



Tehran University of Medical
Sciences Publication
<http://tums.ac.ir>

Iran J Parasitol

Open access Journal at
<http://ijpa.tums.ac.ir>



Iranian Society of Parasitology
<http://isp.tums.ac.ir>

Original Article

Genotyping of *Echinococcus granulosus* Isolates from Human in Khorasan Province, North-Eastern Iran

Fariba BERENJI¹, Seyed Aliakbar SHAMSIAN¹, Marziyeh NOURI DALOEE², Seyed Hossein FATTAHI MASOOM², *Elham MOGHADDAS¹

1. Department of Parasitology and Mycology, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran
2. Department of Thoracic Surgery, Cardiothoracic Surgery and Transplant Research Center, Mashhad University of Medical Sciences, Mashhad, Iran

Received 19 Feb 2018 Accepted 27 Jun 2018	Abstract Background: Human hydatidosis is endemic in northeastern Iran. The present study aimed to investigate molecular diversity of <i>Echinococcus granulosus</i> isolates collected from human surgically. Methods: Sixty human hydatid cysts (58 lung cysts and 2 liver cysts) were collected through surgery from Ghaem and Emam Reza hospitals in Mashhad University of Medical Sciences during 2015-2016. Cysts were characterized using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) analysis of the internal transcribed spacer 1 (ITS1) gene and sequencing fragments of the genes coding for mitochondrial cytochrome c oxidase subunit 1 (<i>cox1</i>) and NADH dehydrogenase subunit I (<i>nad1</i>). Results: Overall, 55 out of 60 <i>Echinococcus granulosus</i> cysts (91.6%) were determined as the G1 strain, 4 cases (6.6%) were determined as the G6 strain and 1 sample was not identified. Conclusion: Although sheep strain (G1) is dominated in human patients in Great Khorasan, the prevention of camel-dog cycle should pay attention in this region.
Keywords: Hydatid; Human; Strain; Iran; <i>Echinococcus granulosus</i>	
*Correspondence Email: Moghaddase@mums.ac.ir	

Introduction

Echinococcus granulosus settle in small intestine of carnivores as definitive host. Almost of herbivores e.g.,

human, sheep, cattle, buffalo, goat, camel, horse, pig and many of wild ungulates are intermediate hosts (1). In intermediate host,

hydatid cyst was created after ingestion egg of *E. granulosus* via contaminated food, vegetable or water. Direct contact with dogs is the most common route of infection transmission. Humans are accidental host of parasitic hydatid disease and it may occur in each body organs (2).

Genetic variability and morphological variations exist within *E. granulosus* that caused G1 to G10 strains (3). Most of the genotype of human cystic echinococcosis in the worldwide is G1 strain (88.44%), after that the genotype G6 (7.34%). G5, G8 and G10 genotypes were recorded rarely from human and no case from G4 genotype have been identified in human up to now (4). Detection of these strains is important for the epidemiology, control, prevention programs, vaccine designs and drug production.

G1, G3 and G6 strain were reported from Iranian patients (5). Nucleic acid sequencing is the gold standard for determining the genotypes of *E. granulosus* (6). Hydatidosis is endemic in Iran (1% of all surgical admissions). Incidence of human hydatid disease in Khorasan area is as high as 4.45 in 100,000 (7, 8). The average number of operated cysts per year was 134.2 (8). Cyst strains were studied in camels, sheep, goats, and cattle in northeast of Iran without any survey on human hydatid cysts (9, 10). Even alveolar echinococcosis in Monkey (*Ateles geoffroyi*) and small mammals were reported in this area (11, 12).

The aim of this study was to strains identification of *E. granulosus* in human in Great Khorasan Province, north-eastern Iran.

Materials and Methods

Sample collection

Sixty pieces of germinal layer of cyst including 58 lung cysts and 2 liver cysts were collected from Ghaem and Emam Reza hospitals in Mashhad University of Medical Sciences, Iran in 2015-2016. All patients lived in Mashhad or small cities that located in Great Khorasan

including North Khorasan, South Khorasan and Khorasan Razavi.

DNA extraction

Germinal layer of every cyst was rinsed several times with sterilized distilled water prior to DNA extraction. Total genomic DNA (gDNA) was extracted from each cyst as manufacturer's instructions (MBST, Tehran, Iran). About 5% Mm of cyst wall lysed in 180 μ l lysis buffers, the proteins were degraded with 20 μ l proteinase K for 10 min at 55 °C. After adding 360 μ l of binding buffer and incubation for 10 min at 70 °C, 270 μ l of ethanol (100%) were added to the solution. After vortexing, the complete volume was transferred into the MBST column. The MBST column was first centrifuged and then washed twice with 500 μ l of washing buffer. Finally, DNA was eluted from the carrier with 50 ml elution buffer. The concentration of DNA was determined by nanodrop and the samples were stored at -20 °C.

PCR

ITS1 Gene

The forward and reverse primers employed in this study for ITS1 were: EgF (forward), 5'-CAGAGCACTTTTGTATGCA-3'); EgR (reverse), (5'-ATGGTTGTATCGCTG CGA-3') (9). The following program was used: 5 min incubation at 95 °C to denature double-stranded DNA, 35 cycles of 45 sec at 94 °C (denaturing step), 45 sec at 50 °C (annealing step) and 45 sec at 72 °C (extension step). Finally, PCR was completed with an additional extension step for 10 min.

PCR-RFLP of ITS1

For PCR-RFLP, the amplified products (20 μ l) were digested with five units of the restriction endonuclease Bsh1236I (5U, Fermentas) in a final volume of 25 μ l for eight hours in 37 °C (9). This enzyme identifies sequences of CG/CG. Restriction fragments were visualised by gel electrophoreses through 4% ethidium bromide agarose gel.

Cox1 and Nad1 Gene Sequencing

Randomly 10 samples of G1 and 1 sample of G6 pattern of RFLP- PCR amplified by PCR method. Two primers, JB3 (forward), 5'-TTT TTT GGG CAT CCT GAG GTT TAT -3' and JB4 (reverse), 5'-TAA AGA AAG AAC ATA ATG AAA ATG -3' were used to amplify a 450 bp fragment of *cox1* gene under the following conditions: initial denaturation 94°C for 5 min, followed by 35 cycles of denaturation at 94 °C for 45 sec, annealing at 50 °C for 45 sec and elongation for 45 sec at 72 °C. Final extension was performed at 72 °C for 7 min (13). Also two primers, MS1 (forward): 5'-CGTAGGTATGTT GGTGTTTGGT-3') and MS2 (reverse): 5'-CCATAATCAAATGGCGTACGAT-3' were used to amplify a 400 bp with 5 min initial denaturation at 94 °C, and 30 cycles of 30 sec denaturation at 94 °C, 45 sec annealing at 50 °C and 30 sec elongation at 72 °C. Final extension was performed at 72 °C for 5 min (14). *Nad1* and *cox1* genes after purification were sent for sequencing (Macrogen Company, South Korea). The sequences were compared with those previously published in the GenBank database using the BLAST system (<http://www.ncbi.nlm.nih.gov/blast/Blast.cgi>).

Ethics

The study adhered to the tenets of the Declaration of Helsinki and was approved by the Ethics Committee at Mashhad University of Medical Sciences (Ethical code: IR.MUMS.fm.REC.1394.443).

Results

We detected fifty-five G1 (sheep strain) and four G6 (camel strain) according to RFLP-PCR of ITS1 following were confirmed by sequencing of *cox1* and *nad1* genes. All *E. canadensis* were isolated from lung. One unknown strain did not get satisfied results in RFLP- PCR as same in sequencing result. PCR product of ITS1 gene showed approximately 462 bp in length (Fig. 1). As we expected the restriction enzyme Bsh1236I made

three and four bands for G1 and G6 genotype respectively (Fig. 2).

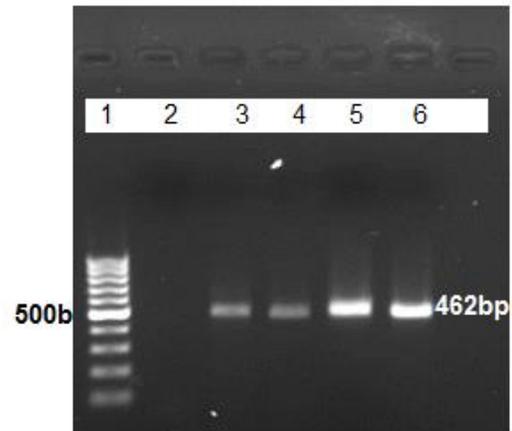


Fig. 1: Agarose gel electrophoresis of ITS1-PCR (462 bp) products of *E. granulosus* isolates from human: Lanes 1: ladder 100bp, Lanes 2: negative control, Lane 3, 4, 5: ITS1, Lane 6: positive control

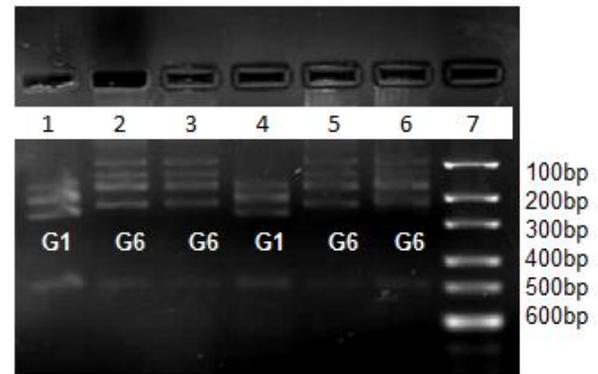


Fig. 2: Digestion pattern of PCR products from rDNA-ITS1 fragment with restriction enzyme Bsh1236I: lane 7: marker with 100bp molecular weight, Lane 1, 4: G1 genotype, Lane 2, 3, 5, 6: G6 genotype

The partial nucleotide of *cox1* (450bp) and *nad1* (400bp) genes from 10 isolates were amplified by PCR method (Fig. 3). Sequencing of *cox1* and *nad1* genes confirmed the RFLP-PCR results. *Cox1* and *nad1* Genes of G6 strain were 100% homology with accession number of Kp751426 and Kp751432 respectively in GenBank. *Cox1* Gene of G1 strain was 99% homology with accession number of

MG 322623 in GenBank and showed in comparison with G1 in GenBank a transition of T to A at position 481, A to G at position 650. Moreover, *nad1* Gene of G1 strain was 99% homology with accession number of MG 322623 in GenBank in comparison with G1 in GenBank, a transition of G to A at position 292, T to A at position 418.

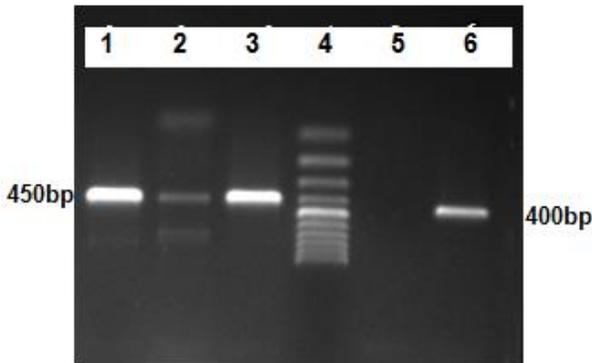


Fig. 3: Agarose gel electrophoresis of *cox1*-PCR (450 bp) and *nad1* (400 bp) products of *E. granulosus* isolates from human: Lanes 1, 2, 3 *cox1* gene, Lane 4: ladder 100bp, lane 5: negative control and lane 6: *nad1* gene

Phylogenetic analysis

Phylogenetic trees were generated by using *cox1* and *nad1* sequencing (Figs. 4 and 5). Alignment was performed using ClustalW and the aligned sequences manually refined in BioEedi software (version 7.2.5) maximum likelihood (ML) trees were inferred by MEGA 7 software. Nodal support was assessed by bootstrapping with 1000 replicates.

Discussion

North-east of Iran is hyperendemic for hydatidosis and also alveolar echinococcosis in human and animals (8, 15). Another study has published one thousand and twenty of pulmonary hydatid cyst in different hospitals of Mashhad (capital city of Khorasan Razavi) from 1981 to 2008 (16). During 9 years 1342 human hydatidosis were operated only in

three hospitals in this area (8). Hydatid cysts from 70 cattle, 50 sheep and 24 goats were shown G1 genotype in Khorasan Province (10).

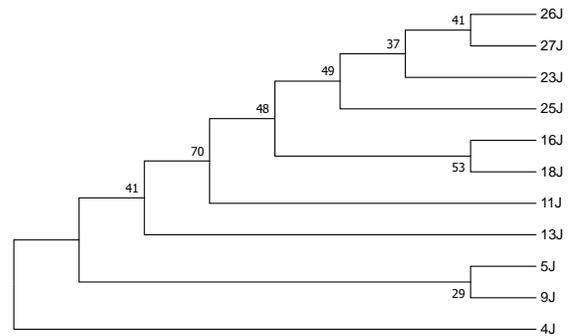


Fig. 4: The evolutionary history was inferred by using the Maximum Likelihood method based on the General Time Reversible model. The tree with the highest log likelihood (-629.50) is shown *cox1* sequences of *Echinococcus canadensis* G6 genotype (4J) and G1 genotype (9J, 11J, 13J, 5J, 16J, 18J, 23J, 25J, 26J, 27J)

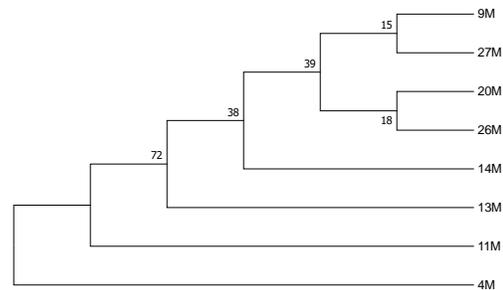


Fig. 5: The evolutionary history was inferred by using the Maximum Likelihood method based on the General Time Reversible model. The tree with the highest log likelihood (-814.94) is shown *nad1* sequences of *Echinococcus canadensis* G6 genotype (4M) and G1 genotype (9M, 11M, 13M, 14M, 20M, 26M, 27M). There were a total of 319 positions in the final dataset. Evolutionary analyses were conducted in MEGA7

In present study dominant strain was G1, followed by G6 which is in accordance with almost other reports in different provinces in Iran (Table 1).

In the present study 4/60 (6.6%) of all genotype were G6 genotype (camel strain) in human patients. Great Khorasan is neighbour of Afghanistan and annually 4500 of dromedary camels were slaughtered in industrial abattoirs. Interestingly, G6 strain was reported from human in Afghanistan (17). Liver and lung camel hydatidosis were recorded 11.1% and 13.2%, respectively in northeastern Iran (18). Moreover, prevalence of canine *Echinococcus* in this region was 22% and it was reported to 10,000 worms in one dog intestine (19). This condition can make potential danger of transmission of camel strain to human in

northeastern Iran. G6 was responsible for human hydatidosis 9.1% and 40.8% from central and southeast of Iran, respectively (20).

Almost the studies conducted on genotyping of *E. granulosus* in Iran including *cox1* and *nad1* sequencing with or without PCR-RFLP of rDNA-ITS1. By using these techniques G1, G3 and G6 obtained in human and domestic animals in whole the country (Table 1).

Because PCR-RFLP is not capable to differentiate G1-G3 from each other, we did gene sequencing *nad1* and *cox1* genes. We did not find any G3 strain in human cases.

Table 1: Published information concerning *Echinococcus granulosus* isolated from humans in different regions of Iran

Number of human cases	Area	Strains	Method	Ref
4	Tehran	Sensu stricto (G1-G3)	Sequencing of <i>cox1</i> and <i>nad1</i> genes PCR-RFLP	(21)
23	Isfahan	Sensu stricto (G1-G3)	Sequencing of <i>cox1</i> and <i>nad1</i> genes	(14)
55	Azerbaijan Province	G1	PCR-RFLP (rdna-ITS1)	(22)
4	Ilam Province	Sensu stricto (G1-G3)	PCR-RFLP (rdna-ITS1)	(23)
29	Tehran	G1, G3 and G6	Sequencing of <i>cox1</i> and <i>nad1</i> genes	(24)
30	Golestan province	G1	PCR-RFLP (rdna-ITS1)	(25)
5	Khuzestan province	G1	PCR-RFLP (rdna-ITS1)	(26)
11	Ardabil Province	G1, G3	Sequencing of <i>cox1</i> and <i>nad1</i> genes	(27)
31	Isfahan province	G6, G1	PCR-RFLP (rdna-ITS1)	(28)
12	Different locations of Iran	Sensu stricto (G1-G3)	PCR-RFLP ITS1	(29)
30	Isfahan Province	Sensu stricto (G1-G3)	PCR-RFLP ITS1	(30)
1	Kerman Province	G6	Sequencing of <i>cox1</i> and <i>nad1</i> genes	(31)
17	Fars Province	8 (G1) and 6 (G6)	Sequencing <i>nad1</i> gene	(32)

Conclusion

G1 (sheep strain) is the dominant genotype that involved human hydatidosis in north-eastern Iran. In additional G6 (camel strain) observed in this district or the first time in human hydatidosis.

Acknowledgements

We thank Mrs. Hosseini for collecting the cyst specimens in surgery centers. This work was supported by Mashhad University of Medical sciences financially supported this

study (Project grants: 940088). The sponsor had no role in the design or conduct of this research.

Conflict of interest

The authors declare that there is no conflict of interests.

References

1. Thompson RC. The taxonomy, phylogeny and transmission of *Echinococcus*. *Exp Parasitol*. 2008; 119(4):439-46.
2. Moro P, Schantz PM. Echinococcosis: a review. *Int J Infect Dis*. 2009; 13(2):125-33.
3. Thompson RC, McManus DP. Towards a taxonomic revision of the genus *Echinococcus*. *Trends Parasitol*. 2002; 18(10):452-7.
4. Alvarez Rojas CA, Romig T, Lightowlers MW. *Echinococcus granulosus* sensu lato genotypes infecting humans—review of current knowledge. *Int J Parasitol*. 2014; 44(1):9-18.
5. Harandi MF, Hobbs RP, Adams PJ et al. Molecular and morphological characterization of *Echinococcus granulosus* of human and animal origin in Iran. *Parasitology*. 2002; 125(Pt 4):367-73.
6. Lymbery AJ, Thompson RC. The molecular epidemiology of parasite infections: tools and applications. *Mol Biochem Parasitol*. 2012; 181(2):102-16.
7. Amoueian S, Tayebi MN, Mohammadian RN. A retrospective study of 1759 cases of hydatid cyst in Mashad University hospitals. *HAKIM*. 2005; 7(4): 7-13.
8. Andalib Aliabadi Z, Berenji F, Fata A et al. Human Hydatidosis/Echinococcosis in North Eastern Iran from 2003–2012. *Iran J Parasitol*. 2015; 10(4):658-62.
9. Moghaddas E, Borji H, Naghibi A et al. Molecular genotyping of *Echinococcus granulosus* from dromedaries (*Camelus dromedarius*) in eastern Iran. *J Helminthol*. 2015; 89(1):100-4.
10. Fadakar B, Tabatabaei N, Borji H et al. Genotyping of *Echinococcus granulosus* from goats and sheep indicating G7 genotype in goats in the Northeast of Iran. *Vet Parasitol*. 2015; 214(1-2):204-7.
11. Borji H, Emami M, Maleki M et al. Alveolar echinococcosis infection in a monkey (*Ateles geoffroyi*) in Mashhad, Iran. *Iran J Public Health*. 2012; 41(2):111-6.
12. Beirumvand M, Akhlaghi L, Fattahi Massom SH et al. Molecular identification of *Echinococcus multilocularis* infection in small mammals from Northeast, Iran. *PLoS Negl Trop Dis*. 2013; 7(7):e2313.
13. Bowles J, Blair D, McManus DP. Genetic variants within the genus *Echinococcus* identified by mitochondrial DNA sequencing. *Mol Biochem Parasitol*. 1992; 54(2):165-73.
14. Sharbatkhori M, Mirhendi H, Jex AR et al. Genetic categorization of *Echinococcus granulosus* from humans and herbivorous hosts in Iran using an integrated mutation scanning-phylogenetic approach. *Electrophoresis*. 2009; 30(15):2648-55.
15. Geramizadeh B, Baghernezhad M. Hepatic Alveolar Hydatid Cyst: A Brief Review of Published Cases from Iran in the Last 20 Years. *Hepat Mon*. 2016: e38920.
16. Bagheri R, Haghi SZ, Amini M et al. Pulmonary hydatid cyst: analysis of 1024 cases. *Gen Thorac Cardiovasc Surg*. 2011; 59(2):105-9.
17. Schneider R, Gollackner B, Schindl M et al. *Echinococcus canadensis* G7 (pig strain): an underestimated cause of cystic echinococcosis in Austria. *Am J Trop Med Hyg*. 2010; 82(5):871-4.
18. Borji H, Azizzadeh M, Afsai A. An abattoir-based study of hydatidosis in the dromedary (*Camelus dromedarius*) in Mashhad, Iran. *J Helminthol*. 2011; 85(4):478-9.
19. Razmi GR, Sardari K, Kamrani A. Prevalence of *Echinococcus granulosus* and other intestinal helminths of stray dogs in Mashhad area, Iran. *Arch Razi Inst*. 2006; 61(3): 143-189.
20. Rostami S, Shariat Torbaghan S, Dabiri S et al. Genetic characterization of *Echinococcus granulosus* from a large number of formalin-fixed, paraffin-embedded tissue samples of human isolates in Iran. *Am J Trop Med Hyg*. 2015; 92(3):588-94.
21. Zhang L, Eslami A, Hosseini SH et al. Indication of the presence of two distinct strains of *Echinococcus granulosus* in Iran by mitochondrial DNA markers. *Am J Trop Med Hyg*. 1998; 59(1):171-4.

22. Vahedi A, Mahdavi M, Ghazanchaei A et al. Genotypic characteristics of hydatid cysts isolated from humans in East Azerbaijan Province (2011-2013). J Anal Res Clin Med. 2014; 2(3): 152-157.
23. Dousti M, Abdi J, Bakhtiyari S et al. Genotyping of hydatid cyst isolated from human and domestic animals in Ilam Province, Western Iran using PCR-RFLP. Iran J Parasitol. 2013; 8(1):47-52.
24. Nikmanesh B, Mirhendi H, Ghalavand Z et al. Genotyping of *Echinococcus granulosus* isolates from human clinical samples based on sequencing of mitochondrial genes in Iran, Tehran. Iran J Parasitol. 2014; 9(1):20-7.
25. Gholami Sh, Sosari M, Fakhar M et al. Molecular characterization of *Echinococcus granulosus* from Hydatid Cysts isolated from human and animals in Golestan Province, North of Iran. Iran J Parasitol. 2012; 7(4):8-16.
26. Khademvatan S, Yousefi E, Rafiei A et al. Molecular characterization of livestock and human isolates of *Echinococcus granulosus* from south-west Iran. J Helminthol. 2013; 87(2):240-4.
27. Pezeshki A, Akhlaghi L, Sharbatkhori M et al. Genotyping of *Echinococcus granulosus* from domestic animals and humans from Ardabil Province, northwest Iran. J Helminthol. 2013; 87(4):387-91.
28. Shahnazi M, Hejazi H, Salehi M et al. Molecular characterization of human and animal *Echinococcus granulosus* isolates in Isfahan, Iran. Acta trop. 2011; 117(1):47-50.
29. Ahmadi N, Dalimi A. Characterization of *Echinococcus granulosus* isolates from human, sheep and camel in Iran. Infect Genet Evol. 2006; 6(2): 85-90.
30. Kia EB, Rahimi H, Sharbatkhori M et al. Genotype identification of human cystic echinococcosis in Isfahan, central Iran. Parasitol Res. 2010; 107(3):757-60.
31. Hajjalilo E, Harandi MF, Sharbatkhori M et al. Genetic characterization of *Echinococcus granulosus* in camels, cattle and sheep from the south-east of Iran indicates the presence of the G3 genotype. J Helminthol. 2012; 86(3):263-70.
32. Sadjjadi SM, Mikaeili F, Karamian M et al. Evidence that the *Echinococcus granulosus* G6 genotype has an affinity for the brain in humans. Int J Parasitol. 2013; 43(11):875-7.