciocalteu assay. Specifically, 500 µL of Folin-Ciocalteu reagent (10% v/v) was added to 100 µL of the extracts at a concentration of 1 mg/mL, followed by an incubation period of 5 minutes at room temperature in the dark. Subsequently, 400 µL of sodium carbonate (7.5% w/v) was introduced to the samples, and the resulting solutions were left at room temperature in darkness for 60 minutes. The absorbance of the samples was measured at 765 nm using a Synergy HTX multi-mode reader. Gallic acid served as the standard, and the phenolic content of the extracts was expressed in micrograms of Gallic Acid Equivalent (GAE) per milligram (µgGAE/mg) (10, 11). The results were

[DOI: 10.18502/jabs.v13i4.13904]

presented as mean ± SD, and all measurements were conducted three times in duplicate.

The power of radical scavenging

In the DPPH clearance assay, the total radical scavenging capacity of the extracts is assessed. The DPPH radical initially possesses a purple color, which transforms to yellow upon reduction by antioxidants. The loss of color indicates the antioxidant activity of the extracts. Various concentrations of the extracts (50, 100, 200, 500, and 1000 µg/mL) were prepared using 70% ethanol. These solutions (40 µL) were combined with 160 µL of DPPH radicals (0.3 mM), thoroughly mixed, and then incubated for 30 minutes at room temperature in darkness. Subsequently, the absorption of the samples was measured at 517 nm using the Synergy HTX multi-mode reader. All measurements were replicated three times in duplicate. The control group comprised 70% ethanol (v/v), which was employed for diluting the essential oils. The percentage of total radical scavenging activity was calculated using the following equation (11-13):

Antioxidant power= [(Optical absorption of the control group - Optical absorption of the experimental group) / Optical absorption of the control group] × 100

Cell cytotoxicity of FAGE and FALE and morphological changes

The MTT assay was employed to evaluate the cell viability of MDA-MB-231 and MCF-7 breast cancer cell lines following exposure to FAGE and FALE. Based on a previous study, 10,000 cells were seeded per well in 150 µL of medium and incubated to form a confluent cell population (14, 15). The cells were then treated with varying concentrations of FAGE and FALE (25-200 µg/mL). The MTT assay procedure followed that of a previous study (16-18). The results are reported as percentages of cell viability. The IC50 values for cell viability were calculated using four-parameter logistic regression. Morphological changes were also evaluated using an inverted microscope (11).

Statistical Analysis

The data are presented as mean ± SD. Statistical analysis was performed using the t-test in GraphPad Prism 8.0.2 software. The IC50 values for cell viability were calculated using four-parameter logistic regression (19).

Results

phenolic content and radical scavenging activity

According to Chart 1, the phenolic content of FAGE was significantly higher (P=0.01) than that of FALE, with values of 142.25± 19.35 µgGAE/mg and 48.15± 3.01 µgGAE/mg, respectively. As shown in Table 1 and Chart 1, there were no statistically significant differences in radical scavenging activity between FAGE and FALE at concentrations of 50 and 100 µg/mL. However, FAGE exhibited significantly higher radical scavenging activity than FALE at concentrations of 200 µg/mL (25.46± 1.99, P= 0.001), 500 µg/mL (45.04± 0.20, P< 0.0001), and 1000 µg/mL (67.70± 1.68, P< 0.0001). Additionally, the IC50 value for radical scavenging activity of FAGE (697.6 µg/mL) was lower than that of FALE (>1000 µg/mL).

[DOI: 10.18502/jabs.v13i4.13904]

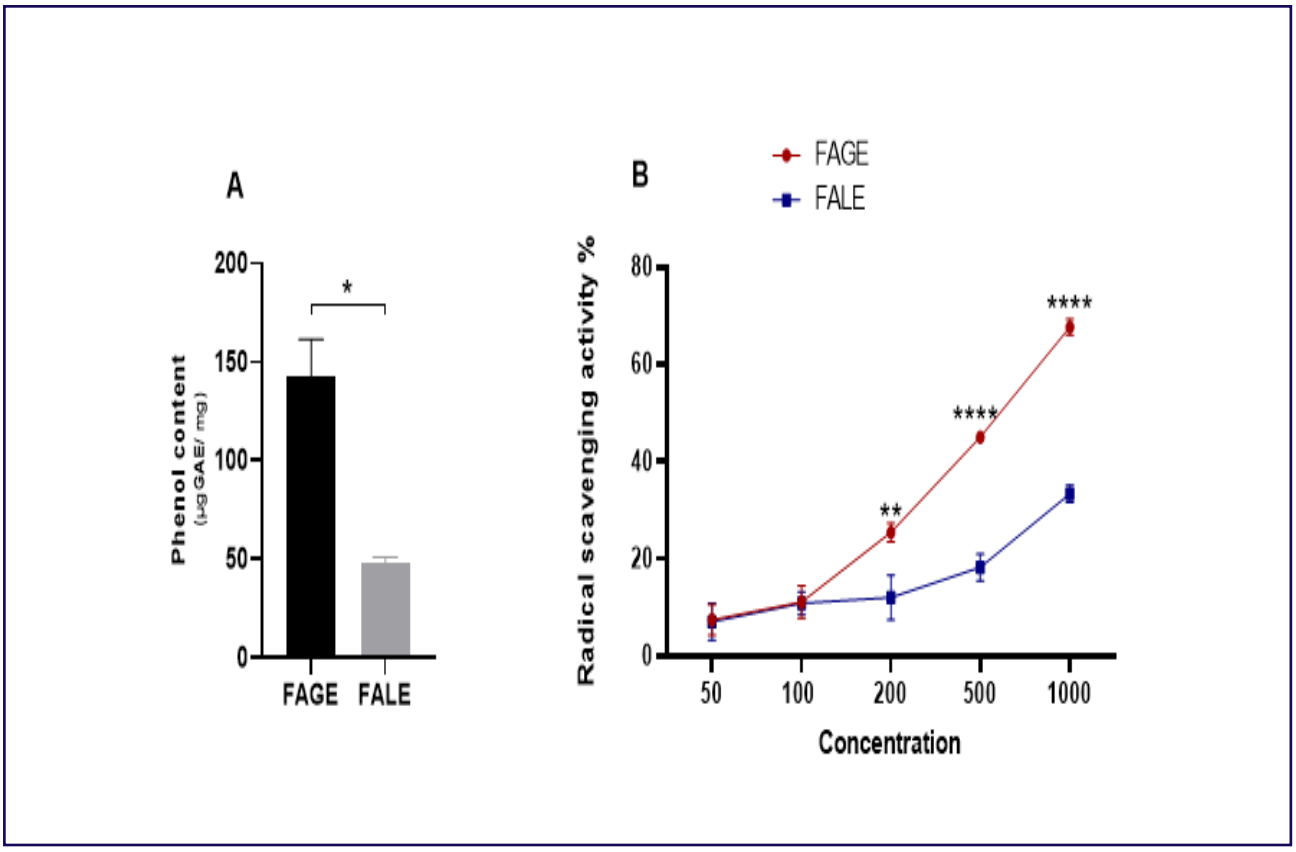

Chart 1. The phenolic content (A) and radical scavenging activity (B) of *Ferula assafoetida* gum extract (FAGE) and leaf extract (FALE)

The (*) bullet indicates the significant differences between the group. *, **, *** and **** respectively indicate $p<0.05$, $p<0.01$, $p<0.001$ and $p<0.0001$

Table 1. Total radical scavenging activity of *Ferula assafoetida* gum extract (FAGE) and leaf extract (FALE) and ascorbic acid

Concentration (µg/mL)	FAGE		FALE		P-value
	Mean± SD	IC50	Mean± SD	IC50	
50	7.45± 3.13	697.6	7.03± 3.78	>1000	0.88
100	11.12± 3.37		10.84± 2.32		0.90
200	25.46± 1.99		12.02± 4.58		0.001
500	45.04± 0.20		18.24± 2.71		<0.0001
1000	67.70± 1.68		33.3± 1.82		<0.0001

[DOI: 10.18502/jabs.v13i4.13904]

Cytotoxicity of the extracts on the MCF-7 cell line

As shown in Table 3, FAGE and FALE significantly reduced the cell viability of the MCF-7 cell line after incubation for 24, 48, and 72 hours. Following 24 hours of incubation, the IC50 values of FAGE and FALE on the MCF-7 cell line were 112.9± 35.82 and >200 (more than 200) µg/mL, respectively. FAGE exhibited significantly higher cytotoxic effects compared to FALE at herbal extract concentrations of 50 (P= 0.004), 100, and 200 µg/mL (P< 0.0001). The highest cytotoxic effect of FAGE was observed at the concentration of 200 µg/mL, reaching 71.90%.

Table 2. Cytotoxic effects of *Ferula assafoetida* gum extract (FAGE) and *Ferula assafoetida* leaf extract (FALE)

Cell line and exposure time		Dose (µg/mL)	FAGE			FALE			P-value
			Viability %		IC_{50}	Viability %		IC_{50}	
			Mean	STD		Mean	STD		
MCF-7	24 h	Control	100.95	4.08	112.9 ± 35.82	100.93	1.31	> 200	0.99
		25	92.30	3.82		97.16	1.10		0.47
		50	86.55	5.03		96.73	2.00		0.004
		100	61.53	6.59		94.17	6.82		< 0.0001
		200	28.10 **	7.66		98.06	0.90		< 0.0001
	48 h	Control	100.01	4.03	98.45 ± 9.8	100.68	2.05	> 200	0.74
		25	94.36	0.36		97.26	1.66		0.36
		50	79.15	8.09		98.16	2.02		0.002
		100	48.65	6.40		87.94	5.65		< 0.0001
		200	8.95	2.20		55.98 ***	4.36		< 0.0001

[DOI: 10.18502/jabs.v13i4.13904]

	72 h	Control	99.46	1.44	80.59 ± 10.98	100.14	2.69	> 200	0.57
		25	88.42	6.76		98.73	0.75		0.04
		50	66.79	6.01		97.75	2.22		< 0.0001
		100	38.82	11.67		96.66	6.56		< 0.0001
		200	5.71	2.60		82.59	7.90		< 0.0001
	24 h	Control	99.83	2.94	173.6 ± 39.51	98.52	2.32	> 200	0.63
		25	88.81	3.05		97.96	2.06		0.01
		50	73.61 ***	5.99		97.50	1.11		< 0.0001
		100	55.82	5.44		95.51	4.93		< 0.0001
		200	39.47	4.71		90.12	6.89		< 0.0001
MDA-MB-231	48 h	Control	98.78	6.89	62.76 ± 7.32	99.18	2.09	> 200	0.87
		25	92.92	6.52		97.51	1.66		0.30
		50	57.81 ***	6.51		97.96	0.28		< 0.0001
		100	27.18 ****	5.09		97.41	7.04		< 0.0001
		200	8.58	0.80		85.82	3.09		< 0.0001
	72 h	Control	98.03	3.41	59.39 ± 15.89	99.39	0.45	> 200	0.63
		25	71.02 **	3.31		99.07	0.38		0.0001
		50	59.73	7.66		98.37	1.40		< 0.0001
		100	21.14 **	6.77		100.57	2.22		< 0.0001
		200	6.70	0.13		80.83	3.74		< 0.0001

The P-value column reports statistical differences between corresponding concentrations of FAGE and FALE through t-test analysis. The (*) bullet also denotes statistical differences between cell lines at similar concentrations and incubation times through t-test analysis. The asterisk is placed on the cell line with lower viability % (indicating higher cytotoxic effects). *, **, ***, and **** respectively indicate $p < 0.05$, $p < 0.01$, $p < 0.001$, and $p < 0.0001$.

[DOI: 10.18502/jabs.v13i4.13904]

After 48 hours of incubation, the IC50 value for FAGE and FALE on the MCF-7 cell line was 98.45 ± 9.8 and >200 µg/mL, respectively. FAGE exhibited higher cytotoxic effects compared to FALE at concentrations of 50 (P = 0.002), 100, and 200 µg/mL (P < 0.0001). The highest cytotoxic effects of FAGE and FALE were observed at the concentration of 200 µg/mL, reaching 91.05% and 44.02%, respectively.

After 72 hours of incubation, the IC50 value for FAGE and FALE on the MCF-7 cell line was 80.59 ± 10.98 and >200 µg/mL, respectively. FAGE demonstrated higher cytotoxic effects compared to FALE at concentrations of 50, 100, and 200 µg/mL (P < 0.0001). The highest cytotoxic effects of FAGE and FALE were observed at the concentration of 200 µg/mL, reaching 94.29% and 17.41%, respectively.

Cytotoxicity of the extracts on the MDA-MB-231 cell line

According to Table 3, both FAGE and FALE significantly reduced the cell viability of MDA-MB-231 after 24, 48, and 72 hours of incubation. After 24 hours of incubation, the IC50 value for FAGE and FALE was 173.6± 39.51 and >200 µg/mL, respectively. FAGE exhibited higher cytotoxic effects compared to FALE at doses of 25 (P= 0.01), 50, 100, and 200 µg/mL (P< 0.0001). The highest cytotoxic effects of FAGE and FALE were observed at the concentration of 200 µg/mL, reaching 60.53% and 9.88%, respectively.

After 48 hours of incubation, the IC50 value for FAGE and FALE was 62.76± 7.32 and >200 µg/mL, respectively. FAGE demonstrated higher cytotoxic effects compared to FALE at concentrations of 50, 100, and 200 µg/mL (P< 0.0001). The highest cytotoxic effects of FAGEand FALE were observed at concentrations of 200 µg/mL, reaching 91.42% and 14.18%, respectively The IC50 value for FAGE and FALE. after 72 hours of incubation was 59.39± 15.89 and >200 µg/mL, respectively. FAGE displayed higher cytotoxic effects compared to FALE at concentrations of 25 (P= 0.0001), 50, 100, and 200 (P< 0.0001). The highest cytotoxic effects of FAGE and FALE were observed at the concentration of 200 µg/mL, reaching 93.3% and 19.17%, respectively.

The cytotoxicity of the extracts between cell lines

The cytotoxic effects of FAGE on the MDA-MB-231 cell line were significantly higher (P=0.0001) than those on the MCF-7 cell line at the 50 µg/mL dose after 24 hours of exposure. However, at the concentration of 200 µg/mL, the cytotoxic effects of FAGE on the MDA-MB-231 cell line were lower than those on the MCF-7 cell line (P=0.0013). After 48 hours of incubation, the cytotoxicity of FAGE on the MDA-MB-231 cell line was significantly higher compared to the MCF-7 cell line at the concentrations of 50 µg/mL (P=0.0001) and 100 µg/mL (P<0.0001). After 72 hours of incubation, the cytotoxicity of FAGE on the MDA-MB-231 cell line was also significantly higher compared to the MCF-7 cell line at the doses of 25 µg/mL (P=0.0065) and 100 µg/mL (P=0.0038). Overall, the cytotoxicity of FAGE on the MDA-MB-231 cell line was higher after incubation times of 48 and 72 hours, respectively, with IC50 values of 62.76± 7.32 µg/mL and 59.39± 15.89 µg/mL. At the 24-hour incubation time, the cytotoxicity of FAGE on the MCF-7 cell line (IC50: 112.9± 35.82 µg/mL) was higher than that on the MDA-MB-231 cell line (IC50: 173.6± 39.51 µg/mL). Regarding FALE, the cytotoxic effects on the MCF-7 cell line were significantly higher than those on the MDA-MB-231 cell line after 48 hours of incubation at the concentration of 200 µg/mL (P=0.0006).

[DOI: 10.18502/jabs.v13i4.13904]

Analysis of cell morphological changes

The morphological changes in the MCF-7 and MDA-MB-231 breast cancer cell lines exposed to FAGE and FALE exhibited a concentration- and time-dependent pattern. The morphological alterations observed in the MDA-MB-231 cell line were more pronounced than those in the MCF-7 cell line. Additionally, FAGE induced more significant morphological changes compared to FALE. The exposure to the herbal extracts led to the rupture of the cell lines, accompanied by a significant increase in cell debris and cell granules.

Discussion

Triple-negative tumors lack the epidermal growth factor receptor 2, progesterone receptor, and estrogen receptor. This tumor type is highly resistant to common treatments and carries a poor prognosis. Ongoing efforts to discover alternative effective therapies are focusing on triple-negative cell lines such as MDA-MB-231 (20). In contrast, MCF-7 possesses the estrogen receptor (ER), making it an ER-positive tumor cell line, unlike the ER-negative MDA-MB-231. The current study revealed that FAGE and FALE exerted higher cytotoxic effects on the MDA-MB-231 triple-negative cell line compared to the MCF-7 cell line. Thus, the cytotoxic effects of FAGE and FALE appear to be independent of estrogen receptors, suggesting a potential therapeutic impact on triple-negative tumors.

The IC50 values of FAGE on the MDA-MB-231 cell line were 62.76± 7.32 and 59.39± 15.89 µg/mL after 48 and 72 hours of incubation, respectively. Consequently, FAGE is considered a relatively active compound according to the Baharum et al. criteria for the classification of anti-cancer herbal medicinal plants (21). The IC50 values of FALE were >200 µg/mL after 48 and 72 hours of incubation, categorizing FALE as a weakly active compound. On the MCF-7 cell line, FAGE induced a higher decrease in cell viability or greater cytotoxic effects. The IC50 values of FAGE were 98.45± 9.8 and 80.59± 10.98 µg/mL on the MCF-7 cell line after 48 and 72 hours of incubation, respectively, confirming its relatively active nature on the MCF-7 cell line.

The heightened cytotoxic effects of FAGE are likely attributed to its higher radical scavenging activity, more potent reducing power, and greater phenolic and flavonoid contents. FAGE appears to possess a substantial capacity for improving oxidative stress, a factor in tumor progression that disrupts the microenvironment of normal cells, leading to changes in physiological parameters such as the suppression of inhibitory signals, production of growth signals, and angiogenesis. Natural antioxidants like polyphenols and flavonoids exhibit high efficacy in improving oxidative stress (22). It is plausible that the polyphenols and flavonoids in FAGE inhibit cancer progression through oxidative stress improvement, whereas FALE, with lower phenolic and flavonoid content, exhibits comparatively lower cytotoxic effects.

In contrast to the present study, Niazmand et al. reported that FALE has higher phenolic and flavonoid content and superior antioxidant effects compared to FAGE (23). This difference may be attributed to variances in the extraction method. Niazmand et al. immersed Ferula assafoetida leaf and gum for 3 hours in the solvent, while the present study employed a one-week incubation period. This prolonged incubation likely led to a more comprehensive extraction, resulting in the extraction of a greater number of compounds, as confirmed by the phenolic and flavonoid content results. Additionally, *Ferula assafoetida* gum exhibits different compositions depending on the time of harvest and climatic conditions, presenting in white, pink, brown, and black colors. Hence, the gum used in this study may have had diverse compositions (24).

Some studies have reported the cytotoxic and anticancer effects of isolated bioactive components from *Ferula assafoetida*. Hasanzadeh et al. documented the cytotoxic and antioxidant effects of Farnesiferol C, one of the major compounds found in *Ferula assafoetida* (36). Kim et al. reported the anti-tumor effect of isolated Galbanic

[DOI: 10.18502/jabs.v13i4.13904]

acid from *Ferula assafoetida* (37). Iranshahy et al. also reported the cytotoxic effect of Gummosin on PC-3 and MCF-7 cell lines. Gummosin, a sesquiterpene coumarin isolated from *Ferula assafoetida*, exhibited cytotoxic effects with IC50 values of 32.1 and 30 µg/mL on MCF-7 and PC-3 cell lines, respectively (38).

To the best of our knowledge, this study is the first to compare the cytotoxic effects of leaf and gum extracts of *Ferula assafoetida* on breast cancer cell lines. FAGE demonstrated more potent cytotoxic effects, likely in an estrogen-independent pattern. It is recommended that future studies evaluate the efficacy of different methods of gum extraction. One of the most significant limitations of the present study was the failure to investigate the cytotoxic effects of FAGE and FALE on non-cancerous cells.

Conclusion

The rising prevalence of breast cancer and the increasing resistance to conventional therapies underscore the urgent need for novel therapeutic approaches. This study demonstrates that FAGE exhibits significantly higher cytotoxic effects compared to FALE, likely due to its richer phytochemical content and enhanced antioxidant activity. Based on the classification criteria for anticancer compounds, FAGE is considered a relatively active agent. FAGE displayed a more pronounced cytotoxic effect on the MDA-MB-231 cell line compared to MCF-7, suggesting an estrogen-independent mechanism of action. Moreover, the cytotoxic effects of FAGE intensified with increasing concentration and incubation time. Therefore, FAGE emerges as a promising drug candidate for the treatment of triple-negative tumors.

Acknowledgment

This research endeavor was generously supported by the esteemed Vice-Chancellor for Research of Fasa University of Medical Sciences. The authors extend their heartfelt gratitude to the Clinical Research Development Unit for their invaluable contributions. Furthermore, the authors acknowledge the unwavering cooperation and assistance provided by the Noncommunicable Diseases Research Center of Fasa University of Medical Sciences. Special recognition is extended to Ms. Mahboubeh Bordbar and Mrs. Soroush Dadvari for their exceptional dedication and support.

Conflict of Interests

None.

Funding

Fasa University of Medical Sciences supported this study, Grant No. 400232.

Ethical Considerations

Non related.

Code of Ethics

IR.FUMS.REC.1400.166.

Author's Contributions

AM assumed the responsibility for conceptualizing and designing the study, conducting the MTT assays, and meticulously analyzing the gathered data. RR diligently carried out the MTT assays and executed the phytochemical analyses with utmost precision. ADA meticulously performed the phytochemical analyses, ensuring the accuracy and reliability of the results. KR provided invaluable assistance in the extraction of plant materials and the execution of phytochemical analyses. AK made significant contributions to the study design, providing expert guidance and direction. MM collaborated extensively in the preparation of the manuscript, lending their expertise to enhance the clarity and coherence of the writing. SN initiated the study and played a pivotal role in its design, providing the foundation for the research endeavor. All authors actively participated

[DOI: 10.18502/jabs.v13i4.13904]

in drafting the manuscript, meticulously crafting each section to convey the findings effectively. Finally, all authors thoroughly reviewed and approved the final manuscript, ensuring the accuracy, completeness, and overall quality of the research presentation.

References

1. Dizdaroglu M. Oxidatively induced DNA damage and its repair in cancer. Mutation Research/Reviews in Mutation Research. 2015;763:212-45.
2. Ferlay J, Colombet M, Soerjomataram I, Parkin DM, Piñeros M, Znaor A, et al. Cancer statistics for the year 2020: An overview. International journal of cancer. 2021;149(4):778-89.
3. Kazeminia M, Salari N, Hosseinian-Far A, Akbari H, Bazrafshan M-R, Mohammadi M. The prevalence of breast cancer in Iranian women: a systematic review and meta-analysis. Indian Journal of Gynecologic Oncology. 2022;20(1):14.
4. Labidi S, Mejri N, Lagha A, Daoud N, El Benna H, Afrit M, et al. Targeted therapies in HER2-overexpressing metastatic breast cancer. Breast Care. 2016;11(6):418-22.
5. Waks AG, Winer EP. Breast cancer treatment: a review. Jama. 2019;321(3):288-300.
6. Nie J, Zhao C, Deng L, Chen J, Yu B, Wu X, et al. Efficacy of traditional Chinese medicine in treating cancer. Biomedical reports. 2016;4(1):3-14.
7. Karimian V, Sepehry A, Barani H, Ebrahimi SN, Mirjalili MH. Productivity, essential oil variability and antioxidant activity of Ferula assa-foetida L. oleo-gum-resin during the plant exploitation period. Journal of Essential Oil Research. 2020;32(6):545-55.
8. Ahmadi E, Abdollahi A, Najafipour S, Meshkibaf MH, FasihiRamandi M, Namdar N, et al. Surveying the effect of the phenol compounds on antibacterial activity of herbal extracts: In vitro assessment of herbal extracts in Fasa-Fars province. Journal of Fasa University of Medical Sciences. 2016;6(2):210-20.
9. Vongsak B, Sithisarn P, Mangmool S, Thongpraditchote S, Wongkrajang Y, Gritsanapan W. Maximizing total phenolics, total flavonoids contents and antioxidant activity of Moringa oleifera leaf extract by the appropriate extraction method. Industrial crops and products. 2013;44:566-71.
10. Moulazadeh A, Ranjbar R, Dakhili Ardestani A, Najafipour S. Evaluation of antioxidant activity and cytotoxic effects of Hypnea musiformis on MCF7 and MDA-MB-231 cell lines. Iranian Journal of Pharmaceutical Sciences. 2021;17(4):33-46.
11. Moulazadeh A, Ranjbar R, Dakhili Ardestani A, Ranjbar K, Farjadfar A, Kouhpayeh SA, et al. Cytotoxic effects of Trachyspermum ammi and Ferula assafoetida on MCF-7 and MDA-MB-468 breast cancer cell lines. Beni-Suef University Journal of Basic and Applied Sciences. 2022;11(1):147.
12. Ranjbar K, Moulazadeh A, Dakhili Ardestani A, Soleimanian M, Meshkibaf Z, Meshkibaf MH. Antinociceptive and antioxidant effects of Onosma platyphyllum riedl extract. Physiology and Pharmacology. 2022;26(3):322-32.
13. Makoolati Z, Bahrami H, Zamanzadeh Z, Mahaldashtian M, Moulazadeh A, Ebrahimi L, et al. Efficacy of Ficus carica leaf extract on morphological and molecular behavior of mice germ stem cells. Animal Reproduction. 2022;19(1):1-18.
14. Moulazadeh A, Kouhpayeh SA. Suitable Concentration of Anti-Inflammatory Herbal Extracts in Cell Culture. Journal of Fasa University of Medical Sciences. 2020;10(3):2396-9.
15. Moulazadeh A, Ranjbar R, Dakhili Ardestani A, Keshavarzi A, Karimzadeh F, Rahnavard M, et al. Evaluation of Phenolic Content, Antioxidant Activity and Cytotoxic Effects of Ulva lactuca and Hypnea musiformis Marine Algae on MDA-MB-468 Cell Line. Journal of Advanced Biomedical Sciences. 2021;11(3):3921-8.
16. Moulazadeh A, Ranjbar R, Hekmat M, Sedaghat F, Yousefzadi M, Najafipour S. Comparison the cytotoxic effects of Ulva fasciata and Ulva lactuca on the MCF-7 and MDA-MB-231 breast cancer cell lines. Physiology and Pharmacology. 2021;25:373-83.
17. Rahmani H, Ghanbariasad A, Meshkibaf MH, Molazade A, Heiran R, Safari M, et al. Chitosan nanoparticles containing α-pinene and Rosmarinus officinalis L. essential oil: effects on human melanoma cells' viability and expression of apoptosis-involved genes. Polymer Bulletin. 2023:80(5):1-19.
18. Moulazadeh A, Kouhpayeh SA, Ranjbar R, Dakhili Ardestani A, Hekmat M, Azarnia S, et al. Antioxidant activity, phenolic and flavonoid content of Lawsonia inermis and Haplophyllum vermiculare. Physiology and Pharmacology. 2021;25(3):261-9.
19. Chen Z, Bertin R, Froldi G. EC50 estimation of antioxidant activity in DPPH assay using several statistical programs. Food chemistry. 2013;138(1):414-20.
20. Lebert J, Lester R, Powell E, Seal M, McCarthy J. Advances in the systemic treatment of triple-negative breast cancer. Current oncology. 2018;25(Suppl 1):S142.
21. Baharum Z, Akim AM, Taufiq-Yap YH, Hamid RA, Kasran R. In vitro antioxidant and antiproliferative activities of methanolic plant part extracts of Theobroma cacao. Molecules. 2014;19(11):18317-31.
22. do Carmo MAV, Pressete CG, Marques MJ, Granato D, Azevedo L. Polyphenols as potential antiproliferative agents: Scientific trends. Current

[DOI: 10.18502/jabs.v13i4.13904]

Opinion in Food Science. 2018;24:26-35.
23. Niazmand R, Razavizadeh BM. Ferula asafoetida: chemical composition, thermal behavior, antioxidant and antimicrobial activities of leaf and gum hydroalcoholic extracts. Journal of Food Science and Technology. 2021;58(6):2148-59.
24. Kavoosi G, Rowshan V. Chemical composition, antioxidant and antimicrobial activities of essential oil obtained from Ferula assa-foetida oleo-gum-resin: effect of collection time. Food chemistry. 2013;138(4):2180-7.
25. Akbarian A, Rahimmalek M, Sabzalian M. Variation in fruit morphological traits and bioactive compounds in different populations of Ferula assa-foetida, F. gummosa, and F. ovina collected from Iran. Journal of Agricultural Science and Technology. 2018. 19(2):425-38.
26. Safari R, Hoseinifar SH, Nejadmoghadam S, Jafar A. Transciptomic study of mucosal immune, antioxidant and growth related genes and non-specific immune response of common carp (Cyprinus carpio) fed dietary Ferula (Ferula assafoetida). Fish & Shellfish Immunology. 2016;55:242-8.
27. Bagheri SM, Yadegari M, Behpur M, Javidmehr D. Antilithiatic and hepatoprotective effects of Ferula assa-foetida oleo-gum-resin on ethylene glycol-induced lithiasis in rats. Urological Science. 2018;29(4):180.
28. Bagheri SM, Javidmehr D, Ghaffari M, Ghoderti-Shatori E. Chemical compositions and antiproliferative effect of essential oil of asafoetida on MCF7 human breast cancer cell line and female wistar rats. Cancer Translational Medicine. 2020;6(2):34.
29. Ahmadvand H, Amiri H, Elmi ZD, Bagheri S. Chemical composition and antioxidant properties of Ferula-assa-foetida leaves essential oil. Iranian Journal of Pharmacology & Therapeutics. 2013;12(2):52-7.
30. Zia-Ul-Haq M, Shahid SA, Ahmad S, Qayum M, Khan I. Antioxidant potential of various parts of Ferula assafoetida L. Journal of Medicinal Plants Research. 2012;6(16):3254-8.
31. Dehpour AA, Ebrahimzadeh MA, Fazel NS, Mohammad NS. Antioxidant activity of the methanol extract of Ferula assafoetida and its essential oil composition. Grasas y aceites. 2009;60(4):405-12.
32. Bagheri SM, Asl AA, Shams A, Mirghanizadeh-Bafghi SA, Hafizibarjin Z. Evaluation of Cytotoxicity Effects of Oleo-Gum-Resin and Its Essential Oil of Ferula assa-foetida and Ferulic Acid on 4T1 Breast Cancer Cells. Indian Journal of Medical and Paediatric Oncology. 2017;38(02):116-20.
33. Bagheri S, Javidmehr D, Ghaffari M, Ghoderti -Shatori E. Chemical compositions and antiproliferative effect of essential oil of asafoetida on MCF7 human breast cancer cell line and female wistar rats. Cancer Translational Medicine. 2020;6(2):34-9.
34. Sadooghi SD, Nezhad Shahrokh Abadi K, Zafar Balanzhad S. Investigating the cytotoxic effects of ethanolic extract of Ferula assa-foetida resin on HepG2 cell line. KAUMS Journal (FEYZ). 2013;17(4):323-30. [In Persian]
35. Bagheri SM, Abdian-Asl A, Moghadam MT, Yadegari M, Mirjalili A, Zare-Mohazabieh F, et al. Antitumor effect of Ferula assa foetida oleo gum resin against breast cancer induced by 4T1 cells in BALB/c mice. Journal of Ayurveda and integrative medicine. 2017;8(3):152-8.
36. Hasanzadeh D, Mahdavi M, Dehghan G, Charoudeh HN. Farnesiferol C induces cell cycle arrest and apoptosis mediated by oxidative stress in MCF-7 cell line. Toxicology reports. 2017;4:420-6.
37. Kim K-H, Lee H-J, Jeong S-J, Lee H-J, Lee E-O, Kim H-S, et al. Galbanic acid isolated from Ferula assafoetida exerts in vivo anti-tumor activity in association with anti-angiogenesis and anti-proliferation. Pharmaceutical research. 2011;28(3):597-609.
38. Iranshahy M, Farhadi F, Paknejad B, Zareian P, Iranshahi M, Karami M, et al. Gummosin, a sesquiterpene coumarin from Ferula assa-foetida is preferentially cytotoxic to human breast and prostate cancer cell lines. Avicenna Journal of Phytomedicine. 2019;9(5):446.

[DOI: 10.18502/jabs.v13i4.13904]