

Tehran University of Medical Sciences Publication http://tums.ac.ir

# **Iran J Parasitol**

Open access Journal at http://ijpa.tums.ac.ir



Iranian Society of Parasitology http://isp.tums.ac.ir

# **Review Article**

# More than Seventy Years of Research (1948 –November 2021) on *Toxoplasma gondii* in Iran: A Narrative Review

Mitra Sadeghi <sup>1,2</sup>, Seyed Abdollah Hosseini <sup>1,2</sup>, Shahabeddin Sarvi <sup>1,2</sup>, Tooran Nayeri <sup>1,2</sup>, Mehdi Sharif <sup>3</sup>, Abdol Sattar Pagheh <sup>4</sup>, Afsaneh Amouei <sup>1,2</sup>, Mahbobeh Montazeri <sup>1,2</sup>, \*Ahmad Daryani <sup>1,2</sup>

1. Toxoplasmosis Research Center, Communicable Diseases Institute, Mazandaran University of Medical Sciences, Sari, Iran

Department of Parasitology, Mazandaran University of Medical Sciences, Sari, Iran
Departments of Parasitology, Sari Branch, Islamic Azad University, Sari, Iran

4. Infection Diseases Research Center, Birjand University of Medical Sciences, Birjand, Iran

Received 15 Jan 2021 Accepted 18 Mar 2022

*Keywords:* Review; *Toxoplasma gondii;* Iran

\*Correspondence Email: daryanii@yahoo.com

#### Abstract

In this review, we intend to provide a summary of the activities of researchers in the field of *Toxoplasma gondii* in Iran, during the past 70 years. Most studies have been limited to epidemiological studies (mostly using ELISA and IFA methods). Designing a standard and reliable method using the specific antigens of this parasite is essential. So far, studies in the field of drug effects have not been able to introduce an effective drug with few side effects. Various types of vaccines have been developed, such as recombinant and DNA vaccines. However, none of them had a good efficacy. The use of multi-epitope vaccines as potential vaccines against toxoplasmosis is recommended. At present, limited studies have been conducted on the patterns of transmission and genetic diversity of isolated isolates in Iran. Future research to determine the genotype of *T. gondii* could play an important role in the study of population structure, and biological characteristics of this parasite. It is hoped that the results of this study will help control, prevent, and reduce the burden of disease caused by this parasite.

# Introduction

*oxoplasma gondii* is an obligate intracellular parasite and infection a wide range of hosts, containing humans and ani-

mals. *T. gondii* infects about one third of the world's population (1-4). Infection with *T. gondii* is acquired by eating food contaminated



Copyright © 2022 Sadeghi et al. Published by Tehran University of Medical Sciences. This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International license. (https://creativecommons.org/licenses/by-nc/4.0/). Non-commercial uses of the work are permitted, provided the original work is properly cited

Available at: <u>http://ijpa.tums.ac.ir</u>

with sporulated oocysts and consuming of raw or undercooked meat infected to tissue cysts. Vertical transmission also occurs, and iatrogenic transmission may happen via organ transplant (5-7).

Considering the importance of this parasite in the congenital toxoplasmosis, the development of opportunistic infection in immunodeficiency individuals, as well as climatic and social conditions in Iran, it is necessary to provide comprehensive information about the situation of toxoplasmosis. In Iran, various studies have been conducted in the fields of epidemiology, pathogenesis, treatment, vaccines preparation, etc. However, a review study is necessary to collect and provide a summary of relevant data to provide proper solutions for the prevention, control, diagnosis and treatment of toxoplasmosis in Iran. It also makes future studies more targeted.

We aimed to give a summary of toxoplasmosis status in Iran from 1948 through 2021 in various fields of epidemiology, determination of genotypes, diagnosis, drug treatment, and vaccine, as well as the role of the environment and food factors in the transmission of infection.

#### Search strategy and primary finding

To collect data, a comprehensive search was performed on all scientific publications in 4 English (PubMed, ScienceDirect, Google Scholar and Scopus) and 4 Persian databases (Magiran, IranDoc, IranMedex and SID) from 1948 to Nov 2021. The search applied the following keywords: "Toxoplasma gondii", "toxoplasmosis", and" Iran". The evaluation of studies was performed according to the inclusion and exclusion criteria. Inclusion criteria: (a) All English and Persian studies related to T. gondii with Iranian Affiliation. Exclusion criteria: (a) All studies related to T. gondii with non-Iranian affiliation; (b) Articles published in non-English and non-Persian languages; (c) Papers presented at scientific conferences and congresses.

# Overview of conducted articles based on subject

The articles published on *T. gondii* from 1948 to 2021 numbered about 1540, included 296 (19.22%) in Persian and 1244 (80.78%) in English languages. Frequency of published papers based on subject in Iran is shown in Fig. 1.



Fig. 1: Frequency of published papers based on subject in Iran since 1948-November 2021

#### *History*

The population of Iran in 2021 was estimated over 83 million (https://www.amar.org.ir/). Iran has four geographical regions comprising 31 provinces. Also, Iran has a long history of research on *T. gondii*, dates back to 1948, and Ansari reported the first case of the disease. The first research on toxoplasmosis in Iran was performed using Sabin-Feldman test in 1954 (1). Jamalian *et al.* conducted the first clinical trial of *T. gondii* in 1973 (published 1974) (2). Presence of anti- *T. gondii* antibodies in Iranian society has been reported in several studies. The results indicate difference in prevalence of infection in different geographical areas; the lowest rate (12%) was in Khuzestan and the highest rate in Mazandaran (87.5%) (3).

#### Epidemiological studies

Epidemiological studies (868) covered 616 (70.97%) human subjects and 252(29.03%) animal subjects.

#### Human Epidemiological studies

Epidemiological studies of *T. gondii* on human (616/1540;4%) are divided into two groups: healthy group (56.81%) including pregnant women, girls before marriage, blood donors, etc., and the clinical cases (43.19%) including AIDS, cancer, Parkinson, schizophrenia, etc. (3-7) (Table 1).

Group	Epidemiological studies	NO. articles	% of study	% T. gondii
Human	General population	186/616	30.2	39.3
	Pregnant women	133/616	21.6	41
	Neonates			0.64
	Infants Suspected of CT	18/616	2.92	4.1
	Blood donor	24/616	3.9	34.4
	Cancer	30/616	4.88	45.06
	HIV+	34/616	5.51	50.05
	Parkinson	3/616	0.49	53 to 85
	Hemodialysis	29/616	4.70	58
	Diabetic	15/616	2.43	35.1 to 60.43
	Schizophrenia	14/616	2.27	34 to 72.5
	Alzheimer	5/616	0.81	61.3 to 66.6
	Ocular	10/616	1.62	8.36 to 82.5
	Mental disorders	32/616	5.2	-
Animals	Sheep and goats	97/252	38.5	31 and 27
	Cattle	38/252	15.07	18.1
	Cats	33/252	13.1	33.6
	Birds	22/252	8.73	16.52
	Rodents	16/252	6.35	15
	Camels	7/252	2.78	9.93 to 28.06
	Dogs	10/252	3.97	10.1 to 77
	Horses	8/252	3.17	13.3 to 71.2
	Cold-blooded	3/252	1.19	Fish (10), Snakes (80.88), seals (83)

Table 1: Frequency of human and animal epidemiological studies in Iran

#### Animal Epidemiological studies

Overall, 252 studies have been performed on the prevalence of toxoplasmosis in animals, as shown in Table 1 (8-11).

#### Animal food products

Twenty studies (1.3%) have been conducted on food including meat, milk and animal products as well as eggs. In the meantime, 10 studies were performed on meat which showed prevalence rate of4-24% (12, 13). Moreover, 9 studies were performed on milk samples that prevalence was reported 5.4%-11.38% (14, 15) and there was also a study on domestic and industrial eggs. Based on the results of this study, *T. gondii* DNA was detected in 11% of the eggs (16).

#### Environment

There have been few studies on the contamination of the soil and water with *T. gondii* oocysts in Iran. Four studies have been performed to diagnose presence of *T. gondii* DNA in soil and water in Tehran, Ahvaz, Arak, Guilan and Mazandaran provinces (prevalence 5.8% to 78.1%) (17-21).

# Development of diagnostic approach of T. gondii

Iranian scientists have conducted several studies (10.39%) to develop diagnostic methods for *T. gondii* infection. Diagnostic tests are mainly three groups, including serological tests detecting *T. gondii* antibodies, and antigens, and molecular methods (DNA detection) (22).

In the last two decades, Iranian researchers have studied the effects of various *T. gondii* antigens, including recombinant antigens on the improvement of serological test, and difference between acute and chronic infection. SAG1, SAG2, SAG3, GRA2, GRA6, GRA7, GRA8, ROP1, SRS3, and ESA are among the antigens examined whose details are presented in Table 2. Overall, the results showed that the use of antigens such as SAG1, GRA7, and GRA6 for serodiagnosis of acute and chronic toxoplasmosis in human sera could be very effective. Javadi et al. (2020) conducted a bioinformatics study on multi-epitope antigens SAG1, GRA7 and ROP1 showed that the use of the antigens can be considered as a diagnostic kit for acute and chronic Toxoplasmosis (23).

# The T. gondii genetic diversity and its population structures in Iran

The genetic characteristics of the reported studies in Iran are summarized in Table 3. In total DNA or isolates separated in 270 animals, 145 humans, 66 meats, and 31 soil samples were analyzed in the 20 studies in Iran. The isolates were often typed by PCR-RFLP based on SAG2 and GRA6 genes. The typing results based on PCR-RFLP and microsatellite markers among the 505 DNA or isolates separated from different hosts revealed Type III (32.62%) had the highest frequency, mix and atypical types had the lowest frequency in Iran. Out of the remaining samples, Type I and II had the frequency rates of 31.64% and 26.76%, respectively. There are no reports on the genetic diversity with more than five typing markers, and only one study has been done by Hosseini et al. (2019) on HIV+ patients with multilocus PCR-RFLP method using 12 genes (44). It is believed that using low or limited markers would not provide reliable results on population structures (45). However, identification of the genetic groups could provide a key role to play in studies on the population structure, epidemiology and biological characteristics of T. gondii.

References	Method	Gen/Peptide	Sensitivity	Specificity
			(%)	(%)
(24)	ELISA	GRA2	100 (IgG)	96.4 (IgG)
			71.4 (IgM)	
(25)	ELISA	GRA6	87.5 (IgG)	94.1 (IgG)
(26)	ELISA	SAG1,30KD	94.5 (IgG)	93.6 (IgG)
(27)	ELISA	ESA	84 (IgG)	92 (IgG)
(28)	ELISA	SAG1	88.4 (IgG)	88 (IgG)
(29)	ELISA avidity	GRA6	85 (IgG)	100 (IgG)
			100 (IgM)	
(30)	ELISA	SAG1	100 (IgG)	96 (IgG)
(31)	ELISA	GRA7	89 (IgG)	90 (IgG)
			96 (IgM)	90 (IgM)
(32)	ELISA	SAG1	93 (IgG)	95 (IgG)
( )			87 (IgM)	
(33)	ELISA	SAG1	80 (IgM)	90 (IgM)
(34)	ELISA	SAG1	93.6 (IgG)	92.9 (IgG)
			39.3 (IgM)	80 (IgM)
		SAG2	100 (IgG)	89.4 (IgG)
			64.3 (IgM)	83.3 (IgM)
		SAG3	95.4 (IgG)	91.2 (IgG)
			17.9 (IgM)	76.7 (IgM)
(35)	ELISA	SAG1	94.1 (IgG)	100 (IgG)
()			100 (IgM)	(8-)
(36)	ELISA	r ROP1		
(37)	ELISA	r GRA7	92 (IgG)	94 (IgG)
(38)	ELIS Avidity	GRA8	84.6 (IgG)	-
		01210	86.9 (IgM)	
(39)	ELISA	r SAG1	92 (IgG)	96 (IgG)
(40)	ELISA	RT-SRS3	82.89 (IgG)	91 (IgG)
()			100 (IgM)	(-8-)
(41)	Flow immune	r GRA7	100 (IgG)	96.7 (IgG)
(11)	assay	1 0101	100 (180)	(180)
(42)	Dot-Elisa	r SAG1	83.7 (IgG)	90.2 (IgG)
()	25 01 21100	101101	81.2 (IgM)	89.3 (IgM)
			(1811)	0710 (1811)
		r GRA7	66.2 (IgG)	81.2 (IgG)
			87.5 (IgM)	83.9 (IgM)
		r SAG1+rGRA7	86.2 (IgG)	91.1 (IgG)
			90.6 (IgM)	92 (IgM)

Table 2: Summary of antigen/peptides used in design of ELISA kit by researchers in Iran

Ref.	Host	No. of DNA/ Isolates	Molecular marker(s)	No. Type I (%)	No. Type II (%)	No. Type III (%)	No. mix and atypical (%)
(46)	Human	13	SAG2	1 (7.6)	12	-	-
	Rodent	8		2 (25)	(92.4)		
(47)	Sheep	8 4	TUB2, W35, TgM-A, B18,	2 (23) -	6 (75) 2	- 2	-
(17)	Bird	7	and B17	-	-	7	_
	Cat	2		-	2	_	_
	Human	3		_	2	1	_
(48)	Aborted	12	B1	12	-	-	_
(10)	fetuses	12		(100)			
	(Sheep)						
(12)	Meat	40	SAG2	-	40	-	-
	product				(100)		
(49)	Aborted	0	GRA6	-	-	-	-
	fetuses						
	(Ovine)						
(17)	Soil	13	SAG2	1 (7.7)	-	8	4; mix
						(61.5)	I&III
					_		(30.8)
(50)	Bird	41	GRA6	-	8	33	-
(51)	A1 / 1	<b>7</b> 5	SAC2	11	(19.5)	(80.5)	
(51)	Aborted fetuses	65	SAG2	11 (16.9)	54 (83.1)	-	-
	(Hu-			(10.9)	(65.1)		
	(11u- man)						
(52)	Human	52	GRA6	-	-	52	_
(0-)	110111011		01210			(100)	
(18)	Soli	18	GRA6	-	6	12	-
					(33.3)	(66.7)	
(53)	Wild	5	Sequencing with 529	1 (20)	-	4 (80)	-
	boar						
(54)	Aborted	5	GRA6	5 (100)	-	-	-
	fetuses						
	(Ovine)						
(55)	Cat	35	SAG2	1 (2.9)	-	32	2; mix
	A1 / 1	10		2 (20)	2(20)	(91.4)	I&III (5.7)
(56)	Aborted	10	SAG2, SAG3, and GRA6	3 (30)	2 (20)	-	5 atypical
	fetuses (Ewe)						(50)
(13)	(Ewe) Chicken	4	SAG2	4 (100)			
(13)	meat	-	5/102	+ (100)	-	-	-
	Beef	8		8 (100)	-	-	_
	meat	Ŭ		0 (100)			
	Lamb	14		14	_	-	-
	meat			(100)			
(57)	Aborted	2	SAG3, GRA6	-	-	2	-
. ,	fetuses					(100)	
	(Hu-						
	man)						

Table 3: The genetic characterization of Toxoplasma gondii isolates from different hosts in Iran

(58)	Sheep	125	SAG2, and GRA6	90 (72)	-	3 (2.4)	9 mix I&II (7.2), 21 I&III (16.8), 1 II&III (0.8), 1 I,II,III
(59)	Hooded	9	GRA6	_	_	9	(0.8)
(3))	crow		Gialo			(100)	
(44)	AIDS	10	SAG1,SAG2,SAG3,alt- SAG2,BTUB,GRA6,Apico ,PK1,C22-8,C29-2,CS3	2 (20)	3 (30)	2 (20)	1 #35 or I variant (10), 1 #27 or I variant (10),
							1 #48 or III variant (10)
(60)	Sheep and Cat- tle	7	GRA6,B1	7(100)			
Total	-	512	TUB2, W35, TgM-A, B18, B17, B1, Sequencing with 529, SAG2, SAG3, ,SAG1, alt-SAG2, BTUB, GRA6, Apico, PK1, C22-8, C29-2, CS3	162 (31.64)	137 (26.7 6)	167 (32.62)	46 (8.98)

#### Treatment studies for T. gondii

Very few investigations (n= 100, 6.49 %) have been carried out on the treatment of T. gondii infection in Iran. The efficacy of synthetic drugs (62%), herbal medicines and other compounds (38%) against T. gondii were evaluated in vitro and in vivo. The synthetic drugs, such as propranolol, Atovaquone, Clindamycin, Co-trimoxazole, etc., have shown good anti-toxoplasmic effects (61). Recently, attention has been drawn to the therapeutic potential of herbal products in Iran due to their lower side effects and higher availability. According to a systematic review, the extracts of Garlic, Achilleamillefolium, Hypericumperforatum, Sambucusnigra were tested for the treatment of toxoplasmosis (62).

Most of the studies were performed on the acute toxoplasmosis using RH strain. Since currently available treatments of toxoplasmosis are insufficiently effective with severe side effects, new therapeutic options for the treatment are urgently needed that effective penetration and concentration in the placenta, trans placental passage, parasitical properties versus the different parasitic stages, penetration into cysts, and distribution in the main sites.

#### Immunization studies against T. gondii

In total, 183 studies (11.88%) on immunization and vaccination against *T. gondii* have been conducted in Iran. Many researchers conducted the investigations regarding the vaccine types against toxoplasmosis. Various strategies have been used to evaluate vaccines against toxoplasmosis including live attenuated vaccines, DNA vaccines, recombinant protein vaccines and epitope-based vaccines. The main outcomes of vaccines evaluation have been summarized in Table 4.

Variable	Parasite strain	Antigens/ adjuvant	Effect	Refer- ences
DNA vaccines	Not determined	SAG1, SAG3, SAG5/ CpG-ODN	Increased survival time 10 and fewer parasite load (15,485 per mg of spleen).	(63)
	#	ROP38	ROP38 was proved a non-allergenic and antigenic protein. It had Sec signal pep- tide (Sec/SPI) with 0.8762 likelihood. It is suitable candidate vaccine against toxo- plasmosis.	(64)
	RH	ROP13 in pcDNA3 Plasmid	ROP13 was success- fully sub cloned into the pcDNA3 expres- sion vector.	(65)
	$2 \times 10^3 \mathrm{RH}$	ROP8 +IL 12	Enhanced the level of anti- <i>T. gondii</i> anti- bodies.	(66)
	#	SAG1-Related Se- quence 3 (SRS3)	SRS3 stimulate the immune system against toxoplasmo- sis.	(67)
	#	ROP16	This protein was immunogenic and non-allergenic.	(68)
	#	GRA12	GRA12 had several excellent B-cells and T-cells epitope, indi- cating that it would become an excellent vaccine against <i>T.</i> <i>gondii.</i>	(69)
	#	SAG1	Analysis showed several immunodom- inant B-cell, cytotox- ic and Helper T- lymphocyte epitopes with excellent im- munogenicity prop- erties, rendering it as a prominent vaccine candidate.	(70)
Recombinant protein vaccines	RH	rSAG1-loaded PLGA	Elicited higher IFN- γ, specific anti- T.gondii IgG and longer survival time in mice.	(71)

Table 4 : Summary of Toxoplasma gondii vaccine candidates from 2020 designed by researchers in Iran

Multi-epitope vac- cine	RH	Th17/GRA14 and ROP13	Enhanced of IL-17, and IL-22 and signif- icant induction in ROS and considera- ble decrease in para- site load was ob- served in mice.	(72)
	$2 \times 10^3 \mathrm{RH}$	ROP8	Induced strong hu- moral and cellular responses and pro- longed the survival time in BALB/c.	(73)
	$1 \times 10^4 \mathrm{RH}$	MIC3, ROP8, and SAG1	Effective protection against the parasite achieved an increase in survival time in the immunized mice, especially in the MRS-CaNPs group.	(74)
Inactivated parasite, crude or purified antigens	#	Calcium-Dependent Protein Kinase 7 (CDPK7)	The protein has im- munogenic and nonallergenic nature.	(75)
	#	calcium-dependent protein kinase-3 (CDPK3)	It is higher affinity for MHC-binding and CTL epitopes	(76)
	1.5 × 10 <sup>6</sup> RH	Soluble total antigen	STAg increased the release of IL1β. IL18 significantly upregu- lated after 24 h.	(77)

#: Bioinformatics investigation

In general, killed *T. gondii* parasite vaccines cannot be effective enough in any of the infection animal models. In contrast, liveattenuated vaccines are capable of enhancing MHCclass1-restricted CD8<sup>+</sup> T-cell immune responses, which it is considered as the most important pathway for the elimination of intracellular parasites. However, there are major concerns that attenuated vaccines have been the risk of reverting to a pathogenic strain.

Currently, appropriate antigens from different parts of *T. gondii* are used as DNA vaccine or recombinant protein vaccine, which stimulates immune responses against *T. gondii* infection. Antigens used as suitable candidates for immunization studies include SAG1, SAG2, SAG3, GRA2, GRA5, GRA6, GRA7, GRA8, GRA14, ROM4, ROP1, ROP2, BAG1 and MIC3. Protein prime/DNA boost have been shown as an efficient strategy to induce both cellular and humoral immune. Recently the use of multi-epitope vaccines has become popular in research institutes in Iran as new potential vaccines against toxoplasmosis. Multi-antigenic vaccinations could overcome for limitation of single antigen and enhance the protective immunity against *T. gondii* infection.

#### Discussion

More than 70 years have passed since the first case of clinical detection of toxoplasmosis in Iran. The relatively high prevalence of *T*.

gondii infection in animals, especially farm animals as the main sources of meat consumed by the Iranian people, as well as environmental pollution (water and soil), increases the risk of human infections. The prevalence of toxoplasmosis in the general population of Iran is reported to be 39% which may be due to geographical location and habitat, eating habits, and lifestyle. Most diagnostic studies in Iran are based on the diagnosis of IgM and IgG anti-Toxoplasma antibodies which today due to false positive and false negative test results especially in pregnant women, their results are controversial. Efforts to establish a fully standardized diagnostic method with high sensitivity and specificity are needed to dissolve this problem using specific antigens of T. gondii and designing diagnostic kits. Information on genetic diversity of the parasite in human and animal toxoplasmosis in Iran using the Mn-PCR-RFLP method is highly limited; however, ToxoDB genotyping of T. gondii can play an important role in studies on population structure, epidemiology, vaccine, and biological characteristics of T. gondii. Over the years, there have been a few studies on the effects of drugs and synthetic compounds, as well as plant extracts on T. gondii. So far, there has been no effective drug to treat and prevent toxoplasmosis with low side effects; although none of the drugs have been effective on the cyst form of T. gondii. Numerous studies have been conducted in the field of designing different models of vaccine candidates. These include live-attenuated vaccines, DNA vaccines, recombinant protein vaccines, and multi-epitope vaccines that are useful for limiting single antigens and enhancing the protective immunity against T. gondii infection. Up to now, several studies using different diagnostic tests have shown the prevalence of T. gondii in cats in Iran.

However, it is necessary to conduct a comprehensive study at the national level and with a single test on cats. To date, no national studies have been conducted on the GIS of *Toxo*- plasma in humans, animals and environmental samples. Therefore, there is a need for a comprehensive study of toxoplasmosis in different populations across the country. This is important for providing information on areas where control efforts should be targeted. Toxoplasmosis is clearly neglected as a human disease, according to reports of the infection. Moreover, a comprehensive investigation has not been carried out in the field of awareness and health education (KAP study) among girls in schools and women on the verge of marriage. Hence, these studies are highly recommended to help prevent and control the infection. Moreover, the incidence of congenital toxoplasmosis in Iranian infants and pregnant women is unknown. Therefore, it is necessary to conduct a comprehensive study in the country to determine the parasitic burden of infection in pregnant women and infants. Future research in various fields of T. gondii, especially in the case of treatment (with emphasis on cystic form) and vaccination, is highly recommended for evaluating various drug derivatives and multi-epitope vaccine candidates.

## Limitation

In this study, we reviewed all Persian and English articles related to *T. gondii* with Iranian affiliation, and therefore, according to the mentioned entry and exit criteria, no study has been lost. Due to the high volume of articles (1540) and also having a limit on the number of words and references for this journal, we tried to include the most important and up-todate articles in the study (in table or references).

## Conclusion

More than half of the conducted studies in Iran were in the field of epidemiology. In order to obtain sufficient information for proper control, prevention and treatment of toxoplasmosis in Iran, it is necessary to perform further studies in the field of genotyping, diagnosis, vaccination and treatment.

### Acknowledgements

We gratefully acknowledge the supporting of the Deputy of Research, Mazandaran University of Medical Sciences, Sari, Iran.

# **Conflict of interest**

None.

### References

- Saebi E. Protozoan Parasitic Diseases in Iran. Tehran: Ayege Publisher. 2011;5th ed.
- Jamalian R, Yalda A, Nassirzadeh MH. Diagnosis of toxoplasmosis with adenopathy by isolation of pathogenic strains (first report in Iran with introducing two patients). Journal of Tehran University of Medical Sciences. 1974;4(3):39.
- 3. Daryani A, Sarvi S, Aarabi M, et al. Seroprevalence of *Toxoplasma gondii* in the Iranian general population: a systematic review and meta-analysis. Acta Trop. 2014;137:185-94.
- 4. Foroutan-Rad M, Khademvatan S, Majidiani H, et al. Seroprevalence of *Toxoplasma gondii* in the Iranian pregnant women: a systematic review and meta-analysis. Acta Trop. 2016;158:160-9.
- 5. Sarvi S, Chegeni TN, Sharif M, et al. Congenital toxoplasmosis among Iranian neonates: a systematic review and meta-analysis. Epidemiol Health. 2019;41: e2019021.
- Mansouri A, Mojarad MRA, Badfar G, et al. Epidemiology of *Toxoplasma gondii* among blood donors in Iran: A systematic review and metaanalysis. Transfus Apher Sci. 2017;56(3):404-9.
- 7. Ahmadpour E, Daryani A, Sharif M, et al. Toxoplasmosis in immunocompromised patients in Iran: a systematic review and metaanalysis. J Infect Dev Ctries. 2014;8(12):1503-10.
- 8. Mizani A, Ahmadpour E, Daryani A, et al. *Toxoplasma gondii* infection among sheep and goats in Iran: a systematic review and meta-analysis. Parasitol Res. 2015;114:1-16.
- 9. Sarvi S, Daryani A, Rahimi MT, et al. Cattle toxoplasmosis in Iran: a systematic review and

meta-analysis. Asian Pac J Trop Med. 2015;8(2):120-6.

- 10. Rahimi MT, Daryani A, Sarvi S, et al. Cats and *Toxoplasma gondii:* a systematic review and metaanalysis in Iran. Onderstepoort J Vet Res. 2015;82(1):1-10.
- 11. Shokri A, Sharif M, Teshnizi SH, et al. Birds and poultries toxoplasmosis in Iran: a systematic review and meta-analysis. Asian Pac J Trop Med. 2017;10(7):635-42.
- Fallah E, Hajizadeh M, Farajnia S, et al. SAG2 locus genotyping of *Toxoplasma gondii* in meat products of East Azerbaijan Province, North West of Iran During 2010-2011 Afr J Biotechnol. 2011;10(62):13631-5.
- Mahami-Oskouei M, Moradi M, Fallah E, et al. Molecular detection and genotyping of *Taxo-plasma gondii* in chicken, beef, and lamb meat consumed in northwestern Iran. Iran J Parasitol. 2017;12(1):38-45.
- Gharekhani J, Yakhchali M, Afshari A, et al. Herd-level contamination of *Neospora caninum*, *Toxoplasma gondii* and *Brucella* in milk of Iranian dairy farms. Foodborne Pathogens and Disease. 2013;10(2):120-5.
- Razmi G, Barati M. Prevalence of Neospora caninum and Toxoplasma gondii antibodies in bulk milk of dairy cattle, Mashhad, Iran. Arch Razi Inst. 2017;72(4):265-9.
- Khademi SZ, Ghaffarifar F, Dalimi A, et al. Molecular detection and genotype identification of *Toxoplasma gondii* in domestic and industrial eggs. J Food Saf. 2018;38(6):e12534.
- Tavalla M, Oormazdi H, Akhlaghi L, et al. Genotyping of *Toxoplasma gondii* isolates from soil samples in Tehran, Iran. Iran J Parasitol. 2013;8(2):227-233.
- Saki J, Khademvatan S, Yousefi E, et al. Detection and genotyping of *Toxoplasma gondii* isolated from soil in Ahvaz, southwest of Iran. J Parasit Dis. 2017;41(1):202-5.
- Solyemane H, Eslamirad Z, Bayat M. Detection of *Toxoplasma Gondii* Oocysts in Soil of Urban Parks, Based on Molecular and Staining Techniques (Arak, Iran). Iran J Public Health. 2014;43(2):299.
- Mahmoudi MR, Kazemi B, Haghighi A, et al. Detection of Acanthamoeba and *Toxoplasma* in River Water Samples by Molecular Methods in Iran. Iran J Parasitol. 2015;10(2):250-7.

- Haghparast-Kenari B, Sarvi S, Sharif M, et al. Isolation and genotypic characterization of *Toxoplasma gondii* based on GRA6 gene from environmental soil samples in Mazandaran Province, North of Iran. Iran J Parasitol. 2020;15(2):158-167.
- 22. Moghazy E, Kandil F, Shaapan R. *Toxoplasma* gondii: comparison of some serological tests for antibody detection in sera of naturally infected pigs. World J Zool. 2011;6:204-8.
- 23. Mamaghani AJ, Tabaei SJS, Ranjbar MM, et al. Designing diagnostic kit for *Toxoplasma gondii* based on GRA7, SAG1, and ROP1 Antigens: An in silico strategy. Int J Pept Res Ther. 2020:1-15.
- 24. Golkar M, Rafati S, Abdel-Latif MS, et al. The dense granule protein GRA2, a new marker for the serodiagnosis of acute *Toxoplasma* infection: comparison of sera collected in both France and Iran from pregnant women. Diagn Microbiol Infect Dis. 2007;58(4):419-26.
- 25. Golkar M, Azadmanesh K, Khalili G, et al. Serodiagnosis of recently acquired *Toxoplasma gondii* infection in pregnant women using enzyme-linked immunosorbent assays with a recombinant dense granule GRA6 protein. Diagn Microbiol Infect Dis. 2008;61(1):31-9.
- Hosseininejad M, Azizi H, Hosseini F, et al. Development of an indirect ELISA test using a purified tachyzoite surface antigen SAG1 for sero-diagnosis of canine *Toxoplasma gondii* infection. Vet Parasitol. 2009;164(2-4):315-9.
- Abdollahi SH, Arababadi MK, Hassanshahi G. Evaluation of excreted/secreted antigens derived from peritoneal of *Toxoplasma* infected small mice to detect IgG against Toxoplasma. Pak J Biol Sci. 2009;12(6):530.
- Jalallou N, Bandepour M, Khazan H, et al. Recombinant SAG1 antigen to detect *Toxoplasma gondii* specific immunoglobulin G in human sera by ELISA test. Iran J Parasitol. 2010;5(2):1-9.
- 29. Elyasi H, Babaie J, Fricker-Hidalgo H, et al. Use of dense granule antigen GRA6 in an immunoglobulin G avidity test to exclude acute *Toxoplasma gondii* infection during pregnancy. Clin Vaccine Immunol. 2010;17(9):1349-55.
- Hosseininejad M. Evaluation of an indirect ELISA using a tachyzoite surface antigen SAG1 for diagnosis of *Toxoplasma gondii* infection in cats. Exp Parasitol. 2012;132(4):556-60.

- Selseleh M, Keshavarz H, Mohebali M, et al. Production and evaluation of *Toxoplasma gondii* recombinant GRA7 for serodiagnosis of human infections. Korean J Parasitol. 2012;50(3):233-8.
- 32. Selseleh MM, Keshavarz H, Mohebali M, et al. Production and evaluation of *Toxoplasma gondii* recombinant surface antigen 1 (SAG1) for serodiagnosis of acute and chronic Toxoplasma infection in human sera. Iran J Parasitol. 2012;7(3):1-9.
- Jalallou N, Bandehpour M, Khazan H, et al. Evaluation of recombinant sag1 protein for detection of *Toxoplasma gondii* specific immunoglobulin M by ELISA test. Iran J Parasitol. 2012;7(4):17-21.
- Khanaliha K, Motazedian MH, Kazemi B, et al. Evaluation of recombinant SAG1, SAG2, and SAG3 antigens for serodiagnosis of toxoplasmosis. Korean J Parasitol. 2014;52(2):137-142.
- Allahyari M, Mohabati R, Babaie J, et al. Production of in-vitro refolded and highly antigenic SAG1 for development of a sensitive and specific *Toxoplasma* IgG ELISA. J Immunol Methods. 2015;416:157-66.
- keshavarzi F. Evaluation of the efficacy of recombinant ROP1 antigen in the diagnosis of *Toxoplasma gondii* infection. J Shaheed Sadoughi Univ Med Sci. 2015;23:2083-95.
- 37. Arab-Mazar Z, Fallahi S, Koochaki A, et al. Immunodiagnosis and molecular validation of *Toxoplasma gondii*-recombinant dense granular (GRA) 7 protein for the detection of toxoplasmosis in patients with cancer. Microbiol Res. 2016;183:53-9.
- 38. Lashkari A, Golkar M, Sohrabi N, et al. Development of ELISA avidity test using recombinant GRA 8 protein to distinguish acute from chronic toxoplasmosis infection. Tehran Payame Noor University, East center Master of Sciences thesis. 2016
- Marashiyan SM, Moradian F, Saadatnia G, et al. Evaluation of recombinant SRS3 antigen for diagnosis of toxoplasmosis by enzyme-linked immunosorbent assay. Arch Clin Infect Dis.2017;12(1): e35612.
- 40. Mirzadeh A, Saadatnia G, Golkar M, et al. Production of refolded *Toxoplasma gondii* recombinant SAG1-related sequence 3 (SRS3) and its use for serodiagnosis of human

toxoplasmosis. Protein Expr Purif. 2017;133:66-74.

- 41. Morovati H, Seyyed Tabaei S, Gholamzad M, et al. Development of a lateral flow immunoassay using recombinant dense granular antigen (GRA) 7 to detect anti-*Toxoplasma gondii* IgG antibodies. Arch Razi Inst. 2019;74(1):39-49.
- 42. Teimouri A, Modarressi MH, Shojaee S, et al. Development, optimization, and validation of an in-house Dot-ELISA rapid test based on SAG1 and GRA7 proteins for serological detection of *Toxoplasma gondii* infections. Infection and drug resistance. Infect Drug Resist .2019;12:2657.
- 43. Sharif M, Amouei A, Sarvi S, et al. Genetic diversity of *Toxoplasma gondii* isolates from ruminants: a systematic review. Int J Food Microbiol. 2017;258:38-49.
- Hosseini SA, Sharif M, Sarvi S, et al. Genetic characterization of *Toxoplasma gondii* in Iranian HIV positive patients using multilocus nested-PCR-RFLP method. Parasitology. 2019;147(3):322-8.
- 45. Chaichan P, Mercier A, Galal L, et al. Geographical distribution of *Toxoplasma gondii* genotypes in Asia: A link with neighboring continents. Infection, Infect Genet Evol. 2017;53:227-38.
- Behzadi R, Razavi M, Hovanessian A, et al. Genotyping of *Toxoplasma gondii* strains isolated from patients and mice by PCR-RFLP assay. Iran J Biotechnol. 2003;1:82–6.
- Zia-Ali N, Fazaeli A, Khoramizadeh M, et al. Isolation and molecular characterization of *Toxoplasma gondii* strains from different hosts in Iran. Parasitol Res. 2007;101(1):111-5.
- Habibi G, Imani A, Gholami M, et al. Detection and identification of *Toxoplasma gondii* type one infection in sheep aborted fetuses in Qazvin Province of Iran. Iran J Parasitol. 2012;7(3):64-72.
- Shahbazi Gr, Hoghooghi Rn, Madani R, et al. Survey on Gra6 gene in differentiating *Toxoplasma Gondii* genotypes, using pcrrflp method, in sheep aborted fetuses in ardabil area, Iran. Comp Pathol IRAN. 2013; 10 (3); 1027 -1032.
- 50. Khademvatan S, Saki J, Yousefi E, et al. Detection and genotyping of *Toxoplasma gondii* strains isolated from birds in the southwest of Iran. Br Poult Sci. 2013;54(1):76-80.

- Asgari Q, Fekri M, Monabati A, et al. Molecular genotyping of *Toxoplasma gondii* in human spontaneous aborted fetuses in Shiraz, Southern Iran. Iran J Public Health. 2013;42(6):620-5.
- 52. Norouzi M, Tabaei SJS, Niyyati M, et al. Genotyping of *Toxoplasma gondii* strains isolated from patients with ocular toxoplasmosis in Iran. Iran J Parasitol. 2016;11(3):316-324.
- 53. Sarkari B, Yaghoobi K, Mansouri M, et al. Seroprevalence and genotyping of *Toxoplasma gondii* in wild boars (Sus scrofa) from Southwestern Iran. Jundishapur J Microbiol. 2017;10(1).
- Danehchin L, Razmi G, Naghibi A. Isolation and genotyping of *Toxoplasma gondii* strains in ovine aborted fetuses in Khorasan Razavi Province, Iran. Korean J Parasitol. 2016;54(1):15-20.
- 55. Tavalla M, Asgarian F, Kazemi F. Prevalence and genetic diversity of *Toxoplasma gondii* oocysts in cats of southwest of Iran. Infect Dis Health. 2017;22(4):203-9.
- 56. Nourmohammadi M, Hamidinejat H, Tabandeh M, et al. Genotyping of zoonotic *Toxoplasm gondii* isolated from aborted fetuses of ewes of Lorestan Province based on SAG2 • SAG3 and GRA6 molecular markers. Journal of Ardabil University of Medical Sciences. 2017;17(3):343-52.
- 57. Abdoli A, Dalimi A, Soltanghoraee H, et al. Molecular detection and genotypic characterization of *Toxoplasma gondii* in paraffinembedded fetoplacental tissues of women with recurrent spontaneous abortion. Int J Fertil Steril. 2017;10(4):327-336.
- Armand B, Solhjoo K, Kordshooli MS, et al. *Toxoplasma gondii* Type I, predominant genotype isolated from sheep in South of Iran. Vet World. 2017;10(4):386-92.
- Abdoli A, Arbabi M, Pirestani M, et al. Molecular assessment of Neospora caninum and *Toxoplasma gondii* in hooded crows (Corvus cornix) in Tehran, Iran. Comp Immunol Microbiol Infect Dis. 2018;57:69-73.
- 60. Rahdar M, Arab L, Samarbaf-zadeh AR. Genotyping of *Toxoplasma gondii* in Sheep and Cattle Meat Using PCR-RFLP Technique. Veterinary Science Research. 2021;2(2):2673.
- 61. Montazeri M, Sharif M, Sarvi S, et al. A systematic review of in vitro and in vivo

activities of anti-*Toxoplasma* drugs and compounds (2006–2016). Front Microbiol. 2017;8:25.

- 62. Sharif M, Sarvi S, Pagheh AS, et al. The efficacy of herbal medicines against *Toxoplasma gondii* during the last 3 decades: a systematic review. Canadian journal of physiology and pharmacology. 2016;94(12):1237-48.
- 63. Khodadadi M, Ghaffarifar F, Dalimi A, et al. Immunogenicity of in-silico designed multiepitope DNA vaccine encoding SAG1, SAG3 and SAG5 of *Toxoplasma gondii* adjuvanted with CpG-ODN against acute toxoplasmosis in BALB/c mice. Acta Trop. 2021;216:105836.
- 64. Nosrati MC, Ghasemi E, Shams M, et al. *Toxoplasma gondii* ROP38 protein: bioinformatics analysis for vaccine design improvement against toxoplasmosis. Microb Pathog. 2020;149:104488.
- Alizadeh P, Daryani A, Ahmadpour E, et al. Cloning and Expression of *Toxoplasma gondii* Rhoptry13 Gene in pcDNA3 Plasmid. Journal of Mazandaran University of Medical Sciences. 2019;28(169):26-35.
- Foroutan M, Barati M, Ghaffarifar F. Enhancing immune responses by a novel multiepitope ROP8 DNA vaccine plus interleukin-12 plasmid as a genetic adjuvant against acute *Toxoplasma gondii* infection in BALB/c mice. Microb Pathog. 2020;147:104435.
- Mirzadeh A, Saadatnia G, Golkar M, et al. In Silico Prediction of T and B Cell Epitopes of SAG1-Related Sequence 3 (SRS3) Gene for Developing *Toxoplasma gondii* Vaccine. Arch Clin Infect Dis. 2020;15(6):1-9.
- 68. Ghaffari AD, Dalimi A, Ghaffarifar F, et al. Structural predication and antigenic analysis of ROP16 protein utilizing immunoinformatics methods in order to identification of a vaccine against *Taxoplasma gondii*: an in silico approach. Microb Pathog. 2020;142:104079.
- Ghaffari AD, Dalimi A, Ghaffarifar F, et al. Antigenic properties of dense granule antigen 12 protein using bioinformatics tools in order to improve vaccine design against *Toxoplasma* gondii. Clin Exp Vaccine Res. 2020;9(2):81-96.

- Majidiani H, Dalimi A, Ghaffarifar F, et al. Computational probing of *Toxoplasma gondii* major surface antigen 1 (SAG1) for enhanced vaccine design against toxoplasmosis. Microb Pathog. 2020;147:104386.
- 71. Allahyari M, Mohabati R, Vatanara A, et al. Invitro and in-vivo comparison of rSAG1-loaded PLGA prepared by encapsulation and adsorption methods as an efficient vaccine against *Toxoplasma gondii*". J Drug Deliv Sci Technol. 2020;55:101327.
- 72. Fatollahzadeh M, Eskandarian A, Darani HY, et al. Evaluation of Th17 immune responses of recombinant DNA vaccine encoding GRA14 and ROP13 genes against *Toxoplasma gondii* in BALB/c mice. Infect Genet Evol. 2021:105150.
- 73. Foroutan M, Ghaffarifar F, Sharifi Z, et al. Vaccination with a novel multi-epitope ROP8 DNA vaccine against acute *Toxoplasma gondii* infection induces strong B and T cell responses in mice. Comp Immunol Microbiol Infect Dis. 2020;69:101413.
- 74. Dodangeh S, Fasihi-Ramandi M, Daryani A, et al. Protective efficacy by a novel multi-epitope vaccine, including MIC3, ROP8, and SAG1, against acute *Toxoplasma gondii* infection in BALB/c mice. Microb Pathog. 2021;153:104764.
- 75. Taghipour A, Tavakoli S, Sabaghan M, et al. Immunoinformatic Analysis of Calcium-Dependent Protein Kinase 7 (CDPK7) Showed Potential Targets for *Toxoplasma gondii* Vaccine. J Parasitol Res. 2021;2021.
- Majidiani H, Soltani S, Ghaffari AD, et al. Indepth computational analysis of calciumdependent protein kinase 3 of *Toxoplasma gondii* provides promising targets for vaccination. Clin Exp Vaccine Res. 2020;9(2):146-158.
- 77. Pazoki H, Rahimi HM, Mirjalali H, et al. Soluble total antigen derived from *Toxoplasma* gondii tachyzoites increased the expression levels of NLRP1, NLRP3, NLRC4, AIM2, and the release of mature form of IL1β, but downregulated the expression of IL1β and IL18 genes in THP-1cell line. Microb Pathog. 2021;158:105072.