Review Article

The Role of Neuroinflammation in Alzheimer's Disease (AD)

Mahdi Rafiyan¹, Hanieh Mojtahedi^{1,2*}

¹ Network of Immunity in Infection, Malignancy and Autoimmunity (NIIMA), Universal Scientific Education and Research Network (USERN), Tehran. Iran

² Molecular Immunology Research Center, Tehran University of Medical Sciences, Tehran, Iran

Received: 02 October 2024; Accepted: 28 December 2024

Abstract

Neuroinflammation is a critical process in Alzheimer's disease (AD) development in which different types of cells and cytokines are involved. Proinflammatory cytokine production and the disturbance of anti-inflammatory pathways play critical roles in AD. Neuroinflammation is affected by various factors such as metabolism (metabolic diseases such as obesity), genetics, and immune cells, especially resident immune cells in the brain. Moreover, the main pro-inflammatory cytokines and inflammatory pathways have different effects on neuroinflammation, neuronal biogenesis, and neuronal apoptosis in AD. Exploration of the relationship between neuroinflammation, risk factors of neuroinflammation, and pro-inflammatory cytokines in AD helps us to understand AD pathogenesis and select therapeutic targets more efficiently.

Keywords: Alzheimer's Disease (AD); Pro-Inflammatory Cytokine; Neuroinflammation; Metabolism

*Corresponding Author: Hanieh Mojtahedi Molecular Immunology Research Center, Tehran University of Medical Sciences, Tehran, Iran

E-mail: haniyeh.mojtahdi@gmail.com

How to cite this article

Rafiyan M, Mojtahedi H. The Role of Neuroinflammation in Alzheimer's Disease (AD). Immunol Genet J, 2025; 8(2): 150-188. https://doi.org/10.18502/igj.v8i2.17998

Introduction

the most common cause of dementia is Alzheimer's creased memory functions, and gradually leads disease (AD), which is responsible for 60-70% of to a total inability to do essential daily life tasks dementia cases. It is estimated that the number of and, eventually, death (3). Based on recent epidepatients with AD will exceed 7 million in 2030 (1, miological studies, the prevalence of AD would

2). AD is characterized by a progressive decline One of the world's most prevalent diseases and in cognitive functions, usually initiated by de-

Copyright © 2025 Tehran University of Medical Sciences. Published by Tehran University of Medical Sciences.

This work is licensed under a Creative Commons Attribution-NonCommercial 4.0 International license (https://creativecommons.org/ licenses/by-nc/4.0/). Non-commercial uses of the work are permitted, provided the original work is properly cited.

double every 20 years, at least until 2040, due to the rapid aging of the nations (3). The etiology of this disease is complicated, and several factors, such as environmental and genetic factors, have been assumed to contribute to the multifactorial etiology of this disease. For instance, infections, diet, and metals (such as aluminum) are effective environmental factors (4). Regarding genetics, different genes have been attributed to the initiation and progression of AD, such as three well-studied genes: the amyloid precursor protein (APP), presenilin 1 (PSEN1), and presenilin 2 (PSEN2) genes (5-7). Studies show that one of the critical mechanisms in developing AD is inflammation, which is defined as the immune system's response to pathogens or organ damage (8). Inflammation occurs in almost all parts of the body, including the central nervous system (CNS), which leads to the activation of astrocytes and microglia, the accumulation of various chemokines and cytokines, and neurodegenerative processes (9). Microglia are the resident macrophages in the CNS, while astrocytes are the most frequent subtype of glial cells, which both are responsible for neuroinflammation by secreting different cytokines and chemokines (10). The most common hallmark of AD is the accumulation of beta-amyloid peptide (A β). Indeed, it could be the first trigger in the pathogenesis of this disease. Furthermore, tubulin-associated unit (tau) protein aggregation, the formation of different cellular and intercellular neuritic plaques, neurofibrillary tangles(NFTs), and neuropil threads are other hallmarks of AD (11).

Taken together, all these aggregations stimulate the activation of astrocytes and microglia, leading to the secretion of pro-inflammatory cytokines and chemokines from microglia and pro- and anti-inflammatory cytokines from astrocytes, which potentially cause synaptic damage, neuronal death, and finally AD development (12). Cytokines and chemokines, as important subcategories of immune mediators, are involved in the induction of inflammation; therefore, they play a prominent role in the development of AD through synaptic dysfunction, neuronal death, and inhibition of neurogenesis (13). However, the importance of microglia in brain functions such as neural plasticity, long-term potentiation (LTP), and brain homeostasis by producing pro-inflam-

matory cytokines at lower concentrations cannot be denied (14, 15). Inflammation is an important factor in various metabolic disorders, such as diabetes, and also in neurodegenerative diseases, such as AD (16). On the other hand, Metabolic disorders such as diabetes mellitus have a significant role in AD development and progression by induction of pathological changes in the body, including vascular changes, inflammation, and blood glucose increase (17, 18). Gut microbiota can also induce neuroinflammation and disturb metabolic homeostasis by either disrupting the blood-brain barrier (BBB) and delivering toxins in the brain or via various pro-inflammatory mediators (19). Considering the high prevalence and the potential heavy burden of AD, understanding the pathophysiology of this disease is of high value. In this review, the effects of inflammation and key factors involved in inducing neuroinflammation and neurodegeneration in the pathogenesis of AD and the possible target therapies are discussed.

Common Hallmarks of AD

Αβ

Beta-amyloid peptide $(A\beta)$ is one of the most important hallmarks of AD, which plays a critical role in the pathogenesis and neuronal and synaptic dysfunction during the progression of AD (20). The *amyloid precursor protein* (*APP*) gene is located on chromosome 21, and its product, APP, is a precursor of the A β protein (21). Recent studies showed that APP has several essential roles for the brain's normal function, such as metal binding and protease inhibition, while it is also a component of the extracellular matrix (21, 22). APP can be cleaved by specific enzymes (secretases) and result in different substrates. Normally, it is cleaved by α-secretase, resulting in the formation of soluble APP (sAPPa) and transmembrane C-terminal fragment (α -CTF). Then, γ -secretase cleaves α-CTF to generate a 23-25 amino acid peptide called P3 and APP intracellular domain (AICD). This pathway is non-amyloidogenic and cannot lead to AD development. However, improper cleavage of APP resulted from β and γ secretase activity. In the first step of an amyloidogenic pathway, β -secretase cleaves APP to produce β -CTF and soluble APP- β . Then, γ -secretase

cleaves β -CTF and forms A β , composed of 38 to 43 amino acids with different solubility, stability, biological, and toxic properties (21, 23, 24). Accumulation of A β (A β 1–42) leads to mast cell activation, which increases blood-brain barrier (BBB) permeability, and the release of inflammatory mediators such as cytokines, chemokines, and other neuroactive mediators, which cause glial cells and neuron activation (25-27). However, Aβ-activated astrocytes and microglia increase uptake of $A\beta$ via microglia and protect neurons from A β toxicity by the secretion of transforming growth factor- β (TGF- β), which is a neurotrophic and anti-inflammatory cytokine (28)(Both pathways of APP proteolytic processing are presented in Figure 1). With regard to the pivotal role of A β in AD development, numerous therapies have been developed based on the inhibition of A β formation, aggregation, or degradation (29).

Tau Protein

The other major hallmark of AD is tau protein aggregations that form neuritic plaques (NP), neurofibrillary tangles, and neuropil threads. Tau is a soluble, microtubule-associated protein (MAPT), playing an essential role in supporting the neuronal cell's microarchitecture complex (30). Moreover, the tau protein is involved in synaptic modulation and neuronal growth (31). Several mechanisms lead to tau protein disturbances, such as the phosphorylation of tau by kinases and the formation of p-tau, which is insoluble in water. The aberrant hyperphosphorylation of tau leads to tau's dissociation from microtubules and the promotion of tau aggregation. Meanwhile, the aggregation of p-tau leads to neurofibrillary tangles and thread formation (32). In addition, inflammation is an important mechanism as well, since the increase in interleukin 1β (IL- 1β), a pro-inflammatory cytokine, leads to the hyperphosphorylation of tau by kinases (33). Diabetes mellitus and genetic factors are also involved in tau protein alterations, with complex underlying mechanisms (34-36). The utilization of tau protein as a therapeutic target remains controversial, and several clinical trials are in process (36-38).

Role of Inflammation in AD

Since the 1980s, when Griffin *et al.* reported the increase in Interleukin-1 (IL-1) in AD patients,

several studies have focused on the central role of inflammation in the development of AD (39, 40). Inflammation is a complex process that consists of several different pathways. Cytokines are important signaling molecules needed for proper homeostasis, which have inflammatory and/ or anti-inflammatory functions depending on the target receptor, cell, and the phase of an immune response (41, 42). Neuroinflammation refers to the inflammation of the neurons, developed by various factors interfering with CNS homeostasis, consisting of external factors including infection, trauma, ischemia, and aging (43), while internal factors are composed of cytokines, chemokines, reactive oxygen species (ROS), microglia, epithelial cells (44). Epithelial cells in CNS can produce various substances such as IL-1b and TNF-a, as pro-inflammatory cytokines (45). Microglia, as another part of neuroinflammation, induce apoptosis in neurons and phagocytosis through pro-inflammatory cytokine secretion (46-49). Moreover, the role of T-cells and B-cells as parts of adaptive immunity in neuroinflammation is inevitable. T-cells can target neurons and induce apoptosis in them. Also, T-cells can interact with activated microglia, resulting in the inflammation of the CNS and demyelination (50).

T helper (Th)17 cells, as one of the subcategories of T-cells with inflammatory features, produce a wide spectrum of pro-inflammatory cytokines such as IL-6, Interleukin-17A (IL-17A), Interleukin-17F (IL-17F), interferon- γ (IFN- γ), and TNF-a, which exacerbate neuroinflammation (51, 52). It should be noted that inflammation is a critical response to trauma, infection, and the normal function of the CNS, leading to the induction of neurogenesis in different parts of the brain, such as the hippocampus, via activation of T lymphocytes (53). Additionally, studies showed that interleukin-4 (IL-4)-producing T cells are needed for cognitive performance (54). Neuroinflammation is required for the regulation of neurons and neurogenesis after an insult. Furthermore, the facilitation of axonal regeneration through M2-macrophages and Th1, but not Th2 or Th17 cells, provision of neurotrophic factors, and its critical role in remyelination of the neurons make this phenomenon an important part of the normal recovery of the CNS after an injury (55-59). However, inflammation plays a double-edged sword in the prolonged form known as chronic neuroinflammation, which exacerbates neuronal damage and neurodegeneration. Neurodegeneration is a critical and central process for cognitive dysfunctions and the development of neurodegenerative diseases like AD (60, 61).

Cellular and Molecular Pathways Involved in Alzheimer's Neuroinflammation

Microglia

Microglia are the resident macrophages of the CNS, which contribute to the homeostasis maintenance of the CNS. Microglia are classified into M1 (pro-inflammatory state) and M2 (repairing and protective state). However, it should be noted that several different subtypes of microglia have been identified in the brain, including KSPG-microglia, satellite microglia, Hox8b-microglia, and Disease-associated microglia (DAM)(62). Neurodegeneration-associated molecular patterns (NAMPs) are danger molecules present on myelin debris, apoptotic bodies of dying neural cells, and the accumulation of abnormal proteins such as A β , and are the triggers of the transition of resident microglia to DAM via TREM2, a main receptor of DAM (63). In the early stages of AD, activation of DAM could reduce the velocity of disease progression, but inappropriate activation of DAM leads to neuroinflammation and deterioration of AD (64). In a normal brain, microglia are in the M2 state. Studies showed that in the context of AD, microglia cells showed phenotypic alteration from M2 to M1 state (65, 66). M1 microglia have a prominent role in inflammation by secretion of various cytokines, especially pro-inflammatory cytokines such as IL-1B, IL-6, IL-18, IL-12, IL23, TNF-a, and neurotoxic substances, which are responsible for blocking neuronal differentiation, attenuating microglial phagocytosis, extracellular matrix damage through the activation of nuclear factor-kB and accumulation of AB and as well as calling other inflammatory cells to the inflammation site through cytokines (67-71). When the amyloid accumulation becomes overt, their phagocytic function is disturbed. Additionally, studies showed that $A\beta$ can activate the molecular pathways of pro-inflammatory cytokine secretion in microglia, such as NF-kB and NLR family pyrin domain containing 3 (NLRP3) in-

flammasome secretion, mediated by cell surface receptors such as CD36, CD47, and a-6/b-1 integrin, leading to neuroinflammation and neurodegeneration (72-76). NF- κ B is a protein family that controls DNA transcription and expression. It is important in inflammation as it increases the pro-inflammatory cytokine expression pathway. Additionally, NF- κ B stimulates the β -secretase (BACE1) cleavage of APP and A β production by enhancing BACE1 expression (77, 78). NLRP3 is a part of the innate immune system and is found in macrophages and inflammasomes. These proteins can trigger immune responses (79, 80). Moreover, this inflammasome is an intracellular protein complex that regulates the maturation of IL-1 β and IL-18 and also increases the cleavage and activity of caspase-1, which are significantly increased in AD brains and associated with the onset and progression of the disease (81, 82).

Astrocyte

Astrocytes are a group of glial cells that reside in the CNS. They have essential roles in the CNS, including repairing the CNS, protecting neurons from harmful agents and neurotoxic substances, modulating synapses (83). These cells are also important in neurodegenerative diseases like Parkinson's disease (PD) and AD (84, 85). Secretion of cytokines such as IL-1a and TNF-a from microglia, resulting in astrocyte activation and the formation of reactive astrocytes (86). There are two forms of reactive astrocytes: A1 and A2. Neuroinflammation gives rise to the A1 form. A proposed mechanism for this phenomenon is through A β and NF- κ B. A β can activate the NF- κ B pathway in astrocytes and induce A1. A1 is a neurotoxic astrocyte and upregulates the expression of the complement cascade gene, which leads to the release of the complement protein C3. This protein binds to the C3aR in microglia and neurons. Activation of the complement-3a receptor (C3aR) in microglia increases phagocytosis, and in neurons, it disrupts dendritic morphology and network function, both of which contribute to AD pathogenesis (28, 87, 88). A2 is a protective form of reactive astrocytes and is induced by ischemia. This form upregulates the expression of neurotrophic genes and promotes survival, growth, and repair of synapses (28). Astrocytes also contribute to glucose hypometabolism through the glutamatergic excitotoxicity mechanism. Increasing glutamate production and decreasing the glutamate receptors of astrocytes, including GLT-1 and GLAST, causes an increase in the amounts of glutamate in the CNS. GLT-1 and GLAST are responsible for glutamate reuptake from CSF, and some studies have suggested that A β can suppress these receptors. Glucose hypometabolism increases stress oxidative production, which can cause neuroinflammation either directly or by producing pro-inflammatory cytokines (89-91).

Cytokines and Signaling Pathways

IL-1

IL-1 (also called "endogenous pyrogen") was the first cytokine that proved to affect the CNS(92). Both isoforms of IL-1 (IL-1a and IL- 1β) are pro-inflammatory cytokines, have similar effects, and are produced in a variety of cells, like microglia and lymphocytes, as precursor proteins called pro-IL-1a and pro-IL-1β. Pro-IL-1a is an active form of IL-1a that can be cleaved to form IL-1a, a smaller active molecule, by a specific enzyme called CAPLAIN. IL-1a acts intracellularly, but it can also be released after neurodegeneration. Pro-IL-1 β is a precursor of IL-1 β , but unlike pro-IL-1 α , it is an inactive form of IL-1 β and should be broken down by the CASPASE-1 enzyme to form an active form, IL-1 β (93, 94). IL-1 α and IL-1 β apply their intracellular signaling via membrane-bound type I IL-1 receptor (IL-1R1) (95). This complex is reinforced by IL-1-receptor accessory protein (IL-1RAcP), as this receptor is needed for the normal function of IL-1R1. Korher *et al.* showed that the response to IL-1 via IL-1RI internalization or IL-2 production in the absence of IL-1RAcP could not occur (96). Another receptor of IL-1 is IL-1R2, considered a decoy receptor, and no intracellular signaling pathway was identified for this receptor; however, recent studies suggested that it is mainly expressed on microglia and can diminish the cytokine-induced microglial activation (95, 97, 98). IL-1 receptors can also be shed from the neurons (99). An antagonist ligand of IL-1R is IL-1RA. Its secreted isoform (also called sIL-1RA) is produced by the IL-1-producing cells (100, 101). IL-1RA and IL-1R have polymorphic genes and are located on chromosome 2 (102, 103). These polymorphisms

could explain the pathogenesis of some early-onset forms of AD (102, 104, 105). IL-1 is needed for normal brain functions. Mason et al. showed that the deficiency of IL-1 β in mice leads to a delay in CNS repair and myelination. IL-1 is also required for normal sleep behavior and non-rapid eye movement (106). Another role of IL-1 is its stimulating effect on magnocellular neurons in the paraventricular nucleus and supraoptic nucleus of the Hypothalamus needed for vasopressin and oxytocin secretion (107). It also stimulates the corticotropin-releasing factor (CRF), which is secreted by neurons in the hypothalamus. In fact, this feature of IL-1 can affect Adrenocorticotropic hormone (ACTH) and cortisol secretion (108, 109).

IL-1 is involved in AD through several mechanisms: 1) It upregulates APP production. D. Goldgaber et al. showed that APP upregulation could occur through protein kinase C (PKC). The upstream binding site for APP promoter production is activator protein-1 (AP-1), which can be utilized by PKC to increase APP transcription(110). 2) The sustained release of IL-1 stimulates tau hyperphosphorylation in the neurons. Y.Li et al. showed the IL-1 effects on tau hyperphosphorylation, at least partly, via the p38p38-Mitogen-activated protein kinase(MAPK) pathway, leading to neuronal structural changes, synaptic loss, and exacerbation of AD (33). 3) The overexpression and constant release of IL-1 and its engagement with IL-1R1 causes neuroinflammation mediated by nuclear factor kappa B (NF- κ b). This pathway leads to an increase in the transcription of pro-inflammatory cytokines, including IL-6 and IL-1. 4) IL-1 can directly attach to the microglia cells, leading to the secretion of several neurotoxic substances, including pro-inflammatory cytokines (like TNF- α), chemokines (such as CC-chemokine ligand2 (CCL2)), eicosanoids (like PGE2), and reactive oxygen species (ROS)(111, 112). Shang *et al.* showed that IL-1 β drives the cellular senescence of rat astrocytes when induced by oxidative stress and oligomerized A β peptide (113). 5) IL-1 changes the pattern of gene expression of astrocytes, resulting in astrocyte proliferation (known as astrogliosis), IL-6 secretion, an increase in the release of metalloproteinases(MMPs)(114). 6) IL-1 causes expression of E-selectin, intercellular adhesion mole-

cule 1 (ICAM-1), vascular cell adhesion protein 1 (VCAM-1), and CXC-chemokine family, such as chemokine ligand 1 (CXCL1) which increase leukocyte adhesion and invasion to the brain parenchyma (114-116). 7) IL-1 has positive effects on neuron function by inhibiting these cells through γ -aminobutyric acid (GABA), inhibiting glutamate release, and calcium entry. However, if this inhibition occurs in the inhibitory pathway, it could reinforce excitatory neurons and cause neuronal death (117). Additionally, IL-1 can cause neuronal death by increasing calcium entry through N-methyl-D-aspartate (NMDA) receptors. Calcium overload in the neurons causes mitochondrial dysfunction, which triggers the apoptotic pathways. Calcium accumulation in the mitochondria causes the opening of the permeability transition pore (PTP). Opening of PTP allows pro-apoptotic factors such as cytochrome C (CytC) and apoptosis-inducing factor (AIF) to be released in the cytoplasm and activate caspases (118). Besides, high calcium influx to the endoplasmic reticulum (ER) causes the C/EBP Homologous Protein (CHOP) overexpression, which induces endoplasmic reticulum stress-mediated apoptosis (119-123)(Figure 2a).

IL-6

Another critical cytokine in AD development is IL-6. Its family has several cytokines, including IL-6, IL-11, LIF (leukemia inhibitory factor), OSM (oncostatin M), CNTF (ciliary neurotrophic factor), CT-1 (corticotrophin-1), and CLC (cardiotrophin-like cytokine)(124-126). IL-6 is a glycosylated protein composed of 4 a helixes, and its molecular weight is 21-28 kDa (127, 128). Its receptors are IL-6R and gp130. Gp130 is expressed in almost every cell in the body, and it is a crucial part of IL-6 signaling, but IL-6R is expressed in specific cells such as leukocytes, hepatocytes, T cells, etc. This selective expression helps IL-6 to act selectively in the body (129-131). IL-6 induces the proliferation and differentiation of B and T cells, increases the liver synthesis of acute-phase protein, regulates APP, increases serum amyloid A (SAA), a major acute-phase protein, induces megakaryocyte maturation and platelet release, increases collagen synthesis and collagen via an effect on fibroblast (132-137). These effects are applied through gp130 and its intracellular proteins:

Janus kinase 1 (JAK1) and STAT3, two principal intracellular signaling pathways involved in IL-6 signaling (138). The activation of JAK via IL-6 activates the PI3/Akt pathway, which leads to NFkb activation. NF-kb binds to the APP promoter and increases A β production (139). Additionally, Aβ can stimulate the release of IL-6 and exacerbate neuroinflammation (140), which activates microglia and astrocytes to produce pro-inflammatory cytokines like TNF-a (141). IL-6 can activate STAT proteins, especially signal transducers and activators of transcription 3 (STAT3), which is important for astrogliosis and astrocyte reactivity. These mechanisms are important in neuroinflammation and neurodegeneration in AD (142, 143). On the other hand, IL-6 is required for normal brain function. Gadient et al. showed that IL-6 and its receptor (IL-6R) increase during postnatal development, and they act as neurotrophic factors (144). IL-6 is involved in cognitive function, as D. Braida et al. showed that in IL-6-deficient mice, the cognitive function becomes disrupted (145). Although S. Toulmond et al. showed the preventive effect on the neurotoxicity of IL-6 after NMDA injection to the striatum (146). IL-6 also has a role in AD development, and its levels increase in the blood of AD patients. However, some studies have reported a decrease or normal level of IL-6 (147). M. Huberman et al. showed the correlation between IL-6 and AD severity. They observed a significant increase in IL-6 in both mild and moderately severe AD patients compared to the control group, but it did not significantly differ between mild and moderate patients (148).

As mentioned above, IL-6 is a strong stimulator for acute-phase protein release from the liver, like SAA, and it can be responsible for the high levels of acute-phase protein and hyperinflammation state in AD patients (149). SAA can interact with amyloid-beta and accumulate in the plaques, and facilitate memory decline (150, 151). IL-6 can increase tau hyperphosphorylation via dysregulation of the cyclin-dependent kinase 5 (cdk5)/p35 pathway and inactivation of phosphatases like protein phosphatase 1 (PP1)(152, 153). An increase in IL-6 induces Th-17 differentiation from naïve T cells. Th-17 cells exacerbate AD via two mechanisms: first, these cells activate astrocytes by IL-17 secretion and cause neurodegeneration. Second, it can cause neurodemyelination and neurodegeneration by secreting inflammatory cytokines, especially IL-23 (154-156). Despite the clear role of IL-6 in AD development, some studies reported its beneficial effect on the early phase of AD by facilitating plaque clearance (157) (**Figure 2b**).

TNF-a

TNF- α is a TNF superfamily member and a powerful cytokine with cytotoxic properties that causes tumor necrosis. Its gene has been located on chromosome 6, and studies have shown that some polymorphisms of this gene can increase the AD risk (158, 159). All of the TNF members have a TNF homology domain (THD) and a trimer structure. TNF-a has two forms: soluble (sT-NF- α) and membrane-bound (tmTNF- α) with 17 kDa and 26 kDa molecular weight, respectively (160, 161). The cleavage of tmTNF- α leads to sT-NF-a formation. Both of them are active biologically and have different roles. sTNF- α has a high affinity for binding to TNFR1, a receptor that is important in apoptosis. Thus, inhibition of this form of TNF- α in the brain can inhibit neuronal apoptosis and reverse AD progression (162). tmT-NF-α has a high affinity for TNFR2; this receptor is important for regulating genes involved in cell survival, myelination, and immunity against pathogens (163). TNF-a receptors (TNFR) are from the TNF receptor superfamily (TNFRSF) with a cysteine-rich domain (CRD), and the THD binds to this domain. Three types of TNFR have been discovered in recent years: 1) the receptors with the death domain (known as TNFR1), involved in TNF intracellular signaling and induction of apoptosis in the cell by using Fas-associated protein with death domain (FADD) (164). TNFR1 has mediated the major impact of TNF- α due to its expression at low levels on all nucleated cells of the body (165). 2) The receptors without the death domain (known as TNFR2) are expressed primarily on cells of hematopoietic origin, but neurons can also express them (166). 3- The decoy receptor binds to TNF-a with high affinity and specificity but cannot induce intracellular signaling (167).

TNF- α has numerous biological effects, including an increase in resistance to microbial infection (168), and cancer (169). Moreover, Shoham *et al.* showed the activity of TNF- α in normal sleep, as the reduced level of TNF-a correlates with a reduction in continuous sleep; the same effect has been seen in the knockout of the TNFR1 gene in animal models by an increase in $A\beta$ production. TNF- α and IFN- γ can increase the expression of β -secretase, which can lead to A β production and decrease its reuptake (170). Furthermore, TNF-a and IFN-y synergistically can decrease soluble APP, which is a protective form of this protein compared to the insoluble form of APP, causing A β production (171). L. Osborn *et al.* showed the potency of TNF-a in NF-kB activation. This scenario can also occur in microglia and increase the release of TNF- α and other cytokines, and exacerbate the inflammation (172). TNF- α can also decrease AB clearance via an effect on microglia and cause synaptic dysfunction (173). N.Hovelmeyer *et al.* showed the involvement of TNF-a in the induction of apoptosis in oligodendrocytes. Numerous studies have shown the vulnerability of oligodendrocytes and the reduction of myelin in AD. This role of TNF- α can explain this vulnerability, even as a part of the mechanisms involved in the myelin breakdown in AD (174). The upregulation of VCAM-1 on endothelium and astrocytes causes the crossing of lymphocytes and other immune cells through the BBB, and further inflammation (175). TNF- α can increase the expression of inducible nitric oxide (iNOS). The result of iNOS activity is NO, which is a neurotoxic substance and increases neuronal loss (176). TNF-α can increase the production of S100B as a zinc binder. Zinc is an important ion in normal synaptic function, and TNF-α can decrease it and induce synaptic dysfunction (177). TNF-a can decrease neurogenesis via NF-KB activation and caspase 3 and 9, two potent apoptotic factors, and related pathways (178, 179). One of the important aspects of AD is mitochondrial dysfunction. Excessive expression of TNF- α can disturb the mitochondrial function of the neurons directly. This disturbance can affect neuronal plasticity and synapses and can exacerbate AD(180)(Figure 2c).

NF-ĸB

Nuclear factor- κ B (NF- κ B) is a regulator of various genes involved in the production of cy-tokines, chemokines, NO, and COX-2, which



Non-Amyloidogenic pathway

Figure 1. Summary of APP metabolism in two different ways: amyloidogenic and non-amyloidogenic pathway: 1-in the amyloidogenic pathway, β -secretase dissociates APP from its A β part from the long side of the APP. The product of this action is soluble APP β and β -CTF, which consists of A β and AICD parts. Finally, γ -secretase cut the other end of A β connected to the AICD and separate them. Accumulation of A β causes Amyloid plaque and AD. 2- in the non-amyloidogenic pathway, first, α secretase dissociates APP to soluble APP α and α -CTF. This dissociation does not occur in the junction of sAPP and A β , but it occurs among A β . Then γ -secretase separates the small part of A β (also called p3), a part of α -CTF, from AICD. This pathway cannot lead to the formation or accumulation of amyloid plaques.



Figure 2. Summary of inflammatory cytokines in Alzheimer's Disease



Figure 3. The interaction of significant etiologies the initiation of neuroinflammation through pro-inflammatory cytokine secretion.

mediate neuroinflammation and the activation of microglia and cause phagocytosis (181). It is a member of a family of inducible transcription factors and has five members: NF- κ B1 (p50), NF- κ B2 (p52), RelA (p65), RelB, and c-Rel (182). The NF- κ B proteins are bound to I κ B family proteins, which inhibit the activity of these proteins (183). The most important I κ B protein is I κ B α . Additionally, p105 and p100, the precursor proteins of NF- κ B1 and NF- κ B2, have a C-terminal portion with a similar structure to I κ B and probably have NF- κ B inhibitory functions (80, 184, 185).

Two major signaling pathways are responsible for the activation of NF- κ B: Canonical and Non-canonical pathways (186, 187). The canonical pathway starts from the activation of the multi-subunit I κ B kinase (IKK) complex. It has two catalytic subunits, including IKK α and IKK β . Besides, a regulatory subunit called NF- κ B essential modulator (NEMO) or IKK γ also gets involved in the IKK complex (181, 188, 189). Cytokines, microbial components, and stress are some of the most common triggers of the canonical pathway, which act via different receptors such as various

cytokine receptors, pattern-recognition receptors (PRRs), T-cell receptor (TCR), B-cell receptors, and TNFR superfamily members (190, 191). Activation of IKK leads to the phosphorylation of IκBa, which triggers a ubiquitin-dependent IκBa degradation in the proteasomes. Finally, NF-KB is released in the cytosol, transferred to the nucleus, and regulates relevant genes (80, 192). Despite canonical pathways that respond to various types of stimuli, non-canonical pathways only respond to a specific group of stimuli, such as LTβR, BAF-FR, CD40, and RANK, which are members of the TNFR superfamily (193-196). Additionally, activation of NF-KB via this pathway depends on p100, an NF-kB2 precursor protein (197, 198). NF-κB-inducing kinase (NIK), in cooperation with IKKa, phosphorylates p100, which further induces its ubiquitination and processing (199, 200). At the final step, degradation of p100 C-terminal IkB-like structure leads to the release of NF-KB2 p52 and activation of further signaling pathways (201-203). The canonical pathway is the main pathway for immune response, and the non-canonical pathway has a complementary

role and, in the adaptive system, cooperates with the canonical pathway (185, 204). T cells, particularly CD4⁺ T-helper (Th) cells, are also involved in this cascade, which activates different proteins and genes, activating proinflammatory cytokines and releasing potentially toxic compounds that cause neurotoxicity, ultimately neuronal dysfunction, and cell death (205). Activation of naïve T cells occurred upon the stimulation of TCR by a specific antigen, which further activates the canonical pathway of NF-KB. RelA and c-Rel, two important members of NF-κB, have a central role in this process (206). Additionally, NF-KB promotes Th1 differentiation. Aronica et al. suggested that inhibition of NF-κB in T-cells leads to the Th1 response impairment (207).

A non-canonical pathway is required for proper differentiation and function (memory/effector) of T cells. Additionally, this pathway is required for Th17-mediated neuroinflammation (208-211). Some studies suggested that NF-KB activation increased *BACE1* and *APP* genes, as both NF- κB and BACE1 are upregulated in the AD patients' brains (212, 213). Furthermore, aging, one of the most important risk factors for AD, leads to the perpetual activation of NF-KB, which further activates microglia, neuroinflammation, and the development of AD (214). Receptors for advanced glycation end products (RAGE) are receptors of advanced glycation end products (AGEs)(215). However, studies showed that $A\beta$ is a ligand for this receptor (216). Interestingly, these receptors are overexpressed during neuroinflammation in microglia (216, 217). Binding of A β to RAGE activates which further induces NO and glutamate release, cytokine production, and BBB amplification (218, 219). NO combines with superoxides, which are associated with oxidative stress and BBB dysregulation (220, 221). Additionally, glutamate release leads to neuronal toxicity and degeneration (222, 223).

NLRP3 Inflammasome

Inflammasomes are large multiprotein complexes assembled by different receptors such as TLRs and NOD-like receptors (NLRs). Inflammasomes induce pyroptosis, characterized by the activation of caspase-1-mediated inflammatory response (224, 225). Several inflammasomes have been discovered, such as NLRP1, NLRP2, NLRP3,

AIM2, and NLRC4 (226). Among these inflammasomes, the most well-studied one is NLRP3. NLRP3, a 118 kDa PRR protein, is a cytosolic protein expressed by different cells such as neurons, microglia, neutrophils, and macrophages. It has a C-terminal leucine-rich repeat (LRR) domain and a central ATPase-containing NACHT domain required for oligomerization. Besides, its N-terminal pyrin (PYD) domain recruits proteins for the formation of the inflammasome complex. NLRP3 inflammasome is composed of a sensor (NLRP3 protein), an adaptor (apoptosis-associated specklike protein, ASC), and an effector (caspase-1) (224, 225, 227, 228). Activation of NLRP3 and its inflammasome formation could be triggered by a plethora of stimuli such as pathogens, uric acid crystals, silica, asbestos, extracellular ATP, and toxins (229, 230). Some studies hypothesized that NLRP3 activation is due to the common cellular events caused by this wide range of stimuli instead of directly binding to them (231, 232). Disruption of the trans-Golgi network (TGN) by multiple NLRP3 stimuli resulted in binding of NIMA-related kinase 7 (NEK7), an important NLRP3 inflammasome modulator, to NLRP3, which further disrupts the NLRP3 double-ring structure, inactive structure, and causes structural rearrangement. Structural rearrangement of NLRP3 exposes its PYD domain, which further associates with NACHT domain oligomerization. After the activation of the NACHT domain, the PYD domain recruits ASC and forms the ASC pyroptosome via PYD-PYD domain interaction (233-236). At the next step, the caspase recruitment domain (CARD) of ASC interacts with the pro-caspase-1 ASC domain, which further converts pro-caspase-1 to its active form, caspase-1. Caspase-1 not only converts pro-IL-1 β and pro-IL-18 into their active forms, IL-1 β and IL-18, respectively, but also activates the membrane pore-forming gasdermin D (GSDMD), a critical protein for pyroptosis (227, 237, 238).

NLRP3 inflammasomes are involved in the pathogenesis of autoinflammatory diseases, including diabetes, obesity, and AD (227). Similar to the NF- κ B pathway, NLRP3 has also been activated via canonical and non-canonical pathways. Two signals are required to activate the canonical pathway. Priming signal (first signal) includes TLR ligands and cytokines such as TNF- α and

IL-1 β , leading to the activation of NF- κ B and further upregulation of NLRP3 and pro-IL-1 β expression. The second signal (an activating signal) activates NLRP3 activation, followed by NLRP3 inflammasome, caspase-1-mediated secretion of IL-1 β and IL-18, and pyroptosis. There are different activating signals, such as mitochondrial dysfunction, ion flux, including K⁺ efflux, Cl⁻ efflux, Na⁺ influx, ROS, and lysosomal disruption (227, 238). Caspase 4 and caspase 5 are involved in the NLRP3 non-canonical pathway in humans. These caspases bind to LPS directly, which leads to their autoproteolysis and activation. Finally, caspase four and caspase 5 induce pyroptosis by activation of GSDMD or triggering of K⁺ efflux (239).

NLRP3 inflammasomes are expressed abundantly in microglia and astrocytes. Interestingly, the NLRP3 inflammasomes expressed in astrocytes are non-functional, and stimuli could not induce IL-1 β and IL-18 secretion in them (240). Halle et al. found that NLRP3 inflammasomes could be activated by A β , which leads to IL-1 β and IL-18 production and further inflammation (241). Some studies suggested that NLRP1 inflammasomes are expressed in neurons, and $A\beta$ could activate them, but the expression of NLRP3 remains controversial (242). Besides, chronic expression of NLRP3 inflammasomes in microglia disturbs their clearance capacity for A β and NFTs, which further exacerbate AD (243). These findings are consistent with Heneka and colleagues' study, which found that NLRP3⁻ or caspase-1-deficient APP/PS1 mice were resistant to neuroinflammation, AD, and amyloid plaque (244).

TREM2

TREM2 (the triggering receptors expressed on myeloid cells 2) is a cell surface transmembrane glycoprotein with a cytoplasmic tail (245), which is expressed in some subgroups of myeloid cells, such as granulocytes and dendritic cells (246-248). TREM2 is expressed by microglia, and it has higher expression in the hippocampus and spinal cord, which suggests its CNS region-dependent expression (249). Inflammation and its related cytokines, such as TNF α and IL1 β , decrease, and anti-inflammatory molecules increase TREM2 expression (250-252). TREM2 acts via an intracellular adaptor called DAP12 (DNAX-activation protein 12, also known as TYROBP) through the TREM2 cytoplasmic short tail. The interaction between a positively-charged lysine in TREM2 and a negatively-charged aspartic acid in DAP12 regulates further intracellular events. TREM2 ligation to DAP12 activates Src family kinases, which generate tyrosine phosphorylation of DAP12 within its immunoreceptor tyrosine-based activation motifs (ITAMS). ITAMS phosphorylation makes a docking site for SH2 domains of different molecules, which is associated with immune response via a cascade of signaling molecules. TREM2 signaling components such as PI3K, Akt, and MAPK are activated via Syk, a principal kinase recruited by ITAM (253-259). TREM2 ligands have not been identified well, but their functions have been studied. It increases phagocytosis rate, AB uptake, and myeloid cell number and survival. Furthermore, it decreases inflammation via modulation of TNFa and NO synthase-2 transcription (NOS2)(253, 260-263). However, studies suggested a dual role of this signaling pathway, with some considering an inflammatory role for TREM2 (264, 265). PD, amyotrophic lateral sclerosis (ALS), stroke, traumatic brain injury, and AD are some of the pathological conditions in which TREM2 expression is upregulated. It seems that TREM2 overexpression in AD recruits microglia to amyloid plaques (227, 266, 267). Interestingly, some studies suggested that A β could directly bind and activate TREM2 (268). On the other hand, lack of TREM2 expression is associated with a reduction of late-stage amyloid plaque accumulation(269). However, reduction of TREM2 expression is associated with Tau spreading around amyloid plaque (266, 270). Studies showed the involvement of TREM2 in different inflammatory pathways, such as NF-кВ. Cosker et al. found that TREM2 inhibits neuroinflammation by inhibition of NF-KB (271). Another similar study found the downregulation of PI3K/AKT by TREM2 for inhibition of neuroinflammation (272). Taken all together, TREM2 has a dual role in AD and could enhance some pathological features and relieve others.

cGas-STING

Detection of foreign DNAs is a crucial part of the immune system. In mammals, cyclic GMP– AMP synthase (cGAS)–stimulator of interferon genes (STING) pathway is responsible for this de-

tection and induces a powerful immune system response against these foreign DNAs (273). cGAS, part of this pathway, is an innate immune system receptor, and its functional part is STING. Activation of cGAS by DNAs leads to the conversion of ATP and GTP to a cyclic dinucleotide 2030-cyclic GMP-AMP (cGAMP). This activation occurs via the C-terminal part of this molecule. This part has a nucleotidyltransferase domain (the catalytic part) with a positively charged DNA-binding site. Binding of DNA to this part leads to the cGAS conformational changes and rearranges its catalytic part, which allows it to convert ATP and GTP (274-277). At the next step, cGMP activates STING, which is located on the ER (278, 279). Conformational changes of STING lead to the binding of this molecule to TBK1, which further phosphorylates the transcription factor interferon regulatory factor 3 (IRF3). IRF3 is transferred to the nucleus, which results in the production of Type-I IFNs and some inflammatory cytokines (273). Studies showed that STING could mediate a non-canonical pathway of autophagy, which requires limited types of molecules such as PI3P effector WIPI2 and the ATG5-12-16L1 complex (280-282). The advantage of this pathway is the restriction of viral propagation (283). Interestingly, some studies suggested that this pathway could prevent tumor growth by induction of autophagy in the cell during proliferation (284). STING upregulates p21 and other cell-cycle inhibitors along with proapoptotic proteins (285, 286). Additionally, phosphorylated IRF3 could induce apoptosis by interaction with BAK and BAX (287-290). Even under conditions in which apoptosis is restricted, activation of STING leads to the RIPK3-dependent necroptosis development, which occurs via type I interferon and TNF signaling pathways (291-293).

One of the critical signaling pathways in neuroinflammation is cGAS/STING/IFNs. Several studies showed the abundance of cGAS/STING in different neuroinflammation-related disorders. One of the main elevated types of interferons during neuroinflammation is IFN-I, a main product of the cGAS/STING pathway. It has receptors on microglia, astrocytes, and neurons (227, 294). Activation of microglia by IFN-I leads to the pro-inflammation (295, 296). Wang *et al.*,

showed that STING could regulate the activation of NLRP3 (297). Another study held by Jin and colleagues showed that polyglutamine binding protein 1 (PQBP1), an important protein for splicing, transcription, and cognitive functions of the brain, interacts with tau 3R/4R proteins, resulting in cGAS/STING activation and further immune response (298).

Genetics

The trace of genetics can be found in almost every disease, and it is also important in neuroinflammation and neurodegeneration. Several genes have been identified for early onset AD, PD, neuroprotection against inflammation, induction, and early resolution of inflammation in the CNS(299, 300). Four well-studied genes involved in AD are *PSEN1*, *PSEN2*, *APP*, and *APOE4*.

PSEN1

PSEN1 is located on chromosome 14q24.3 and encodes PS1, a multi-spanning transmembrane protein with a hydrophilic loop (301, 302). Its N-terminal and hydrophilic loop are available for interaction with other proteins. Besides, PS1 is a highly conserved 50 kDa protein found in different brain regions such as the dentate gyrus, neocortex (especially in layers II and IV), the CA1-CA3 layers, and the subiculum of the hippocampus. The intracellular location of PS1 is in intracellular membranous organelles such as the ER, nuclear envelope, and Golgi apparatus. Mutations of *PSEN1* are the first cause of familial Alzheimer's disease (FAD)(303). PS1 is a catalytic subunit of y-secretase, and as mentioned above, this enzyme plays a crucial role in the production of A β and the development of AD (304, 305). However, the exact role of PS1 in the development of AD remains debatable. Two main hypotheses have been developed to explain the role of PS1 in FAD: the amyloid and presenilin hypotheses. The former one proposed that mutations of *PSEN1* lead to the overproduction of $A\beta$ and further development of AD (306, 307). This hypothesis has been evolved and proposed that *PSEN1* mutations increase the $A\beta 42/A\beta 40$ ratio. (308). Presenilin hypothesis provides an alternative view and proposes that mutations of PSEN1 cause loss of function of its protein, which is associated with dementia and neuroinflammation

(309). This hypothesis is supported by several studies, which showed that *PSEN1* is important in the survival of neurons during aging, as well as learning and memory (310-312). Besides, *PSEN1* loss of function disturbs γ -secretase activity. As Xi *et al.* showed, despite decreased production of both A β 42 and A β 40 after γ -secretase loss of function, A β 42/A β 40 increases, which is further associated with AD development (313).

PSEN2

PSEN2 was first reported as a causative gene for AD development in 1995 (314). It is located on chromosome 1q42.13 and encodes the PSEN2 protein (315). PS1 and PSEN2 are homologous proteins with 67% similarity (316). Their hydrophobic region is highly conserved, and their difference is in their N-terminal and hydrophilic loop (315). PSEN2 has two isoforms: Isoform 1 is found in different tissues such as the placenta, liver, and kidney, and Isoform 2 is found in the brain, placenta, and skeletal muscles (315). Like PS1, this protein is also a subunit of γ -secretase, and PSEN2 mutations could change the activity of this enzyme and further AD development. PSEN2 like PS1 resides in ER and Golgi appartus (317). Studies showed that some mutations of PSEN2 increase $A\beta$ production, while others change the intracellular calcium signaling (318-320).

APP

APP or amyloid beta precursor protein is the substrate of enzymes for the production of $A\beta$ and further development of AD. It is a highly conserved protein encoded by the APP gene (321-323). APP is an integral membrane protein with a large extracellular and small intracellular region. Its extracellular region has two subdomains: E1 and E2, which are linked to each other by an acidic domain (324). APP is expressed in different tissues such as skin, adipose tissue, muscles, and CNS, but its main functions are in the CNS, which are the regulation of synapse formation, enhancing synapse adhesion, increasing neuronal viability, and axon pruning (325-327). APP mutations could increase the $A\beta 42/A\beta 40$ ratio and increase Aß generation. Additionally, mutations could impair α -secretase action on the APP and increase the hydrophobicity of $A\beta$, which is further associated with amyloid plaque formation (328-330). Several mutations have been identified, including: 1) Mutations in the N-Terminal of A β Domain: *K*670*N*/*M*671*L* is an example of this type of mutation, which leads to the lysine-to-asparagine substitution at codon 670 and a methionine-to-leucine substitution at codon 671, are within the extracellular part of APP at the β -secretase cleavage site and increase both A β 40 and A β 42 (331, 332). 2) mutations in the $A\beta$ Domain: For instance, E693G increases A β protofibril formation (333, 334). 3) Mutations in the C-Terminal of $A\beta$ Domain: These mutations disturb their respective secretases and lead to the production of longer A β (A β 42), which aggregates easily (330, 335). 4) A673V mutation (substitution of alanine to valine at codon 673) increases production of A β (336). 5) A673T (alanine-to-threonine substitution) decreases $A\beta$ formation and has a protective role against AD, probably via impairment of BACE1 cleavage of APP (337, 338).

APOE4

Apolipoprotein E (APOE) is located on the chromosome 19q13.32 and is a primary apolipoprotein lipid and cholesterol transporter in the CNS (339, 340). It also enhances the transportation of lipids via different cells by acting as a ligand of low-density lipoprotein receptor (LDLR) and lipoprotein receptor-related protein (LRP)(341). *APOE* is composed of N-terminal and C-terminal domains linked by a hinge, and its isoforms are mainly different in the N-terminal of this protein (339).

APOE has 3 isoforms: E2, E3, and E4. APOE4 is the strongest risk factor for AD development, as it influences different aspects of AD (342). APOE4 is associated with more Aß accumulation and aggregation in the brain (343-345). Additionally, some studies reported that APOE4 could change the clearance of A β (346). It also influences tau pathology via induction of neuroinflammation, increase of neuronal accumulation, and redistribution of this protein (347-349). Furthermore, induction of a more damaging related reactive astroglia signature in APOE4 harboring mice indicates its effect on astrocytes (347, 350, 351). Prasad et al. showed that APOE4 could restrict the ability of astrocytes in amyloid clearance (351). APOE4 could also promote disease-associated microglia (DAM) in the brain. These microglia are involved in the induction of neuroinflammation and tau pathology (352-354).

Non-coding RNAs

Regulation of gene transcription depends on several factors, and non-coding RNAs (ncRNAs) are one of the crucial parts of gene transcription regulation. Long non-coding RNAs (LncRNAs), microRNAs (miRNAs), and circular RNAs (Currans) are three important members of this family. LncRNAs are RNAs with more than 200 nucleotides that interfere with mRNA and/or miRNAs. miRNAs have 18-24 nucleotides. miRNAs bind to mRNA and lead to the degradation of mRNA. CircRNAs are closed LncRNAs that have similar roles to LncRNAs. Some of the ncRNAs and their targets are shown in **Table 1**.

miR-155 is a highly conserved miRNA and is important for the immune system and T-helpers. Studies showed that this miRNA could lead to neuroinflammation. miR-155 decreases the endogenous anti-inflammatory cell response, which is further associated with neuroinflammation and brain damage. It is also involved in the inflammatory response after CNS ischemia, Parkinson's disease, MS, ALS, and traumatic brain injury (355-360). Guedes et al. investigated the functional role of miR-155 in AD using the 3xTg AD animal model. They showed a strong upregulation of miR-155 levels in the brain of 3xTg AD animals. Simultaneously, the rise of microglia and astrocytes activation rate suggests neuroinflammation. Furthermore, they investigated whether miR-155 and c-Jun are involved in the A β -mediated activation of glial cells. Results showed the upregulation of these two molecules in mouse models and Aβ-activated microglia and astrocytes. Taken all together, miR-155 could be a promising therapeutic factor for the reduction of neuroinflammation in AD (361).

Neuroblastoma differentiation marker 29 (NDM29) is an LncRNA, and its transcription is mediated by RNA pol III (362). Its expression could be influenced by the expression of pro-in-flammatory cytokines such as TNF- α and IL-1 α (363). Interestingly, this LncRNA could induce differentiation of neuroblastoma (NB) cells to a non-malignant neuron-like phenotype (364, 365). Massone *et al.* investigated the role of NDM-

29 on APP synthesis. Results showed an increase in A β secretion and A β 42/A β 40 ratio. Furthermore, expression of this LncRNA and further A β formation could be influenced by inflammation (increase of NDM29 and A β formation). The expression of NDM29 is increased in the brain of neurodegenerative disease patients, indicating that NDM29 provides a situation in which A β could be formed in the extracellular space (366).

ciRS-7, a 1500 nt circular RNA located on chromosome Xq27.1, was first identified in 2011 (367, 368). It is an antisense of cerebellar degeneration-related protein 1 (CDR1AS) without a 3' poly-A tail and a 5' cap, indicating its circular structure. It has more than 70 seed regions for miRNAs, and most of them are for miR-7 (368, 369). Short interspersed nuclear elements upstream and downstream of the *ciRS-7* gene induce its transcription (369). Zhao et al. investigated the ciRS-7 role in AD development. They found a disruption of the ciRS-7-miRNA-7-UBE2A axis in the hippocampal CA1 and Broadmann A22 of the AD patients' brains. ciRS-7 acts as a sponge in the brain, and its deficit leads to higher levels of miR-7, which are further associated with downregulation of this miRNA targets. One of the miR-7 targets is the ubiquitin conjugating enzyme E2A (UBE2A), which is required for the ubiquitin-26S proteasome system, a critical complex for amyloid peptides clearance. Taken all together, the results indicated that dysfunction of the aforementioned axis leads to the formation of amyloid plaque and AD development (370).

Possible Etiologies of Neuroinflammation

Pathogen Aspects of Neuroinflammation and AD

Studies showed that bacterial and viral infections are important in the progression and development of cognitive decline in AD patients. Furthermore, AD itself increases the vulnerability to the effects of peripheral infection with bacteria or viruses (371).

Herpes simplex virus-1 (also called HSV-1) is one of these pathogens involved in AD development. It is a latent infection of the CNS, and studies confirmed the presence of HSV-1 DNA in brain regions that are also involved in AD, such

	Rafiyan	and Mo	itahedi:	Neuroin	flammatio	on in AD
--	---------	--------	----------	---------	-----------	----------

Name	Target	Expression	Function	Reference	
			miRNAs		
miR-206	BDNF	Up	Decrease BDNF, a neuroprotective protein against cell death	(1)	
miR-613	BDNF	Up	Decrease BDNF, a neuroprotective protein against cell death	(2)	
miR-155	SOCS-1	Up	Increase production of pro- inflammatory cytokines such as IL-6 and IFN-β	(3)	
miR-339-5p	BACE-1	Down	Regulation of BACE-1 expression	(4)	
miR-144	ADAM10	Up	important AB inhibitor	(5)	
miR-1908	APOE	Up	Inhibition of APOE expression	(6)	
miR-33	ABCA1	Up	-Impairment of cellular cholesterol efflux -increase Aβ secretion and its clearance inhibition	(7)	
miR-219	Tau	Down	Decrease Tau expression	(8)	
miR-146a	ROCK1	Up	ROCK1/PTEN pathway is involved in Tau hyperphosphorylation in early neurofibrillary tangles	(9)	
miR-200b/c	S6K1	Down	reduction of IRS-1pSer and further Aβ clearance improvement.	(10)	
miR-125b	DUSP6, PPP1CA and Bcl-W	Up	Tau hyperphosphorylation due to downregulation of DUSP6, PPP1CA, and Bcl-W	(11)	
miR-137	CACNA1C	Down	miR-137 and CACNA1C decrease Tau hyperphosphorylation	(12)	
miR-124	PTPN1	Up	Memory deficit and synaptic failure via miR-124/PTPN1	(13)	
miR-134-5p	CREB-1 and BDNF	Up	long-term potentiation (LTP) and synaptic tagging and capture (STC) were disturbed	(14)	
miR-342-5p	AnkG	Up	AnkG is important in the axon initial segment, and its downregulation leads to the axonopathy	(15)	
miR-188-5p	Nrp-2	Down	Reduction of dendritic spine density and mEPSCs, and further cognitive dysfunction	(16)	
miR-34c	SYT1	Up	The ROS-JNK-p53 pathway is important in miR-34c upregulation and further SYT1 downregulation, which are associated with synaptic and cognitive dysfunction	(17)	
miR-214-3p	Atg12	Down	Upregulation of miR-214-3p decreased neuronal apoptosis and autophagy	(18)	
miR-98	HEY-2	Down	miR-98 suppresses apoptosis by inhibition of the Notch signaling pathway via HEY-2 suppression	(19)	
Lnc RNAs					
NDM29	APP	Up	Increase in APP synthesis and further Aβ secretion	(20)	
51A	SORL1	Up	Decreased SORL1 expression leads to impaired APP processing and increased Aβ formation.	(21)	
BACE1-AS	BACE1	Up	BACE1-AS increased BACE1	(22)	

Table 1. The connection of Non-coding RNAs (ncRNAs) with neuroinflan	mmation and Alzheimer's disease
--	---------------------------------

BDNF-AS	BDNF	Up	Decreased viability and induction of apoptosis due to BDNF-AS upregulation	(23)		
SNHG1	miR-137	Up	SNHG1/miR-137/ KREMEN1 axis is involved in neuronal viability and apoptosis	(24)		
NAT-RAD18	RAD18	Up	-	(25)		
NEAT1	CAV2, TGFB2 and TGFBR1	Down	Endocytosis-related genes, which are important in Aβ clearance, and also H3K27Ac and H3K27Cro, which are important for several gene expressions, are affected by NEAT1	(26)		
MALAT1	miR-125b	Down	MALAT1/miR-125b axis regulated neuronal apoptosis and inflammation	(27)		
BC200	BACE1	Up	BC200 decreased cell viability and induced apoptosis and BACE-1 expression	(28)		
CircRNAs						
CircHDAC9	miR-138	Down	circHDAC9/miR-138/Sirt1 axis regulates synaptic function and APP processing	(29)		
Circ-0000950	miR-103	Up	Circ-0000950/miR-103 pathway is involved in neuroinflammation, neurite outgrowth, and neuron apoptosis	(30)		
ciRS-7	miR-7	Down	ciRS-7/miRNA-7/UBE2A axis is involved in the Aβ process and clearance	(31)		

as the hippocampus (372). Additionally, this virus increases amyloid- β production, and it is a prominent risk factor in patients with Apolipoprotein E4 (APOE4) for further AD development (373-375). Another lifelong infection is cytomegalovirus (CMV), which is linked with accelerated cognitive decline and AD development (376, 377). Human Immunodeficiency Virus (HIV) is another chronic lifelong infection that can penetrate the BBB and proliferate in the CNS and cerebrospinal fluid (CSF). This virus could induce chronic neuroinflammation through its gp120 and TAT protein, which exacerbates neurodegeneration. Besides, it increases amyloid deposition in the brain, which could be a risk factor for further AD development (378, 379). Several cohort studies confirmed the increased rate of Human Herpes-Virus (HHV)-6A and HHV-7 infection in AD patients. Like other viruses mentioned above, these viruses cause persistent infection, chronic inflammation, and further glia activation, which could accelerate AD development and progression (380, 381). Some studies confirmed that Epstein-Barr Virus (EBV) is a risk factor for AD development

and progression, especially in APOEɛ4 carriers. Besides, serologic EBV positivity in patients with AD and EBV IgG plasma levels is correlated with cognitive decline and AD progression (376, 382).

Several bacterial pathogens have also been associated with AD. Chlamydia pneumoniae, an obligate intracellular, Gram-negative bacterium, increases the AD development risk up to fivefold (383). It passes through the BBB, infects microglia, astrocytes, and neurons, and causes chronic inflammation. Additionally, it increases Aß deposition in the brain and tends to aggregate in the hippocampus more than in other parts of the brain (384, 385). Heliobacter pylori (H. pylori), another Gram-negative bacterium, is also significantly associated with AD and dementia in the elderly. H. pylori increases the severity of cognitive decline, pro-inflammatory cytokines, and tau protein in the brain (386, 387). An interesting risk factor for AD is periodontitis. Studies showed that healthy elderly individuals with periodontitis have higher levels of amyloid in the CNS (388). Treponema species are involved in periodontitis, and some studies showed an increased sus-

Table 1. Continued

ceptibility to infection with Treponema species in AD patients, but there is no cause-and-effect relationship between these pathogens and AD (389). Borrelia burgdorferi is also involved in periodontitis. It could induce Aß accumulation in the brain through neurons and glia (390). Studies suggest a tenfold increase in the AD development in the context of spirochetes such as B. burgdorferi infection (383). P. gingivalis, a Gram-negative bacterium involved in chronic periodontitis, is a strong risk factor for AD, which could reproduce hallmarks of AD in wild-type mouse models due to the ability of *P. gingivalis* to induce $A\beta$ and tau production in the brain (384, 391). Dominy et al. found a positive correlation between the level of some toxic proteases of P. gingivalis called gingipains in the AD patients' brains and tau and ubiquitin pathology (392). Besides infections, various etiologies can contribute to the initiation of neuroinflammation and the subsequent neurodegeneration.

Metabolic Aspects of Neuroinflammation and AD

Another important aspect of AD is metabolic dysfunction. Metabolic syndrome (MetS), which is a consequence of modern lifestyle, is a risk factor for a wide range of chronic diseases. MetS is characterized by overweight, insulin resistance, high glucose levels, and hypertension, and studies showed that it has a critical role in AD development and progression, especially in LOAD (393). The exact mechanisms are currently unknown, but some studies have suggested that MetS induces neuroinflammation and also increases amyloid plaque production (394, 395). The interaction of MetS and neuroinflammation in the development of AD is mentioned in **Figure 3**.

Diabetes Mellitus (DM)

Diabetes mellitus can potentially induce inflammation in the CNS (396) by two main mechanisms. Firstly, insulin resistance can occur in the CNS, causing an increase in insulin in the blood and the impairment of insulin signaling. An excess amount of insulin induces the secretion of different cytokines and causes inflammation in the CNS. Insulin resistance is a risk factor for cognitive impairment and seems essential for the conversion of these impairments to AD. Molecular mechanisms need to be elucidated, but it seems that insulin resistance causes Ser-phosphorylation of the insulin receptor substrate 1 (IRS1) instead of normal Tyr-phosphorylation. Furthermore, insulin resistance results in decreased phosphorylation of Akt, affecting several downstream components of the insulin pathway, including Glycogen synthase kinase-3 (GSK-3). The increase of unphosphorylated GSK-3 (active form) is correlated with Tau hyperphosphorylation and NFTs formation (397-399). It can also suppress the BBB insulin transporters (400, 401), which are essential for glucose transportation and metabolism across the neurons; therefore, the neurons' available insulin would be decreased (401). Impaired insulin signaling and a decrease in insulin amounts cause neuroinflammation and neurodegeneration. Another important mechanism is amyloidogenesis. In the hyperglycemic state, which can be expected in DM, the APP degradation has been disturbed (402, 403). Thus, the increment of A β can be associated with neuroinflammation and neurodegeneration. These observations have been seen in both streptozotocin (STZ) induced type 1 diabetes mellitus (DMT1) and high-fat diet-induced type 2 diabetes mellitus (DMT2) rodents (404, 405). Cao et al. investigated the role of sugar in AD in a transgenic AD mouse fed with sucrose-sweetened water and reported that, in comparison with the control group, sugar could accelerate the amyloidogenesis and exacerbate AD (405). In another study, Insub et al. found a relationship between AD and DMT2 through A β autoantibodies, as the level of A β autoantibodies was dramatically elevated in the patient serum of T2DM (406). Further studies revealed that diabetes mellitus is a potent risk factor for the development and exacerbation of AD by the acceleration of not only A β but also tau protein production and aggregation (407-409).

Obesity

Obesity induces a hyperinflammatory state, a situation in which inflammatory cytokines increase and immune cells become activated (410, 411). Although the mechanism of how obesity leads to a hyperinflammation state has not been completely understood, some studies suggested the role of leptin as an essential hormone for conducting inflammation in obese patients (412-

414). Leptin has a similar structure to cytokines (415) and has receptors on immune cells such as macrophages, T cells, and microglia. The activated microglia by leptin can cause IL-6 and IL-1B production (416, 417). Another important mechanism is the contribution of obesity to metabolic syndrome. Increased body fat, accompanied by adipocyte hypertrophy and hyperplasia, excessive cholesterol and glucose in blood, induces stress in adipocytes, causing the secretion of TNF-a and IL-6 alongside the ROS activation, causing a hyperinflammatory state in the body and increasing the risk of neuroinflammation and degeneration (67, 418-421). Additionally, ROS disrupts the BBB as detected by the serum increase of calcium-binding protein B (S100B), a glial-specific protein expressed primarily in astrocytes, and neuron-specific enolase (NSE)(422, 423). Therefore, BBB dysfunction could result in altered permeability and cerebrovascular integrity loss in the human hippocampus, a region involved in learning and memory that is early damaged in AD (423-425). Brain cholesterol levels directly influence Aβ formation through stimulation of the amyloidogenic pathway, since different experiments strongly suggest that cholesterol has an elevated affinity for APP and A β (423, 426, 427). Another possible mechanism for the hyperinflammatory state in obesity is the endoplasmic reticulum stress. ER is an important site for protein synthesis and folding, and the stress resulting from fat deposition and compression of adipocytes increases the need for protein and protein synthesis, triggering unfolded protein response (UPR) in it. UPR is the accumulation of unfolded proteins in the ER, which causes the pro-inflammatory cytokine release from adipocytes and inflammation (428-430). Obesity has a strong correlation with DMT2 and insulin resistance. ROS, hyperinflammation state, adiponectin secretion dysfunction, Excess lipid substrates and lipotoxicity, and changes in the gut microbiome are the main causes of insulin resistance in obesity. As mentioned above, insulin resistance could exacerbate amyloidogenesis and AD (431-433).

Gut Microbiome

The microbiota-gut-brain axis is a mutual communication system that is connected via neural, immune, endocrine, and metabolic pathways.

Alteration in gut microbiome is associated not only with gastrointestinal disorders but also with some neurodegenerative disorders, such as AD (434). Gut microbiome dysbiosis can increase the permeability of both the intestine and the blood-brain-barrier and expose the CNS to some microbiome products, such as lipopolysaccharides (LPS) and small-chain fatty acids (SCFAs). These products could induce inflammation, increase the production of pro-inflammatory cytokines, and facilitate neuronal apoptosis (435, 436). Besides, dysbiosis could induce an unresolved inflammation in the gut and activate immune system cells such as CD4⁺ T cells. These cells could further pass through the BBB and induce neuroinflammation (437). Microglia activation can also occur in this context and not only exacerbate the neuroinflammation but also impact astrocytes indirectly via microglia-astrocyte communication (438). Studies showed that Bacteroidetes, Rikenellaceae, and Tenericutes were increased, and Firmicutes, Verrucomicrobia, Proteobacteria, Akkermansia, Allobacilum, and Actinobacteria were decreased in the gut in animal models of AD. These changes could enhance amyloid production and plaque-localized inflammation in the brain by a change in the activation of glia in the brain (439-442). Germ-free mice studies revealed a reduction in brain-derived neurotrophic factor in germ-free mice. This factor is essential for synaptic plasticity and cognitive function, besides studies confirmed its reduced expression in AD (443, 444). Probiotics are living microorganisms ingested for some gastrointestinal disorders as they can modulate the gut microbiome. Several studies confirmed that certain probiotics, such as Lactobacillus helveticus NS8, could ameliorate cognitive impairments and restore brain-derived neurotrophic factor (BDNF) content in the brain in the context of chronic stress (445). However, further studies are needed to determine the best combination of different probiotics for the maximum effect. The gut microbiome could also act directly through the vagus nerve. The acetylcholine neurotransmitter is released in response to vagus nerve stimulation and modulates the inflammation of the CNS by controlling the activity of immune cells. Thus, gut microbiome could alter the secretion of acetylcholine, and further changes in immune system function (446, 447).

Treatments for AD

Development of new drugs and therapeutic targets is an urgent need for AD due to its severe cognitive and neuropsychiatric symptoms. More than 100 agents in more than 150 clinical trials are in progress for the treatment of this disease, and disease-modifying therapies (DMTs) are the most common agents. More than 20 agents are in phase 3 of clinical trials (448). Treatment for the AD is classified into different types based on their mechanisms of action and several other variables. Two main types based on their mechanisms of the action are DMTs such as Aducanumab, Atuzaginstat (COR388), Azeliragon, and Blarcamesine (ANAVEX2-73), which act via different mechanisms including monoclonal antibody against AB plaques, reduction of neurodegeneration and neuroinflammation by inhibition of P. gingivalis protease inhibitor, reduction of inflammation by antagonizing RAGE and inhibition of A β transport to the brain, reduction of oxidative stress, protein misfolding, mitochondrial dysfunction, and inflammation by targeting M2 and Sigma-1 receptors respectively. These are some of the common mechanisms of the DMT drugs for the reduction of AD development and symptoms (449-452). Another group is neuropsychiatric and cognitive symptoms relievers such as AVP-786, Brexpiprazole, Ginkgo biloba, Guanfacine, and Nabilone. These drugs act via different mechanisms, including inhibition of acetylcholinesterase, inactivating NMDA receptors, and activation of Sigmal receptors (453-457). These two types of AD treatments (DMT and neuropsychiatric/ cognitive symptoms reliever) account for about 86% and 13% of all treatments, respectively (448). Additionally, there is a one-vaccine trial in the phase 3 (CAD106)(448). CAD106 is composed of different copies of A β 1–6 with a carrier that is composed of 180 copies of bacteriophage QB coat protein. This vaccine induces Aß antibodies without involvement of Aβ-specific T-cell response (458-460). Collectively, there are numerous treatments under investigation for AD treatment, and each of them acts via specific mechanisms. These drugs make new hopes for AD treatment soon. Targeting signaling pathways, especially inflammatory pathways and pro-inflammatory cytokines, to reduce neuronal damage and AD progression is another strategy. Anakinra and

rilonacept are developed against IL-1, and canakinumab is against IL-1 β (461-463). Several drugs are also developed based on the NLRP3 pathway. JC124, a NLRP3 inflammasome and caspase-1 inhibitor, CY-09, which binds to the NACHT domain to inhibit NLRP3 ATPase activity, and several similar drugs are among this group (464, 465). Use of curcumin, phytochemicals such as Resveratrol, MW01-2-069A-SRM, a p38a MAPK inhibitor, are the strategies to inhibit NF-KB signaling pathway (466-468). Nicotinamide riboside (NR) is a promising therapeutic target for normalizing cGAS-STING in preclinical studies (469). Targeting macrophages could also be a successful strategy. Increase in microglial phagocytosis and switch from M1 to M2 are two main strategies. PPARa is a nuclear receptor that promotes microglia recruitment and phagocytosis and further increases Aβ clearance. Gemfibrozil and Wy14643 are two PPARa agonists used to increase autophagy of microglia and structural neuroplasticity (469, 470). Another member of the PPAR family is PPARy with opposite effects. PPARy antagonist T0070907 could enhance microglial autophagy via Liver kinase B1 (LKB1)-AMPK pathway (470). LC3 is an important part of autophagy and phagosome formation. Anti-inflammatory drugs and cytokines such as dimethyl fumarate (DMF) and IL-4 could be used for upregulation of LC3 and increased microglial autophagy (471-474). Switching from M1 to M2 macrophages due to their beneficial effects against AD progression could be used as another strategy for AD treatment. L-cysteine-derived hydrogen sulfide (H2S), CaMKK inhibitor (STO-609), and T0070907 are used for this strategy (475-478).

Conclusion

AD is a multifactorial disease, and different mechanisms are involved in its pathogenesis. Inflammation can be considered a principal mechanism in AD initiation and progression. Various studies have been done over the years to examine different parts of inflammation in the brain. Pro-inflammatory cytokines and inflammatory pathways are vital parts of inflammation. They are double-edged, either having a preventive and protective role against diseases or being harmful and damaging to the cells. Overall, in acute immune response, activation of inflammatory pathways and production of pro-inflammatory cytokines are essential for proper immune system function, but if the immune system's stimulator remained, the inflammation could be chronic, and in this situation, the cytokines can be harmful to the body the same as what occurs in Alzheimer's disease. This article summarizes and discusses different aspects of AD, from its molecular pathways to gut microbiome and current treatments and clinical trials, and provides a new insight into AD development and its promising therapeutic targets. Further studies would be needed to investigate the hidden aspects of this disease.

Funding

This research did not receive any specific grant from funding agencies in the public, commercial, or not-for-profit sectors.

Conflict of Interest

Authors approve that they have no conflict of interest.

References

- 1. Litke R, Garcharna LC, Jiwani SNeugroschl J. Modifiable Risk Factors in Alzheimer Disease and Related Dementias: A Review. Clin Ther. 2021.
- Scheltens P, Blennow K, Breteler M, De Strooper B, Frisoni GSalloway S. Van der Flier WM: Alzheimer's disease. Lancet. 2016;388(10043):505-17.
- Mayeux RStern Y. Epidemiology of Alzheimer disease. Cold Spring Harb Perspect Med. 2012;2(8):a006239.
- 4. Grant WB, Campbell A, Itzhaki RFSavory J. The significance of environmental factors in the etiology of Alzheimer's disease. J Alzheimers Dis. 2002;4:179-89.
- 5. Khanahmadi M, Farhud DDMalmir M. Genetic of Alzheimer's disease: A narrative review article. Iran J Public Health. 2015;44(7):892.
- 6. Azizi GMirshafiey A. The potential role of proinflammatory and antiinflammatory cytokines in Alzheimer disease pathogenesis. Immunopharmacol Immunotoxicol. 2012;34(6):881-95.
- Lanoiselée H-M, Nicolas G, Wallon D, Rovelet-Lecrux A, Lacour M, Rousseau S, et al. APP, PSEN1, and PSEN2 mutations in early-onset Alzheimer disease: A genetic screening study of familial and sporadic cases. PLoS Med. 2017;14(3):e1002270.
- 8. Fuster-Matanzo A, Llorens-Martín M, Hernández FAvila J. Role of neuroinflammation in adult neurogenesis and Alzheimer disease: therapeutic

approaches. Mediators Inflamm. 2013;2013.

- Chen L, Deng H, Cui H, Fang J, Zuo Z, Deng J, et al. Inflammatory responses and inflammation-associated diseases in organs. Oncotarget. 2017;9(6):7204-18.
- 10. Fakhoury M. Microglia and astrocytes in Alzheimer's disease: Implications for therapy. Curr Neuropharmacol. 2018;16(5):508-18.
- 11. Leng FEdison P. Neuroinflammation and microglial activation in Alzheimer disease: where do we go from here? Nat Rev Neurol. 2020:1-16.
- 12. Zotova E, Nicoll JA, Kalaria R, Holmes CBoche D. Inflammation in Alzheimer's disease: relevance to pathogenesis and therapy. Alzheimers Res Ther. 2010;2(1):1-9.
- 13. Lyman M, Lloyd DG, Ji X, Vizcaychipi MPMa D. Neuroinflammation: the role and consequences. Neurosci Res. 2014;79:1-12.
- Augusto-Oliveira M, Arrifano GP, Lopes-Araújo A, Santos-Sacramento L, Takeda PY, Anthony DC, et al. What do microglia really do in healthy adult brain? Cells. 2019;8(10):1293.
- 15. Bourgognon J-MCavanagh J. The role of cytokines in modulating learning and memory and brain plasticity. Brain Neurosci Adv. 2020;4:2398212820979802.
- Kinney JW, Bemiller SM, Murtishaw AS, Leisgang AM, Salazar AMLamb BT. Inflammation as a central mechanism in Alzheimer's disease. Alzheimers Dement Transl Res Clin Interv. 2018;4:575-90.
- 17. Sun Y, Ma C, Sun H, Wang H, Peng W, Zhou Z, et al. Metabolism: a novel shared link between diabetes mellitus and Alzheimer's disease. J Diabetes Res. 2020;2020.
- de Araújo Boleti AP, de Oliveira Flores TM, Moreno SE, Dos Anjos L, Mortari MRMigliolo L. Neuroinflammation: An overview of neurodegenerative and metabolic diseases and of biotechnological studies. Neurochem Int. 2020;136:104714.
- Gauthier S, Aisen P, Cummings J, Detke M, Longo F, Raman R, et al. Non-amyloid approaches to disease modification for Alzheimer's disease: an EU/ US CTAD Task Force Report. J Prev Alzheimers Dis. 2020;7:152-57.
- Gouras GK, Tampellini D, Takahashi RHCapetillo-Zarate E. Intraneuronal β-amyloid accumulation and synapse pathology in Alzheimer's disease. Acta Neuropathol. 2010;119(5):523-41.
- 21. Thinakaran GKoo EH. Amyloid precursor protein trafficking, processing, and function. J Biol Chem. 2008;283(44):29615-19.
- 22. Nalivaeva NNTurner AJ. The amyloid precursor protein: A biochemical enigma in brain de-

velopment, function and disease. FEBS Lett. 2013;587(13):2046-54.

- 23. Volloch VRits S. Results of beta secretase-inhibitor clinical trials support amyloid precursor protein-independent generation of beta amyloid in sporadic Alzheimer's disease. Med Sci. 2018;6(2):45.
- 24. Selkoe DJWolfe MS. Presenilin: running with scissors in the membrane. Cell. 2007;131(2):215-21.
- 25. Kempuraj D, Thangavel R, Selvakumar GP, Zaheer S, Ahmed ME, Raikwar SP, et al. Brain and peripheral atypical inflammatory mediators potentiate neuroinflammation and neurodegeneration. Front Cell Neurosci. 2017;11:216.
- 26. Wang W, Zhao F, Ma X, Perry GZhu X. Mitochondria dysfunction in the pathogenesis of Alzheimer's disease: Recent advances. Mol Neurodegener. 2020;15(1):1-22.
- 27. Kempuraj D, Mentor S, Thangavel R, Ahmed ME, Selvakumar GP, Raikwar SP, et al. Mast cells in stress, pain, blood-brain barrier, neuroinflammation and Alzheimer's disease. Front Cell Neurosci. 2019:54.
- 28. Arranz AMDe Strooper B. The role of astroglia in Alzheimer's disease: pathophysiology and clinical implications. Lancet Neurol. 2019;18(4):406-14.
- 29. Chen G-f, Xu T-h, Yan Y, Zhou Y-r, Jiang Y, Melcher K, et al. Amyloid beta: structure, biology and structure-based therapeutic development. Acta Pharmacol Sin. 2017;38(9):1205-35.
- Gao Y, Tan L, Yu JTTan L. Tau in Alzheimer's Disease: Mechanisms and Therapeutic Strategies. Curr Alzheimer Res. 2018;15(3):283-300.
- 31. Lee G, Newman ST, Gard DL, Band HPanchamoorthy G. Tau interacts with src-family non-receptor tyrosine kinases. J Cell Sci. 1998;111(21):3167-77.
- 32. Kitagishi Y, Nakanishi A, Ogura YMatsuda S. Dietary regulation of PI3K/AKT/GSK-3β pathway in Alzheimer's disease. Alzheimers Res Ther. 2014;6(3):1-7.
- 33. Li Y, Liu L, Barger SWGriffin WST. Interleukin-1 mediates pathological effects of microglia on tau phosphorylation and on synaptophysin synthesis in cortical neurons through a p38-MAPK pathway. J Neurosci. 2003;23(5):1605-11.
- 34. Ghosh S, Wu MD, Shaftel SS, Kyrkanides S, LaFerla FM, Olschowka JA, et al. Sustained interleukin-1β overexpression exacerbates tau pathology despite reduced amyloid burden in an Alzheimer's mouse model. J Neurosci. 2013;33(11):5053-64.
- Farfel JM, Yu L, De Jager PL, Schneider JABennett DA. Association of APOE with tau-tangle pathology with and without β-amyloid. Neurobiol Aging. 2016;37:19-25.

- 36. Bedse G, Di Domenico F, Serviddio GCassano T. Aberrant insulin signaling in Alzheimer's disease: current knowledge. Front Neurosci. 2015;9:204.
- 37. Gauthier S, Feldman HH, Schneider LS, Wilcock GK, Frisoni GB, Hardlund JH, et al. Efficacy and safety of tau-aggregation inhibitor therapy in patients with mild or moderate Alzheimer's disease: a randomised, controlled, double-blind, parallel-arm, phase 3 trial. Lancet. 2016;388(10062):2873-84.
- Congdon EESigurdsson EM. Tau-targeting therapies for Alzheimer disease. Nat Rev Neurol. 2018;14(7):399-415.
- 39. Griffin W, Stanley L, Ling C, White L, MacLeod V, Perrot L, et al. Brain interleukin 1 and S-100 immunoreactivity are elevated in Down syndrome and Alzheimer disease. Proc Natl Acad Sci U S A. 1989;86(19):7611-15.
- 40. Grande G, Marengoni A, Vetrano DL, Roso-Llorach A, Rizzuto D, Zucchelli A, et al. Multimorbidity burden and dementia risk in older adults: The role of inflammation and genetics. Alzheimers Dement. 2021;17(5):768-776.
- 41. Borish LCSteinke JW. 2. Cytokines and chemokines. J Allergy Clin Immunol. 2003;111(2 Suppl):S460-75.
- 42. Sahibzada HA, Khurshid Z, Khan RS, Naseem M, Siddique KM, Mali M, et al. Salivary IL-8, IL-6 and TNF-α as potential diagnostic biomarkers for oral cancer. Diagnostics. 2017;7(2):21.
- 43. Kim YK, Na KS, Myint AMLeonard BE. The role of pro-inflammatory cytokines in neuroinflammation, neurogenesis and the neuroendocrine system in major depression. Prog Neuropsychopharmacol Biol Psychiatry. 2016;64:277-84.
- 44. Megur A, Baltriukienė D, Bukelskienė VBurokas A. The Microbiota–Gut–Brain Axis and Alzheimer's Disease: Neuroinflammation Is to Blame? Nutrients. 2021;13(1):37.
- 45. Hoffman WH, Casanova MF, Cudrici CD, Zakranskaia E, Venugopalan R, Nag S, et al. Neuroinflammatory response of the choroid plexus epithelium in fatal diabetic ketoacidosis. Exp Mol Pathol. 2007;83(1):65-72.
- 46. Liu B, Wang K, Gao HM, Mandavilli B, Wang JY-Hong JS. Molecular consequences of activated microglia in the brain: overactivation induces apoptosis. J Neurochem. 2001;77(1):182-89.
- 47. De Simone R, Ajmone-Cat MAMinghetti L. Atypical antiinflammatory activation of microglia induced by apoptotic neurons. Mol Neurobiol. 2004;29(2):197-212.
- 48. Smith JA, Das A, Ray SKBanik NL. Role of pro-inflammatory cytokines released from microglia

in neurodegenerative diseases. Brain Res Bull. 2012;87(1):10-20.

- Brown GCNeher JJ. Microglial phagocytosis of live neurons. Nat Rev Neurosci. 2014;15(4):209-16.
- Dong YYong VW. When encephalitogenic T cells collaborate with microglia in multiple sclerosis. Nat Rev Neurol. 2019;15(12):704-17.
- 51. Morianos I, Trochoutsou AI, Papadopoulou G, Semitekolou M, Banos A, Konstantopoulos D, et al. Activin-A limits Th17 pathogenicity and autoimmune neuroinflammation via CD39 and CD73 ectonucleotidases and Hif1-α-dependent pathways. Proc Natl Acad Sci U S A. 2020;117(22):12269-80.
- 52. Tesmer LA, Lundy SK, Sarkar SFox DA. Th17 cells in human disease. Immunol Rev. 2008;223:87-113.
- 53. Ziv Y, Ron N, Butovsky O, Landa G, Sudai E, Greenberg N, et al. Immune cells contribute to the maintenance of neurogenesis and spatial learning abilities in adulthood. Nat Neurosci. 2006;9(2):268-75.
- 54. Derecki NC, Cardani AN, Yang CH, Quinnies KM, Crihfield A, Lynch KR, et al. Regulation of learning and memory by meningeal immunity: a key role for IL-4. J Exp Med. 2010;207(5):1067-80.
- 55. Lei C, Wu B, Cao T, Liu MHao Z. Brain recovery mediated by toll-like receptor 4 in rats after intracerebral hemorrhage. Brain Res. 2016;1632:1-8.
- 56. Danilov AI, Covacu R, Moe MC, Langmoen IA, Johansson CB, Olsson T, et al. Neurogenesis in the adult spinal cord in an experimental model of multiple sclerosis. Eur J Neurosci. 2006;23(2):394-400.
- 57. Ishii H, Jin X, Ueno M, Tanabe S, Kubo T, Serada S, et al. Adoptive transfer of Th1-conditioned lymphocytes promotes axonal remodeling and functional recovery after spinal cord injury. Cell Death Dis. 2012;3(8):e363-e63.
- 58. Hauben E, Butovsky O, Nevo U, Yoles E, Moalem G, Agranov E, et al. Passive or active immunization with myelin basic protein promotes recovery from spinal cord contusion. J Neurosci. 2000;20(17):6421-30.
- 59. Yong HYF, Rawji KS, Ghorbani S, Xue MYong VW. The benefits of neuroinflammation for the repair of the injured central nervous system. Cell Mol Immunol. 2019;16(6):540-46.
- 60. Plascencia-Villa GPerry G. Preventive and Therapeutic Strategies in Alzheimer's Disease: Focus on Oxidative Stress, Redox Metals, and Ferroptosis. Antioxid Redox Signal. 2021;34(8):591-610.
- 61. Lashley T, Schott JM, Weston P, Murray CE, Wellington H, Keshavan A, et al. Molecular biomark-

ers of Alzheimer's disease: progress and prospects. Dis Model Mech. 2018;11(5).

- 62. Cai Y, Liu J, Wang B, Sun MYang H. Microglia in the neuroinflammatory pathogenesis of Alzheimer's disease and related therapeutic targets. Front Immunol. 2022:1868.
- 63. Deczkowska A, Keren-Shaul H, Weiner A, Colonna M, Schwartz MAmit I. Disease-Associated Microglia: A Universal Immune Sensor of Neurodegeneration. Cell. 2018;173(5):1073-81.
- 64. Ulland TKColonna M. TREM2—a key player in microglial biology and Alzheimer disease. Nat Rev Neurol. 2018;14(11):667-75.
- 65. Yao KZu H-b. Microglial polarization: novel therapeutic mechanism against Alzheimer's disease. Inflammopharmacology. 2020;28(1):95-110.
- 66. Cui W, Sun C, Ma Y, Wang S, Wang XZhang Y. Inhibition of TLR4 induces M2 microglial polarization and provides neuroprotection via the NLRP3 inflammasome in Alzheimer's disease. Front Neurosci. 2020;14:444.
- 67. Rondinone CM. Adipocyte-derived hormones, cytokines, and mediators. Endocrine. 2006;29(1):81-90.
- 68. Wang W-Y, Tan M-S, Yu J-TTan L. Role of pro-inflammatory cytokines released from microglia in Alzheimer's disease. Ann Transl Med. 2015;3(10):136-36.
- 69. Sastre M, Klockgether THeneka MT. Contribution of inflammatory processes to Alzheimer's disease: molecular mechanisms. Int J Dev Neurosci. 2006;24(2-3):167-76.
- 70. Ojala J, Alafuzoff I, Herukka S-K, van Groen T, Tanila HPirttilä T. Expression of interleukin-18 is increased in the brains of Alzheimer's disease patients. Neurobiol Aging. 2009;30(2):198-209.
- 71. Abbas N, Bednar I, Mix E, Marie S, Paterson D, Ljungberg A, et al. Up-regulation of the inflammatory cytokines IFN-γ and IL-12 and down-regulation of IL-4 in cerebral cortex regions of APPSWE transgenic mice. J Neuroimmunol. 2002;126(1-2):50-57.
- 72. Mulder SD, Nielsen HM, Blankenstein MA, Eikelenboom PVeerhuis R. Apolipoproteins E and J interfere with amyloid-beta uptake by primary human astrocytes and microglia in vitro. Glia. 2014;62(4):493-503.
- 73. Maezawa I, Zimin PI, Wulff HJin L-W. Amyloid- β protein oligomer at low nanomolar concentrations activates microglia and induces microglial neurotoxicity. J Biol Chem. 2011;286(5):3693-706.
- 74. Trojan E, Tylek K, Schröder N, Kahl I, Brandenburg L-O, Mastromarino M, et al. The N-Formyl Peptide Receptor 2 (FPR2) Agonist MR-39

Improves Ex Vivo and In Vivo Amyloid Beta (1–42)-Induced Neuroinflammation in Mouse Models of Alzheimer's Disease. Mol Neurobiol. 2021;58(12):6203-21.

- 75. Nakanishi A, Kaneko N, Takeda H, Sawasaki T, Morikawa S, Zhou W, et al. Amyloid β directly interacts with NLRP3 to initiate inflammasome activation: identification of an intrinsic NLRP3 ligand in a cell-free system. Inflamm Regen. 2018;38:27.
- 76. Bonaiuto C, McDonald PP, Rossi FCassatella MA. Activation of nuclear factor-kappa B by beta-amyloid peptides and interferon-gamma in murine microglia. J Neuroimmunol. 1997;77(1):51-6.
- 77. Ma L-Y, Liu S-F, Du J-H, Niu Y, Hou P-F, Shu Q, et al. Chronic ghrelin administration suppresses IKK/NF- κ B/BACE1 mediated Aβ production in primary neurons and improves cognitive function via upregulation of PP1 in STZ-diabetic rats. Neurobiol Learn Mem. 2020;169:107155.
- 78. Qiao A, Li J, Hu Y, Wang JZhao Z. Reduction BACE1 expression via suppressing NF-κB mediated signaling by Tamibarotene in a mouse model of Alzheimer's disease. IBRO Neurosci Rep. 2021;10:153-60.
- 79. Swanson KV, Deng MTing JP-Y. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. Nat Rev Immunol. 2019;19(8):477-89.
- Zinatizadeh MR, Schock B, Chalbatani GM, Zarandi PK, Jalali SAMiri SR. The Nuclear Factor Kappa B (NF-kB) signaling in cancer development and immune diseases. Genes Dis. 2021;8(3):287-97.
- Thawkar BSKaur G. Inhibitors of NF-κB and P2X7/NLRP3/Caspase 1 pathway in microglia: Novel therapeutic opportunities in neuroinflammation induced early-stage Alzheimer's disease. J Neuroimmunol. 2019;326:62-74.
- 82. Hong Y, Liu Y, Yu D, Wang MHou Y. The neuroprotection of progesterone against Aβ-induced NLRP3-Caspase-1 inflammasome activation via enhancing autophagy in astrocytes. Int Immunopharmacol. 2019;74:105669.
- Blasko I, Stampfer-Kountchev M, Robatscher P, Veerhuis R, Eikelenboom PGrubeck-Loebenstein B. How chronic inflammation can affect the brain and support the development of Alzheimer's disease in old age: the role of microglia and astrocytes. Aging Cell. 2004;3(4):169-76.
- 84. De Vellis J. Neuroglia in the aging brain: Springer Science & Business Media; 2001.
- 85. Sidoryk-Wegrzynowicz M, Wegrzynowicz M, Lee E, Bowman ABAschner M. Role of astrocytes in brain function and disease. Toxicol Pathol.

2011;39(1):115-23.

- 86. Phillips EC, Croft CL, Kurbatskaya K, O'Neill MJ, Hutton ML, Hanger DP, et al. Astrocytes and neuroinflammation in Alzheimer's disease. Portland Press Ltd.; 2014.
- 87. Westacott LJ, Haan N, Evison C, Marei O, Hall J, Hughes TR, et al. Dissociable effects of complement C3 and C3aR on survival and morphology of adult born hippocampal neurons, pattern separation, and cognitive flexibility in male mice. Brain Behav Immun. 2021;98:136-50.
- 88. Xu X, Zhang A, Zhu Y, He W, Di W, Fang Y, et al. MFG-E8 reverses microglial-induced neurotoxic astrocyte (A1) via NF-κB and PI3K-Akt pathways. J Cell Physiol. 2019;234(1):904-14.
- 89. Tarczyluk MA, Nagel DA, Parri HR, Tse EH, Brown JE, Coleman MD, et al. Amyloid β 1-42 induces hypometabolism in human stem cell-derived neuron and astrocyte networks. J Cereb Blood Flow Metab. 2015;35(8):1348-57.
- 90. Simpson IA, Chundu KR, Davies-Hill T, Honer WGDavies P. Decreased concentrations of GLUT1 and GLUT3 glucose transporters in the brains of patients with Alzheimer's disease. Ann Neurol. 1994;35(5):546-51.
- 91. Simpson IA, Carruthers AVannucci SJ. Supply and demand in cerebral energy metabolism: the role of nutrient transporters. J Cereb Blood Flow Metab. 2007;27(11):1766-91.
- Berkenbosch F, Van Oers J, Del Rey A, Tilders FBesedovsky H. Corticotropin-releasing factor-producing neurons in the rat activated by interleukin-1. Science. 1987;238(4826):524-26.
- 93. March CJ, Mosley B, Larsen A, Cerretti DP, Braedt G, Price V, et al. Cloning, sequence and expression of two distinct human interleukin-1 complementary DNAs. Nature. 1985;315(6021):641-47.
- 94. Thornberry NA, Bull HG, Calaycay JR, Chapman KT, Howard AD, Kostura MJ, et al. A novel heterodimeric cysteine protease is required for interleukin-1β processing in monocytes. Nature. 1992;356(6372):768-74.
- 95. Subramaniam S, Stansberg CCunningham C. The interleukin 1 receptor family. Dev Comp Immunol. 2004;28(5):415-28.
- 96. Korherr C, Hofmeister R, Wesche HFalk W. A critical role for interleukin-1 receptor accessory protein in interleukin-1 signaling. Eur J Immunol. 1997;27(1):262-67.
- 97. O'Léime CS, Cryan JFNolan YM. Nuclear deterrents: intrinsic regulators of IL-1β-induced effects on hippocampal neurogenesis. Brain Behav Immun. 2017;66:394-412.
- 98. Molgora M, Supino D, Mantovani AGarlanda C.

Tuning inflammation and immunity by the negative regulators IL-1R2 and IL-1R8. Immunol Rev. 2018;281(1):233-47.

- 99. Allan SM, Tyrrell PJRothwell NJ. Interleukin-1 and neuronal injury. Nat Rev Immunol. 2005;5(8):629-40.
- 100. Hannum CH, Wilcox CJ, Arend WP, Joslin FG, Dripps DJ, Heimdal PL, et al. Interleukin-1 receptor antagonist activity of a human interleukin-1 inhibitor. Nature. 1990;343(6256):336-40.
- 101. Malyak M, Smith MF, Abel AA, Hance KRArend WP. The differential production of three forms of IL-1 receptor antagonist by human neutrophils and monocytes. J Immunol. 1998;161(4):2004-10.
- 102. Du Y, Dodel R, Eastwood B, Bales K, Gao F, Lohmüller F, et al. Association of an interleukin 1α polymorphism with Alzheimer's disease. Neurology. 2000;55(4):480-84.
- 103. Grimaldi LM, Casadei VM, Ferri C, Veglia F, Licastro F, Annoni G, et al. Association of early-onset Alzheimer's disease with an interleukin-1α gene polymorphism. Ann Neurol. 2000;47(3):361-65.
- 104. Déniz-Naranjo M, Muñoz-Fernandez C, Alemany-Rodríguez M, Pérez-Vieitez M, Aladro-Benito Y, Irurita-Latasa J, et al. Cytokine IL-1 beta but not IL-1 alpha promoter polymorphism is associated with Alzheimer disease in a population from the Canary Islands, Spain. Eur J Neurol. 2008;15(10):1080-84.
- 105. Babić Leko M, Nikolac Perković M, Klepac N, Štrac DŠ, Borovečki F, Pivac N, et al. IL-1β, IL-6, IL-10, and TNF α Single Nucleotide Polymorphisms in Human Influence the Susceptibility to Alzheimer's Disease Pathology. J Alzheimers Dis. 2020(Preprint):1-19.
- 106. Mason JL, Suzuki K, Chaplin DDMatsushima GK. Interleukin-1β promotes repair of the CNS. J Neurosci. 2001;21(18):7046-52.
- 107. Landgraf R, Neumann I, Holsboer FPittman QJ. Interleukin-1 β stimulates both central and peripheral release of vasopressin and oxytocin in the rat. Eur J Neurosci. 1995;7(4):592-98.
- 108. Sapolsky R, Rivier C, Yamamoto G, Plotsky PVale W. Interleukin-1 stimulates the secretion of hypothalamic corticotropin-releasing factor. Science. 1987;238(4826):522-24.
- 109. Uehara A, Gottschall PE, Dahl RRArimura A. Interleukin-1 stimulates ACTH release by an indirect action which requires endogenous corticotropin releasing factor. Endocrinology. 1987;121(4):1580-82.
- 110. Goldgaber D, Harris HW, Hla T, Maciag T, Donnelly RJ, Jacobsen JS, et al. Interleukin 1 regulates synthesis of amyloid beta-protein precursor

mRNA in human endothelial cells. Proc Natl Acad Sci U S A. 1989;86(19):7606-10.

- 111. Basu A, Krady JKLevison SW. Interleukin-1: a master regulator of neuroinflammation. J Neurosci Res. 2004;78(2):151-56.
- 112. Kopitar-Jerala N. Innate Immune Response in Brain, NF-Kappa B Signaling and Cystatins. Front Mol Neurosci. 2015;8(73).
- 113. Shang D, Hong Y, Xie W, Tu ZXu J. Interleukin-1β Drives Cellular Senescence of Rat Astrocytes Induced by Oligomerized Amyloid β Peptide and Oxidative Stress. Front Neurol. 2020;11.
- 114. John GR, Lee SC, Song X, Rivieccio MBrosnan CF. IL-1-regulated responses in astrocytes: Relevance to injury and recovery. Glia. 2005;49(2):161-76.
- 115. Wyble CW, Hynes KL, Kuchibhotla J, Marcus BC, Hallahan DGewertz BL. TNF-alpha and IL-1 upregulate membrane-bound and soluble E-selectin through a common pathway. J Surg Res. 1997;73(2):107-12.
- 116. Calkins CM, Bensard DD, Shames BD, Pulido EJ, Abraham E, Fernandez N, et al. IL-1 regulates in vivo C-X-C chemokine induction and neutrophil sequestration following endotoxemia. J Endotoxin Res. 2002;8(1):59-67.
- 117. Allan SMRothwell NJ. Cytokines and acute neurodegeneration. Nat Rev Neurosci. 2001;2(10):734-44.
- 118. Dubois C, Prevarskaya NAbeele F. The calcium-signaling toolkit: Updates needed. Biochim Biophys Acta Mol Cell Res. 2015;1863.
- 119. Yang S, Liu Z-W, Wen L, Qiao H-F, Zhou W-XZhang Y-X. Interleukin-1β enhances NMDA receptor-mediated current but inhibits excitatory synaptic transmission. Brain Res. 2005;1034(1-2):172-79.
- 120. Viviani B, Bartesaghi S, Gardoni F, Vezzani A, Behrens M, Bartfai T, et al. Interleukin-1 β enhances NMDA receptor-mediated intracellular calcium increase through activation of the Src family of kinases. J Neurosci. 2003;23(25):8692-700.
- 121. Marambaud P, Dreses-Werringloer UVingtdeuxV. Calcium signaling in neurodegeneration. Mol Neurodegener. 2009;4:20.
- 122. Pivovarova NBAndrews SB. Calcium-dependent mitochondrial function and dysfunction in neurons. FEBS J. 2010;277(18):3622-36.
- 123. Guo Y, Hao DHu H. High doses of dexamethasone induce endoplasmic reticulum stress-mediated apoptosis by promoting calcium ion influx-dependent CHOP expression in osteoblasts. Mol Biol Rep. 2021;48(12):7841-51.
- 124. Urbańska-Ryś H, Wiersbowska A, Stepień HRobak T. Relationship between circulating interleu-

kin-10 (IL-10) with interleukin-6 (IL-6) type cytokines (IL-6, interleukin-11 (IL-11), oncostatin M (OSM)) and soluble interleukin-6 (IL-6) receptor (sIL-6R) in patients with multiple myeloma. Eur Cytokine Netw. 2000;11(3):443-51.

- 125. Rose-John S. Interleukin-6 family cytokines. Cold Spring Harb Perspect Biol. 2018;10(2):a028415.
- 126. Murakami M, Kamimura DHirano T. Mini ReviewNew IL-6 (gp130) Family Cytokine Members, CLC/NNT1/BSF3 and IL-27. Growth Factors. 2004;22(2):75-77.
- 127. Boulanger MJ, Chow D-c, Brevnova EEGarcia KC. Hexameric structure and assembly of the interleukin-6/IL-6 α-receptor/gp130 complex. Science. 2003;300(5628):2101-04.
- 128. Simpson RJ, Hammacher A, Smith DK, Matthews JMWard LD. Interleukin-6: Structure-function relationships. Protein Sci. 1997;6(5):929-55.
- 129. Wolf J, Rose-John SGarbers C. Interleukin-6 and its receptors: a highly regulated and dynamic system. Cytokine. 2014;70(1):11-20.
- 130. Jones SA, Scheller JRose-John S. Therapeutic strategies for the clinical blockade of IL-6/gp130 signaling. J Clin Invest. 2011;121(9):3375-83.
- 131. Garbers C, Aparicio-Siegmund SRose-John S. The IL-6/gp130/STAT3 signaling axis: recent advances towards specific inhibition. Curr Opin Immunol. 2015;34:75-82.
- 132. Ishibashi T, Kimura H, Uchida T, Kariyone S, Friese PBurstein SA. Human interleukin 6 is a direct promoter of maturation of megakaryocytes in vitro. Proc Natl Acad Sci U S A. 1989;86(15):5953-57.
- 133. Chalaris A, Garbers C, Rabe B, Rose-John SScheller J. The soluble Interleukin 6 receptor: generation and role in inflammation and cancer. Eur J Cell Biol. 2011;90(6-7):484-94.
- 134. Rose-John S. IL-6 trans-signaling via the soluble IL-6 receptor: importance for the pro-inflammatory activities of IL-6. Int J Biol Sci. 2012;8(9):1237.
- 135. Haegeman G, Content J, Volckaert G, Derynck R, Tavernier JFiers W. Structural analysis of the sequence coding for an inducible 26-kDa protein in human fibroblasts. Eur J Biochem. 1986;159(3):625-32.
- 136. Kushner I, Jiang S-L, Zhang D, Lozanski GSamols D. Do post-transcriptional mechanisms participate in induction of C-reactive protein and serum amyloid A by IL-6 and IL-1? Ann N Y Acad Sci. 1995;762:102-07.
- 137. Ringheim GE, Szczepanik AM, Petko W, Burgher KL, Zu Zhu SChao CC. Enhancement of beta-amyloid precursor protein transcription and expression by the soluble interleukin-6 receptor/

interleukin-6 complex. Brain Res Mol Brain Res. 1998;55(1):35-44.

- 138. Peter C, BEHRMANN I, Gerhard M, LLER-NE-WEN FSGRAEVE L. Interleukin-6-type cytokine signalling through the gp130/Jak/STAT pathway1. Biochem J. 1998;334:297-314.
- 139. Ait-Ghezala G, Volmar CH, Frieling J, Paris D, Tweed M, Bakshi P, et al. CD40 promotion of amyloid beta production occurs via the NF-κB pathway. Eur J Neurosci. 2007;25(6):1685-95.
- 140. Forloni G, Mangiarotti F, Angeretti N, Lucca EDe Simoni MG. β-amyloid fragment potentiates IL-6 and TNF-α secretion by LPS in astrocytes but not in microglia. Cytokine. 1997;9(10):759-62.
- 141. Rothaug M, Becker-Pauly CRose-John S. The role of interleukin-6 signaling in nervous tissue. Biochim Biophys Acta Mol Cell Res. 2016;1863(6, Part A):1218-27.
- 142. Ben Haim L, Ceyzériat K, Carrillo-de Sauvage MA, Aubry F, Auregan G, Guillermier M, et al. The JAK/STAT3 pathway is a common inducer of astrocyte reactivity in Alzheimer's and Huntington's diseases. J Neurosci. 2015;35(6):2817-29.
- 143. Reichenbach N, Delekate A, Plescher M, Schmitt F, Krauss S, Blank N, et al. Inhibition of Stat3-mediated astrogliosis ameliorates pathology in an Alzheimer's disease model. EMBO Mol Med. 2019;11(2):e9665.
- 144. Gadient ROtten U. Expression of interleukin-6 (IL-6) and interleukin-6 receptor (IL-6R) mRNAs in rat brain during postnatal development. Brain Res. 1994;637(1-2):10-14.
- 145. Braida D, Sacerdote P, Panerai AE, Bianchi M, Aloisi AM, Iosuè S, et al. Cognitive function in young and adult IL (interleukin)-6 deficient mice. Behav Brain Res. 2004;153(2):423-29.
- 146. Toulmond S, Vige X, Fage DBenavides J. Local infusion of interleukin-6 attenuates the neurotoxic effects of NMDA on rat striatal cholinergic neurons. Neurosci Lett. 1992;144(1-2):49-52.
- 147. Lanzrein A-S, Johnston CM, Perry VH, Jobst KA, King EMSmith AD. Longitudinal study of inflammatory factors in serum, cerebrospinal fluid, and brain tissue in Alzheimer disease: interleukin-1beta, interleukin-6, interleukin-1 receptor antagonist, tumor necrosis factor-alpha, the soluble tumor necrosis factor receptors I and II, and alpha1-antichymotrypsin. Alzheimer Dis Assoc Disord. 1998;12(3):215-27.
- 148. Huberman M, Sredni B, Stern L, Kott EShalit F. IL-2 and IL-6 secretion in dementia: correlation with type and severity of disease. J Neurol Sci. 1995;130(2):161-4.
- 149. Cojocaru IM, Cojocaru M, Miu GSapira V. Study

of interleukin-6 production in Alzheimer's disease. Rom J Intern Med. 2011;49(1):55-8.

- 150. Kindy MS, Yu J, Guo JTZhu H. Apolipoprotein Serum Amyloid A in Alzheimer's Disease. J Alzheimers Dis. 1999;1(3):155-67.
- 151. Jang S, Jang WY, Choi M, Lee J, Kwon W, Yi J, et al. Serum amyloid A1 is involved in amyloid plaque aggregation and memory decline in amyloid beta abundant condition. Transgenic Res. 2019;28(5-6):499-508.
- 152. Quintanilla RA, Orellana DI, González-Billault CMaccioni RB. Interleukin-6 induces Alzheimer-type phosphorylation of tau protein by deregulating the cdk5/p35 pathway. Exp Cell Res. 2004;295(1):245-57.
- 153. Morales I, Farías GMaccioni RB. Neuroimmunomodulation in the pathogenesis of Alzheimer's disease. Neuroimmunomodulation. 2010;17(3):202-04.
- 154. Rothaug M, Becker-Pauly CRose-John S. The role of interleukin-6 signaling in nervous tissue. Biochim Biophys Acta Mol Cell Res. 2016;1863(6):1218-27.
- 155. Ivanov II, Zhou LLittman DR, editors. Transcriptional regulation of Th17 cell differentiation. Semin Immunol. 2007: Elsevier.
- 156. Morishima N, Mizoguchi I, Takeda K, Mizuguchi JYoshimoto T. TGF- β is necessary for induction of IL-23R and Th17 differentiation by IL-6 and IL-23. Biochem Biophys Res Commun. 2009;386(1):105-10.
- 157. Chakrabarty P, Jansen-West K, Beccard A, Ceballos-Diaz C, Levites Y, Verbeeck C, et al. Massive gliosis induced by interleukin-6 suppresses Aβ deposition in vivo: evidence against inflammation as a driving force for amyloid deposition. FASEB J. 2010;24(2):548-59.
- 158. Carswell EA, Old LJ, Kassel R, Green S, Fiore NWilliamson B. An endotoxin-induced serum factor that causes necrosis of tumors. Proc Natl Acad Sci U S A. 1975;72(9):3666-70.
- 159. Laws SM, Perneczky R, Wagenpfeil S, Müller U, Förstl H, Martins RN, et al. TNF polymorphisms in Alzheimer disease and functional implications on CSF beta-amyloid levels. Hum Mutat. 2005;26(1):29-35.
- 160. Luettig B, Decker TLohmann-Matthes M. Evidence for the existence of two forms of membrane tumor necrosis factor: an integral protein and a molecule attached to its receptor. J Immunol. 1989;143(12):4034-38.
- 161. Kriegler M, Perez C, DeFay K, Albert ILu S. A novel form of TNF/cachectin is a cell surface cyto-toxic transmembrane protein: ramifications for the

complex physiology of TNF. Cell. 1988;53(1):45-53.

- 162. McAlpine FE, Lee J-K, Harms AS, Ruhn KA, Blurton-Jones M, Hong J, et al. Inhibition of soluble TNF signaling in a mouse model of Alzheimer's disease prevents pre-plaque amyloid-associated neuropathology. Neurobiol Dis. 2009;34(1):163-77.
- 163. Alexopoulou L, Kranidioti K, Xanthoulea S, Denis M, Kotanidou A, Douni E, et al. Transmembrane TNF protects mutant mice against intracellular bacterial infections, chronic inflammation and autoimmunity. Eur J Immunol. 2006;36(10):2768-80.
- 164. Ware CF. Tumor Necrosis Factors. In: Bertino JR, editor. Encyclopedia of Cancer (Second Edition). New York: Academic Press; 2002. p. 475-89.
- 165. Kinouchi K, Brown G, Pasternak GDonner DB. Identification and characterization of receptors for tumor necrosis factor-alpha in the brain. Biochem Biophys Res Commun. 1991;181(3):1532-8.
- 166. Cheng B, Christakos SMattson MP. Tumor necrosis factors protect neurons against metabolic-excitotoxic insults and promote maintenance of calcium homeostasis. Neuron. 1994;12(1):139-53.
- 167. Ashkenazi A. Targeting death and decoy receptors of the tumour-necrosis factor superfamily. Nat Rev Cancer. 2002;2(6):420-30.
- 168. Rothe J, Lesslauer W, Lötscher H, Lang Y, Koebel P, Köntgen F, et al. Mice lacking the tumour necrosis factor receptor 1 are resistant to IMF-mediated toxicity but highly susceptible to infection by Listeria monocytogenes. Nature. 1993;364(6440):798-802.
- 169. van Horssen R, Ten Hagen TLEggermont AM. TNF-α in cancer treatment: molecular insights, antitumor effects, and clinical utility. Oncologist. 2006;11(4):397-408.
- 170. Yamamoto M, Kiyota T, Horiba M, Buescher JL, Walsh SM, Gendelman HE, et al. Interferon- γ and tumor necrosis factor- α regulate amyloid- β plaque deposition and β -secretase expression in Swedish mutant APP transgenic mice. Am J Pathol. 2007;170(2):680-92.
- 171. Blasko I, Marx F, Steiner E, Hartmann TGrubeck-Loebenstein B. TNF α plus IFN γ induce the production of Alzheimer β -amyloid peptides and decrease the secretion of APPs. FASEB J. 1999;13(1):63-68.
- 172. Osborn L, Kunkel SNabel GJ. Tumor necrosis factor alpha and interleukin 1 stimulate the human immunodeficiency virus enhancer by activation of the nuclear factor kappa B. Proc Natl Acad Sci U S A. 1989;86(7):2336-40.

- 173. Chang R, Yee K-LSumbria RK. Tumor necrosis factor α Inhibition for Alzheimer's Disease. J Cent Nerv Syst Dis. 2017;9:1179573517709278-78.
- 174. Hövelmeyer N, Hao Z, Kranidioti K, Kassiotis G, Buch T, Frommer F, et al. Apoptosis of oligodendrocytes via Fas and TNF-R1 is a key event in the induction of experimental autoimmune encephalomyelitis. J Immunol. 2005;175(9):5875-84.
- 175. Shen J, T-To SS, Schrieber LKing N. Early E-selectin, VCAM-1, ICAM-1, and late major histocompatibility complex antigen induction on human endothelial cells by flavivirus and comodulation of adhesion molecule expression by immune cytokines. J Virol. 1997;71(12):9323-32.
- 176. Fonseca SG, Romão PR, Figueiredo F, Morais RH, Lima HC, Ferreira SH, et al. TNF-alpha mediates the induction of nitric oxide synthase in macrophages but not in neutrophils in experimental cutaneous leishmaniasis. Eur J Immunol. 2003;33(8):2297-306.
- 177. Edwards MRobinson SR. TNF alpha affects the expression of GFAP and S100B: implications for Alzheimer's disease. J Neural Transm. 2006;113(11):1709-15.
- 178. Monje ML, Toda HPalmer TD. Inflammatory blockade restores adult hippocampal neurogenesis. Science. 2003;302(5651):1760-65.
- 179. Chen ZPalmer TD. Differential roles of TNFR1 and TNFR2 signaling in adult hippocampal neurogenesis. Brain Behav Immun. 2013;30:45-53.
- 180. Busquets S, Aranda X, Ribas-Carbo M, Azcon-Bieto J, López-Soriano FJArgilés JM. Tumour necrosis factor-alpha uncouples respiration in isolated rat mitochondria. Cytokine. 2003;22(1-2):1-4.
- 181. Oeckinghaus AGhosh S. The NF-κB family of transcription factors and its regulation. Cold Spring Harb Perspect Biol. 2009;1(4):a000034.
- 182. Sun S-C, Chang J-HJin J. Regulation of nuclear factor-κB in autoimmunity. Trends Immunol. 2013;34(6):282-89.
- 183. Beinke SLey SC. Functions of NF-κB1 and NF-κB2 in immune cell biology. Biochem J. 2004;382(2):393-409.
- 184. Almowallad S, Alqahtani LSMobashir M. NFkB in Signaling Patterns and Its Temporal Dynamics Encode/Decode Human Diseases. Life. 2022;12(12):2012.
- 185. Liu T, Zhang L, Joo DSun S-C. NF-κB signaling in inflammation. Signal Transduct Target Ther. 2017;2(1):17023.
- 186. Vallabhapurapu SKarin M. Regulation and function of NF-κB transcription factors in the immune system. Annu Rev Immunol. 2009;27:693-733.
- 187. Sun S-C. Non-canonical NF-кВ signaling path-

way. Cell Res. 2011;21(1):71-85.

- 188. Sun S-CLey SC. New insights into NF-κB regulation and function. Trends Immunol. 2008;29(10):469-78.
- 189. Karin MDelhase M, editors. The IκB kinase (IKK) and NF-κB: key elements of proinflammatory signalling. Semin Immunol. 2000: Elsevier.
- 190. Israël A. The IKK complex, a central regulator of NF-κB activation. Cold Spring Harb Perspect Biol. 2010;2(3):a000158.
- 191. Zhang HSun S-C. NF-κB in inflammation and renal diseases. Cell Biosci. 2015;5(1):1-12.
- 192. Hayden MSGhosh S. Shared principles in NF-κB signaling. Cell. 2008;132(3):344-62.
- 193. Lampl S, Janas MK, Donakonda S, Brugger M, Lohr K, Schneider A, et al. Reduced mitochondrial resilience enables non-canonical induction of apoptosis after TNF receptor signaling in virus-infected hepatocytes. J Hepatol. 2020;73(6):1347-59.
- 194. Choudhary S, Kalita M, Fang L, Patel KV, Tian B, Zhao Y, et al. Inducible tumor necrosis factor (TNF) receptor-associated factor-1 expression couples the canonical to the non-canonical NFκB pathway in TNF stimulation. J Biol Chem. 2013;288(20):14612-23.
- 195. Sun S-CLiu Z-G. A special issue on NF-κB signaling and function. Cell Res. 2011;21(1):1-2.
- 196. Sun SC. The noncanonical NF-κB pathway. Immunol Rev. 2012;246(1):125-40.
- 197. Yu H, Lin L, Zhang Z, Zhang HHu H. Targeting NF-κB pathway for the therapy of diseases: mechanism and clinical study. Signal Transduct Target Ther. 2020;5(1):1-23.
- 198. Morgan D, Garg M, Tergaonkar V, Tan SYSethi G. Pharmacological significance of the non-canonical NF-κB pathway in tumorigenesis. Biochim Biophys Acta Rev Cancer. 2020;1874(2):188449.
- 199. House CD, Grajales V, Ozaki M, Jordan E, Wubneh H, Kimble DC, et al. IKKε cooperates with either MEK or non-canonical NF-kB driving growth of triple-negative breast cancer cells in different contexts. BMC Cancer. 2018;18(1):1-13.
- 200. Pippione AC, Sainas S, Federico A, Lupino E, Piccinini M, Kubbutat M, et al. N-Acetyl-3-aminopyrazoles block the non-canonical NF-kB cascade by selectively inhibiting NIK. MedChemComm. 2018;9(6):963-68.
- 201. Bobardt M, Kuo J, Chatterji U, Chanda S, Little SJ, Wiedemann N, et al. The inhibitor apoptosis protein antagonist Debio 1143 Is an attractive HIV-1 latency reversal candidate. PLoS One. 2019;14(2):e0211746.
- 202. AlQasrawi D, Naser ENaser SA. Nicotine increases macrophage survival through α7nAChR/NF-κB

pathway in Mycobacterium avium paratuberculosis infection. Microorganisms. 2021;9(5):1086.

- 203. Jimi EKatagiri T. Critical roles of NF-κB signaling molecules in bone metabolism revealed by genetic mutations in osteopetrosis. Int J Mol Sci. 2022;23(14):7995.
- 204. Xiang H, Zhu F, Xu ZXiong J. Role of inflammasomes in kidney diseases via both canonical and non-canonical pathways. Front Cell Dev Biol. 2020;8:106.
- 205. Zhu J, Yamane HPaul WE. Differentiation of effector CD4 T cell populations (*). Annu Rev Immunol. 2010;28:445-89.
- 206. Oh HGhosh S. NF-κB: roles and regulation in different CD4(+) T-cell subsets. Immunol Rev. 2013;252(1):41-51.
- 207. Aronica MA, Mora AL, Mitchell DB, Finn PW, Johnson JE, Sheller JR, et al. Preferential role for NF-κB/Rel signaling in the type 1 but not type 2 T cell-dependent immune response in vivo. J Immunol. 1999;163(9):5116-24.
- 208. Li Y, Wang H, Zhou X, Xie X, Chen X, Jie Z, et al. Cell intrinsic role of NF-κB-inducing kinase in regulating T cell-mediated immune and autoimmune responses. Sci Rep. 2016;6(1):1-11.
- 209. Murray SE, Polesso F, Rowe AM, Basak S, Koguchi Y, Toren KG, et al. NF-κB–inducing kinase plays an essential T cell–intrinsic role in graft-versus-host disease and lethal autoimmunity in mice. J Clin Invest. 2011;121(12).
- 210. Rowe AM, Murray SE, Raué H-P, Koguchi Y, Slifka MKParker DC. A Cell-Intrinsic Requirement for NF-κB–Inducing Kinase in CD4 and CD8 T Cell Memory. J Immunol. 2013;191(7):3663-72.
- 211. Yu J, Wang Y, Yan F, Zhang P, Li H, Zhao H, et al. Noncanonical NF-κB activation mediates STAT3-stimulated IDO upregulation in myeloid-derived suppressor cells in breast cancer. J Immunol. 2014;193(5):2574-86.
- 212. Grilli M, Ribola M, Alberici A, Valerio A, Memo MSpano P. Amyloid precursor protein (APP) gene expression is controlled by a NFkB/Rel related protein. Alzheimer's and Parkinson's Diseases: Springer; 1995. p. 105-10.
- 213. Chen C-H, Zhou W, Liu S, Deng Y, Cai F, Tone M, et al. Increased NF-κB signalling up-regulates BACE1 expression and its therapeutic potential in Alzheimer's disease. Int J Neuropsychopharmacol. 2012;15(1):77-90.
- 214. Sadagurski M, Cady GMiller RA. Anti-aging drugs reduce hypothalamic inflammation in a sex-specific manner. Aging Cell. 2017;16(4):652-60.
- 215. Egaña-Gorroño L, López-Díez R, Yepuri G,

Ramirez LS, Reverdatto S, Gugger PF, et al. Receptor for advanced glycation end products (RAGE) and mechanisms and therapeutic opportunities in diabetes and cardiovascular disease: insights from human subjects and animal models. Front Cardiovasc Med. 2020;7:37.

- 216. Kong Y, Liu C, Zhou Y, Qi J, Zhang C, Sun B, et al. Progress of RAGE molecular imaging in Alzheimer's disease. Front Aging Neurosci. 2020;12:227.
- 217. Park S-S, Park H-S, Kim C-J, Baek S-S, Park S-Y, Anderson C, et al. Combined effects of aerobic exercise and 40-Hz light flicker exposure on early cognitive impairments in Alzheimer's disease of 3× Tg mice. J Appl Physiol. 2022;132(4):1054-68.
- 218. Wang G, Wang X, Zheng X, Sun S, Zhao J, Long Y, et al. Acidic oligosaccharide sugar chain combined with hyperbaric oxygen delays D-galactose-induced brain senescence in mice via attenuating oxidative stress and neuroinflammation. Neurosci Res. 2022.
- 219. Gonzalez-Reyes RERubiano MG. Astrocyte s RAGE: more than just a question of mood. Cent Nerv Syst Agents Med Chem. 2018;18(1):39-48.
- 220. Morris M, Evans D, Bienias J, Tangney C, Wilson R, Qu W, et al. Tao WY, Yu LJ, Jiang S, Cao X, Chen J, Bao XY, Li F, Xu Y, Zhu XL (2020) Neuroprotective effects of ZL006 in Aβ1-42-treated neuronal cells. Neural Regen Res. 2020;15(12):2296-2305. doi: 10.4103/1673-5374.285006. J Neurosci. 1999;19(13):5360-69.
- 221. Logsdon AF, Meabon JS, Cline MM, Bullock KM, Raskind MA, Peskind ER, et al. Blast exposure elicits blood-brain barrier disruption and repair mediated by tight junction integrity and nitric oxide dependent processes. Sci Rep. 2018;8(1):1-13.
- 222. Deane R, Du Yan S, Submamaryan RK, LaRue B, Jovanovic S, Hogg E, et al. RAGE mediates amyloid- β peptide transport across the blood-brain barrier and accumulation in brain. Nat Med. 2003;9(7):907-13.
- 223. Versele R, Sevin E, Gosselet F, Fenart LCandela P. TNF- α and IL-1 β Modulate Blood-Brain Barrier Permeability and Decrease Amyloid- β Peptide Efflux in a Human Blood-Brain Barrier Model. Int J Mol Sci. 2022;23(18):10235.
- 224. Swanson KV, Deng MTing JPY. The NLRP3 inflammasome: molecular activation and regulation to therapeutics. Nat Rev Immunol. 2019;19(8):477-89.
- 225. Guo H, Callaway JBTing JPY. Inflammasomes: mechanism of action, role in disease, and therapeutics. Nat Med. 2015;21(7):677-87.
- 226. Ketelut-Carneiro NFitzgerald KA. Inflammasomes. Curr Biol. 2020;30(12):R689-r94.

- 227. Li T, Lu L, Pember E, Li X, Zhang BZhu Z. New insights into neuroinflammation involved in pathogenic mechanism of Alzheimer's disease and its potential for therapeutic intervention. Cells. 2022;11(12):1925.
- 228. Yang Y, Wang H, Kouadir M, Song HShi F. Recent advances in the mechanisms of NLRP3 inflammasome activation and its inhibitors. Cell Death Dis. 2019;10(2):128.
- 229. Rahman T, Nagar A, Duffy EB, Okuda K, Silverman NHarton JA. NLRP3 sensing of diverse inflammatory stimuli requires distinct structural features. Front Immunol. 2020;11:1828.
- 230. Zheng D, Liwinski TElinav E. Inflammasome activation and regulation: toward a better understanding of complex mechanisms. Cell Discov. 2020;6(1):36.
- 231. Akbal A, Dernst A, Lovotti M, Mangan MSJ, McManus RMLatz E. How location and cellular signaling combine to activate the NLRP3 inflammasome. Cell Mol Immunol. 2022;19(11):1201-14.
- 232. Kelley N, Jeltema D, Duan YHe Y. The NLRP3 Inflammasome: An Overview of Mechanisms of Activation and Regulation. Int J Mol Sci. 2019;20(13).
- 233. Oroz J, Barrera-Vilarmau S, Alfonso C, Rivas Gde Alba E. ASC Pyrin Domain Self-associates and Binds NLRP3 Protein Using Equivalent Binding Interfaces. J Biol Chem. 2016;291(37):19487-501.
- 234. Dekker C, Mattes H, Wright M, Boettcher A, Hinniger A, Hughes N, et al. Crystal Structure of NLRP3 NACHT Domain With an Inhibitor Defines Mechanism of Inflammasome Inhibition. J Mol Biol. 2021;433(24):167309.
- 235. He Y, Zeng MY, Yang D, Motro BNúñez G. NEK7 is an essential mediator of NLRP3 activation downstream of potassium efflux. Nature. 2016;530(7590):354-57.
- 236. Chen JChen ZJ. PtdIns4P on dispersed trans-Golgi network mediates NLRP3 inflammasome activation. Nature. 2018;564(7734):71-76.
- 237. Bergsbaken T, Fink SLCookson BT. Pyroptosis: host cell death and inflammation. Nat Rev Microbiol. 2009;7(2):99-109.
- 238. Felderhoff-Mueser U, Schmidt OI, Oberholzer A, Bührer CStahel PF. IL-18: a key player in neuroinflammation and neurodegeneration? Trends Neurosci. 2005;28(9):487-93.
- 239. Lamkanfi MDixit VM. Mechanisms and functions of inflammasomes. Cell. 2014;157(5):1013-22.
- 240. Gustin A, Kirchmeyer M, Koncina E, Felten P, Losciuto S, Heurtaux T, et al. NLRP3 inflammasome is expressed and functional in mouse

brain microglia but not in astrocytes. PLoS One. 2015;10(6):e0130624.

- 241. Halle A, Hornung V, Petzold GC, Stewart CR, Monks BG, Reinheckel T, et al. The NALP3 inflammasome is involved in the innate immune response to amyloid- β . Nat Immunol. 2008;9(8):857-65.
- 242. Yap JKY, Pickard BS, Chan EWLGan SY. The role of neuronal NLRP1 inflammasome in Alzheimer's disease: bringing neurons into the neuroinflammation game. Mol Neurobiol. 2019;56(11):7741-53.
- 243. Tejera D, Mercan D, Sanchez-Caro JM, Hanan M, Greenberg D, Soreq H, et al. Systemic inflammation impairs microglial Aβ clearance through NLRP3 inflammasome. EMBO J. 2019;38(17):e101064.
- 244. Heneka MT, Kummer MP, Stutz A, Delekate A, Schwartz S, Vieira-Saecker A, et al. NLRP3 is activated in Alzheimer's disease and contributes to pathology in APP/PS1 mice. Nature. 2013;493(7434):674-8.
- 245. Bouchon A, Dietrich JColonna M. Cutting edge: inflammatory responses can be triggered by TREM-1, a novel receptor expressed on neutrophils and monocytes. J Immunol. 2000;164(10):4991-95.
- 246. Bouchon A, Hernández-Munain C, Cella MColonna M. A DAP12-mediated pathway regulates expression of CC chemokine receptor 7 and maturation of human dendritic cells. J Exp Med. 2001;194(8):1111-22.
- 247. Hu N, Tan M-S, Yu J-T, Sun L, Tan L, Wang Y-L, et al. Increased expression of TREM2 in peripheral blood of Alzheimer's disease patients. J Alzheimers Dis. 2014;38(3):497-501.
- 248. Colonna M. TREMs in the immune system and beyond. Nat Rev Immunol. 2003;3(6):445-53.
- 249. Forabosco P, Ramasamy A, Trabzuni D, Walker R, Smith C, Bras J, et al. Insights into TREM2 biology by network analysis of human brain gene expression data. Neurobiol Aging. 2013;34(12):2699-714.
- 250. Bhattacharjee S, Zhao Y, Dua P, Rogaev EILukiw WJ. microRNA-34a-mediated down-regulation of the microglial-enriched triggering receptor and phagocytosis-sensor TREM2 in age-related macular degeneration. PLoS One. 2016;11(3):e0150211.
- 251. Zheng H, Liu C-C, Atagi Y, Chen X-F, Jia L, Yang L, et al. Opposing roles of the triggering receptor expressed on myeloid cells 2 and triggering receptor expressed on myeloid cells-like transcript 2 in microglia activation. Neurobiol Aging. 2016;42:132-41.
- 252. Turnbull IR, Gilfillan S, Cella M, Aoshi T, Mill-

er M, Piccio L, et al. Cutting edge: TREM-2 attenuates macrophage activation. J Immunol. 2006;177(6):3520-24.

- 253. Gratuze M, Leyns CEGHoltzman DM. New insights into the role of TREM2 in Alzheimer's disease. Mol Neurodegener. 2018;13(1):66.
- 254. Sun M, Zhu M, Chen K, Nie X, Deng Q, Hazlett LD, et al. TREM-2 promotes host resistance against Pseudomonas aeruginosa infection by suppressing corneal inflammation via a PI3K/ Akt signaling pathway. Invest Ophthalmol Vis Sci. 2013;54(5):3451-62.
- 255. Sada K, Takano T, Yanagi SYamamura H. Structure and function of Syk protein-tyrosine kinase. J Biochem. 2001;130(2):177-86.
- 256. Mócsai A, Ruland JTybulewicz VL. The SYK tyrosine kinase: a crucial player in diverse biological functions. Nat Rev Immunol. 2010;10(6):387-402.
- 257. Takahashi K, Rochford CDNeumann H. Clearance of apoptotic neurons without inflammation by microglial triggering receptor expressed on myeloid cells-2. J Exp Med. 2005;201(4):647-57.
- 258. Lanier LL, Corliss BC, Wu J, Leong CPhillips JH. Immunoreceptor DAP12 bearing a tyrosine-based activation motif is involved in activating NK cells. Nature. 1998;391(6668):703-07.
- 259. Call ME, Wucherpfennig KWChou JJ. The structural basis for intramembrane assembly of an activating immunoreceptor complex. Nat Immunol. 2010;11(11):1023-29.
- 260. Saber M, Kokiko-Cochran O, Puntambekar SS, Lathia JDLamb BT. Triggering receptor expressed on myeloid cells 2 deficiency alters acute macrophage distribution and improves recovery after traumatic brain injury. J Neurotrauma. 2017;34(2):423-35.
- 261. Kawabori M, Kacimi R, Kauppinen T, Calosing C, Kim JY, Hsieh CL, et al. Triggering receptor expressed on myeloid cells 2 (TREM2) deficiency attenuates phagocytic activities of microglia and exacerbates ischemic damage in experimental stroke. J Neurosci. 2015;35(8):3384-96.
- 262. Takahashi K, Prinz M, Stagi M, Chechneva ONeumann H. TREM2-transduced myeloid precursors mediate nervous tissue debris clearance and facilitate recovery in an animal model of multiple sclerosis. PLoS Med. 2007;4(4):e124.
- 263. Atagi Y, Liu C-C, Painter MM, Chen X-F, Verbeeck C, Zheng H, et al. Apolipoprotein E is a ligand for triggering receptor expressed on myeloid cells 2 (TREM2). J Biol Chem. 2015;290(43):26043-50.
- 264. Li R, Zhang J, Wang Q, Cheng MLin B. TPM1 mediates inflammation downstream of TREM2 via the PKA/CREB signaling pathway. J Neuroin-

flammation. 2022;19(1):1-22.

- 265. Kobayashi M, Konishi H, Sayo A, Takai TKiyama H. TREM2/DAP12 signal elicits proinflammatory response in microglia and exacerbates neuropathic pain. J Neurosci. 2016;36(43):11138-50.
- 266. Gratuze M, Chen Y, Parhizkar S, Jain N, Strickland MR, Serrano JR, et al. Activated microglia mitigate Aβ-associated tau seeding and spreading. J Exp Med. 2021;218(8):e20210542.
- 267. Zhao Y, Wu X, Li X, Jiang L-L, Gui X, Liu Y, et al. TREM2 is a receptor for β-amyloid that mediates microglial function. Neuron. 2018;97(5):1023-31. e7.
- 268. Daws MR, Sullam PM, Niemi EC, Chen TT, Tchao NKSeaman WE. Pattern recognition by TREM-2: binding of anionic ligands. J Immunol. 2003;171(2):594-99.
- 269. Meilandt WJ, Ngu H, Gogineni A, Lalehzadeh G, Lee S-H, Srinivasan K, et al. Trem2 deletion reduces late-stage amyloid plaque accumulation, elevates the A β 42: A β 40 ratio, and exacerbates axonal dystrophy and dendritic spine loss in the PS2APP Alzheimer's mouse model. J Neurosci. 2020;40(9):1956-74.
- 270. Gratuze M, Leyns CE, Sauerbeck AD, St-Pierre M-K, Xiong M, Kim N, et al. Impact of TREM2 R47H variant on tau pathology-induced gliosis and neurodegeneration. J Clin Invest. 2020;130(9):4954-68.
- 271. Cosker K, Mallach A, Limaye J, Piers TM, Staddon J, Neame SJ, et al. Microglial signalling pathway deficits associated with the patient derived R47H TREM2 variants linked to AD indicate inability to activate inflammasome. Sci Rep. 2021;11(1):1-15.
- 272. Chen S, Peng J, Sherchan P, Ma Y, Xiang S, Yan F, et al. TREM2 activation attenuates neuroin-flammation and neuronal apoptosis via PI3K/Akt pathway after intracerebral hemorrhage in mice. J Neuroinflammation. 2020;17(1):1-16.
- 273. Decout A, Katz JD, Venkatraman SAblasser A. The cGAS–STING pathway as a therapeutic target in inflammatory diseases. Nat Rev Immunol. 2021;21(9):548-69.
- 274. Xie W, Lama L, Adura C, Tomita D, Glickman JF, Tuschl T, et al. Human cGAS catalytic domain has an additional DNA-binding interface that enhances enzymatic activity and liquid-phase condensation. Proc Natl Acad Sci U S A. 2019;116(24):11946-55.
- 275. Zhang X, Wu J, Du F, Xu H, Sun L, Chen Z, et al. The cytosolic DNA sensor cGAS forms an oligomeric complex with DNA and undergoes switchlike conformational changes in the activation loop. Cell Rep. 2014;6(3):421-30.
- 276. Gao P, Ascano M, Wu Y, Barchet W, Gaffney BL,

Zillinger T, et al. Cyclic [G (2', 5') pA (3', 5') p] is the metazoan second messenger produced by DNA-activated cyclic GMP-AMP synthase. Cell. 2013;153(5):1094-107.

- 277. Civril F, Deimling T, de Oliveira Mann CC, Ablasser A, Moldt M, Witte G, et al. Structural mechanism of cytosolic DNA sensing by cGAS. Nature. 2013;498(7454):332-37.
- 278. Ishikawa H, Ma ZBarber GN. STING regulates intracellular DNA-mediated, type I interferon-dependent innate immunity. Nature. 2009;461(7265):788-92.
- 279. Ishikawa HBarber GN. STING is an endoplasmic reticulum adaptor that facilitates innate immune signalling. Nature. 2008;455(7213):674-78.
- 280. Liu D, Wu H, Wang C, Li Y, Tian H, Siraj S, et al. STING directly activates autophagy to tune the innate immune response. Cell Death Differ. 2019;26(9):1735-49.
- 281. Prabakaran T, Bodda C, Krapp C, Zhang Bc, Christensen MH, Sun C, et al. Attenuation of c GAS-STING signaling is mediated by a p62/ SQSTM 1-dependent autophagy pathway activated by TBK1. EMBO J. 2018;37(8):e97858.
- 282. Gui X, Yang H, Li T, Tan X, Shi P, Li M, et al. Autophagy induction via STING trafficking is a primordial function of the cGAS pathway. Nature. 2019;567(7747):262-66.
- 283. Margolis SR, Wilson SCVance RE. Evolutionary origins of cGAS-STING signaling. Trends Immunol. 2017;38(10):733-43.
- 284. Nassour J, Radford R, Correia A, Fusté JM, Schoell B, Jauch A, et al. Autophagic cell death restricts chromosomal instability during replicative crisis. Nature. 2019;565(7741):659-63.
- 285. Gulen MF, Koch U, Haag SM, Schuler F, Apetoh L, Villunger A, et al. Signalling strength determines proapoptotic functions of STING. Nat Commun. 2017;8(1):1-10.
- 286. Glück S, Guey B, Gulen MF, Wolter K, Kang T-W, Schmacke NA, et al. Innate immune sensing of cytosolic chromatin fragments through cGAS promotes senescence. Nat Cell Biol. 2017;19(9):1061-70.
- 287. Zierhut C, Yamaguchi N, Paredes M, Luo J-D, Carroll TFunabiki H. The cytoplasmic DNA sensor cGAS promotes mitotic cell death. Cell. 2019;178(2):302-15.e23.
- 288. Petrasek J, Iracheta-Vellve A, Csak T, Satishchandran A, Kodys K, Kurt-Jones EA, et al. STING-IRF3 pathway links endoplasmic reticulum stress with hepatocyte apoptosis in early alcoholic liver disease. Proc Natl Acad Sci U S A. 2013;110(41):16544-49.

- 289. Liu SGuan W. STING signaling promotes apoptosis, necrosis, and cell death: an overview and update. Mediators Inflamm. 2018;2018.
- 290. White MJ, McArthur K, Metcalf D, Lane RM, Cambier JC, Herold MJ, et al. Apoptotic caspases suppress mtDNA-induced STING-mediated type I IFN production. Cell. 2014;159(7):1549-62.
- 291. Morgan MJKim Y-S. Roles of RIPK3 in necroptosis, cell signaling, and disease. Exp Mol Med. 2022:1-10.
- 292. Brault M, Olsen TM, Martinez J, Stetson DBOberst A. Intracellular nucleic acid sensing triggers necroptosis through synergistic type I IFN and TNF signaling. J Immunol. 2018;200(8):2748-56.
- 293. DeRoo E, Zhou TLiu B. The role of RIPK1 and RIPK3 in cardiovascular disease. Int J Mol Sci. 2020;21(21):8174.
- 294. Hui CW, Zhang YHerrup K. Non-Neuronal Cells Are Required to Mediate the Effects of Neuroinflammation: Results from a Neuron-Enriched Culture System. PLoS One. 2016;11(1):e0147134.
- 295. Chen K, Lai C, Su Y, Bao WD, Yang LN, Xu P-P, et al. cGAS-STING-mediated IFN-I Response in Host Defense and Neuroinflammatory Diseases. Curr Neuropharmacol. 2022;20(2):362-71.
- 296. Tan P-H, Ji J, Hsing C-H, Tan RJi R-R. Emerging Roles of Type-I Interferons in Neuroinflammation, Neurological Diseases, and Long-Haul COVID. Int J Mol Sci. 2022;23(22):14394.
- 297. Wang W, Hu D, Wu C, Feng Y, Li A, Liu W, et al. STING promotes NLRP3 localization in ER and facilitates NLRP3 deubiquitination to activate the inflammasome upon HSV-1 infection. PLoS Pathog. 2020;16(3):e1008335.
- 298. Jin M, Shiwaku H, Tanaka H, Obita T, Ohuchi S, Yoshioka Y, et al. Tau activates microglia via the PQBP1-cGAS-STING pathway to promote brain inflammation. Nat Commun. 2021;12(1):1-22.
- 299. Zhao J, Xu C, Cao H, Zhang L, Wang XChen S. Identification of target genes in neuroinflammation and neurodegeneration after traumatic brain injury in rats. PeerJ. 2019;7:e8324.
- 300. Boyd RJ, Avramopoulos D, Jantzie LLMcCallion AS. Neuroinflammation represents a common theme amongst genetic and environmental risk factors for Alzheimer and Parkinson diseases. J Neuroinflammation. 2022;19(1):223.
- 301. Fraser PE, Yang D-S, Yu G, Lévesque L, Nishimura M, Arawaka S, et al. Presenilin structure, function and role in Alzheimer disease. Biochim Biophys Acta. 2000;1502(1):1-15.
- 302. Sherrington R, Rogaev E, Liang Y, Rogaeva E, Levesque G, Ikeda M, et al. Cloning of a gene

bearing missense mutations in early-onset familial Alzheimer's disease. Nature. 1995;375:754-60.

- 303. Kelleher RJ, 3rdShen J. Presenilin-1 mutations and Alzheimer's disease. Proc Natl Acad Sci U S A. 2017;114(4):629-31.
- 304. Restrepo LJ, DePew AT, Moese ER, Tymanskyj SR, Parisi MJ, Aimino MA, et al. γ-secretase promotes Drosophila postsynaptic development through the cleavage of a Wnt receptor. Dev Cell. 2022.
- 305. Monacelli F, Martella L, Parodi MN, Odetti P, Fanelli FTabaton M. Frontal variant of Alzheimer's disease: a report of a novel PSEN1 mutation. J Alzheimers Dis. 2019;70(1):11-15.
- 306. Duff K, Eckman C, Zehr C, Yu X, Prada C-M, Perez-Tur J, et al. Increased amyloid- β 42 (43) in brains of mice expressing mutant presenilin 1. Nature. 1996;383(6602):710-13.
- 307. Hardy JSelkoe DJ. The amyloid hypothesis of Alzheimer's disease: progress and problems on the road to therapeutics. Science. 2002;297(5580):353-56.
- 308. Selkoe DJHardy J. The amyloid hypothesis of Alzheimer's disease at 25 years. EMBO Mol Med. 2016;8(6):595-608.
- 309. Shen JKelleher III RJ. The presenilin hypothesis of Alzheimer's disease: evidence for a loss-offunction pathogenic mechanism. Proc Natl Acad Sci U S A. 2007;104(2):403-09.
- 310. Wines-Samuelson M, Schulte EC, Smith MJ, Aoki C, Liu X, Kelleher III RJ, et al. Characterization of age-dependent and progressive cortical neuronal degeneration in presenilin conditional mutant mice. PLoS One. 2010;5(4):e10195.
- 311. Watanabe H, Xia D, Kanekiyo T, Kelleher RJShen J. Familial frontotemporal dementia-associated presenilin-1 c. 548G> T mutation causes decreased mRNA expression and reduced presenilin function in knock-in mice. J Neurosci. 2012;32(15):5085-96.
- 312. Saura CA, Choi S-Y, Beglopoulos V, Malkani S, Zhang D, Rao BS, et al. Loss of presenilin function causes impairments of memory and synaptic plasticity followed by age-dependent neurodegeneration. Neuron. 2004;42(1):23-36.
- 313. Xia D, Watanabe H, Wu B, Lee SH, Li Y, Tsvetkov E, et al. Presenilin-1 knockin mice reveal loss-offunction mechanism for familial Alzheimer's disease. Neuron. 2015;85(5):967-81.
- 314. Levy-Lahad E, Wasco W, Poorkaj P, Romano DM, Oshima J, Pettingell WH, et al. Candidate gene for the chromosome 1 familial Alzheimer's disease locus. Science. 1995;269(5226):973-77.
- 315. Cai Y, An SSKim S. Mutations in presenilin 2 and

its implications in Alzheimer's disease and other dementia-associated disorders. Clin Interv Aging. 2015;10:1163-72.

- 316. Rademakers R, Cruts MVan Broeckhoven C. Genetics of early-onset Alzheimer dementia. ScientificWorldJournal. 2003;3:497-519.
- 317. Annaert WG, Levesque L, Craessaerts K, Dierinck I, Snellings G, Westaway D, et al. Presenilin 1 controls γ-secretase processing of amyloid precursor protein in pre-Golgi compartments of hippocampal neurons. J Cell Biol. 1999;147(2):277-94.
- 318. Leissring MA, Yamasaki TR, Wasco W, Buxbaum JD, Parker ILaFerla FM. Calsenilin reverses presenilin-mediated enhancement of calcium signaling. Proc Natl Acad Sci U S A. 2000;97(15):8590-93.
- 319. Leissring MA, LaFerla FM, Callamaras NParker I. Subcellular mechanisms of presenilin-mediated enhancement of calcium signaling. Neurobiol Dis. 2001;8(3):469-78.
- 320. Citron M, Westaway D, Xia W, Carlson G, Diehl T, Levesque G, et al. Mutant presenilins of Alzheimer's disease increase production of 42-residue amyloid β -protein in both transfected cells and transgenic mice. Nat Med. 1997;3(1):67-72.
- 321. Goldgaber D, Lerman MI, McBride OW, Saffiotti UGajdusek DC. Characterization and chromosomal localization of a cDNA encoding brain amyloid of Alzheimer's disease. Science. 1987;235(4791):877-80.
- 322. Müller UCZheng H. Physiological functions of APP family proteins. Cold Spring Harb Perspect Med. 2012;2(2):a006288.
- 323. Tharp WGSarkar IN. Origins of amyloid-β. BMC Genomics. 2013;14(1):290.
- 324. Dahms SO, Hoefgen S, Roeser D, Schlott B, Gührs KHThan ME. Structure and biochemical analysis of the heparin-induced E1 dimer of the amyloid precursor protein. Proc Natl Acad Sci U S A. 2010;107(12):5381-6.
- 325. Guénette S, Strecker PKins S. APP protein family signaling at the synapse: insights from intracellular APP-binding proteins. Front Mol Neurosci. 2017;10:87.
- 326. Nikolaev A, McLaughlin T, O'Leary DDTessier-Lavigne M. APP binds DR6 to trigger axon pruning and neuron death via distinct caspases. Nature. 2009;457(7232):981-9.
- 327. Zheng HKoo EH. Biology and pathophysiology of the amyloid precursor protein. Mol Neurodegener. 2011;6(1):27.
- 328. Haass C, Hung AY, Selkoe DJTeplow DB. Mutations associated with a locus for familial Alzheimer's disease result in alternative processing of amyloid beta-protein precursor. J Biol Chem.

1994;269(26):17741-48.

- 329. Suzuki N, Cheung TT, Cai X-D, Odaka A, Otvos Jr L, Eckman C, et al. An increased percentage of long amyloid β protein secreted by familial amyloid β protein precursor (βAPP717) mutants. Science. 1994;264(5163):1336-40.
- 330. Tcw JGoate AM. Genetics of β-Amyloid Precursor Protein in Alzheimer's Disease. Cold Spring Harb Perspect Med. 2017;7(6).
- 331. Busciglio J, Gabuzda DH, Matsudaira PYankner BA. Generation of beta-amyloid in the secretory pathway in neuronal and nonneuronal cells. Proc Natl Acad Sci U S A. 1993;90(5):2092-96.
- 332. Mullan M, Crawford F, Axelman K, Houlden H, Lilius L, Winblad B, et al. A pathogenic mutation for probable Alzheimer's disease in the APP gene at the N-terminus of β -amyloid. Nat Genet. 1992;1(5):345-47.
- 333. Levy E, Carman MD, Fernandez-Madrid IJ, Power MD, Lieberburg I, van Duinen SG, et al. Mutation of the Alzheimer's disease amyloid gene in hereditary cerebral hemorrhage, Dutch type. Science. 1990;248(4959):1124-26.
- 334. Nilsberth C, Westlind-Danielsson A, Eckman CB, Condron MM, Axelman K, Forsell C, et al. The'Arctic'APP mutation (E693G) causes Alzheimer's disease by enhanced Aβ protofibril formation. Nat Neurosci. 2001;4(9):887-93.
- 335. Hardy J. Amyloid, the presenilins and Alzheimer's disease. Trends Neurosci. 1997;20(4):154-59.
- 336. Giaccone G, Morbin M, Moda F, Botta M, Mazzoleni G, Uggetti A, et al. Neuropathology of the recessive A673V APP mutation: Alzheimer disease with distinctive features. Acta Neuropathol. 2010;120:803-12.
- 337. Maloney JA, Bainbridge T, Gustafson A, Zhang S, Kyauk R, Steiner P, et al. Molecular mechanisms of Alzheimer disease protection by the A673T allele of amyloid precursor protein. J Biol Chem. 2014;289(45):30990-1000.
- 338. Jonsson T, Atwal JK, Steinberg S, Snaedal J, Jonsson PV, Bjornsson S, et al. A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. Nature. 2012;488(7409):96-99.
- 339. Parhizkar SHoltzman DM, editors. APOE mediated neuroinflammation and neurodegeneration in Alzheimer's disease. Semin Immunol. 2022;59:101594.
- 340. Olaisen B, Teisberg PGedde-Dahl T. The locus for apolipoprotein E (apoE) is linked to the complement component C3 (C3) locus on chromosome 19 in man. Hum Genet. 1982;62(3):233-36.
- 341. Kim J, Castellano JM, Jiang H, Basak JM, Parsadanian M, Pham V, et al. Overexpression of low-den-

sity lipoprotein receptor in the brain markedly inhibits amyloid deposition and increases extracellular A β clearance. Neuron. 2009;64(5):632-44.

- 342. Di Battista AM, Heinsinger NMRebeck GW. Alzheimer's Disease Genetic Risk Factor APOE-ε4 Also Affects Normal Brain Function. Curr Alzheimer Res. 2016;13(11):1200-07.
- 343. Liu C-C, Zhao N, Fu Y, Wang N, Linares C, Tsai C-W, et al. ApoE4 accelerates early seeding of amyloid pathology. Neuron. 2017;96(5):1024-32.e3.
- 344. Verghese PB, Castellano JM, Garai K, Wang Y, Jiang H, Shah A, et al. ApoE influences amyloid- β (A β) clearance despite minimal apoE/A β association in physiological conditions. Proc Natl Acad Sci U S A. 2013;110(19):E1807-E16.
- 345. LaDu MJ, Pederson TM, Frail DE, Reardon CA, Getz GSFalduto MT. Purification of apolipoprotein E attenuates isoform-specific binding to β -amyloid. J Biol Chem. 1995;270(16):9039-42.
- 346. Wildsmith KR, Holley M, Savage JC, Skerrett RLandreth GE. Evidence for impaired amyloid β clearance in Alzheimer's disease. Alzheimers Res Ther. 2013;5(4):33.
- 347. Litvinchuk A, Huynh T-PV, Shi Y, Jackson RJ, Finn MB, Manis M, et al. Apolipoprotein E4 Reduction with Antisense Oligonucleotides Decreases Neurodegeneration in a Tauopathy Model. Ann Neurol. 2021;89(5):952-66.
- 348. Shi Y, Andhey PS, Ising C, Wang K, Snipes LL, Boyer K, et al. Overexpressing low-density lipoprotein receptor reduces tau-associated neurodegeneration in relation to apoE-linked mechanisms. Neuron. 2021;109(15):2413-26.e7.
- 349. Parhizkar SHoltzman DM. APOE mediated neuroinflammation and neurodegeneration in Alzheimer's disease. Semin Immunol. 2022;59:101594.
- 350. Shi Y, Yamada K, Liddelow SA, Smith ST, Zhao L, Luo W, et al. ApoE4 markedly exacerbates tau-mediated neurodegeneration in a mouse model of tauopathy. Nature. 2017;549(7673):523-27.
- 351. Prasad HRao R. Amyloid clearance defect in ApoE4 astrocytes is reversed by epigenetic correction of endosomal pH. Proc Natl Acad Sci U S A. 2018;115(28):E6640-E49.
- 352. Keren-Shaul H, Spinrad A, Weiner A, Matcovitch-Natan O, Dvir-Szternfeld R, Ulland TK, et al. A unique microglia type associated with restricting development of Alzheimer's disease. Cell. 2017;169(7):1276-90.e17.
- 353. Lin Y-T, Seo J, Gao F, Feldman HM, Wen H-L, Penney J, et al. APOE4 causes widespread molecular and cellular alterations associated with Alzheimer's disease phenotypes in human iPSC-derived brain cell types. Neuron. 2018;98(6):1141-54.e7.

- 354. Muzio L, Viotti AMartino G. Microglia in neuroinflammation and neurodegeneration: from understanding to therapy. Front Neurosci. 2021;15.
- 355. Murugaiyan G, Beynon V, Mittal A, Joller NWeiner HL. Silencing microRNA-155 ameliorates experimental autoimmune encephalomyelitis. J Immunol. 2011;187(5):2213-21.
- 356. Lv R, Du L, Zhou F, Yuan X, Liu XZhang L. Rosmarinic Acid Alleviates Inflammation, Apoptosis, and Oxidative Stress through Regulating miR-155-5p in a Mice Model of Parkinson's Disease. ACS Chem Neurosci. 2020;11(20):3259-66.
- 357. Wen Y, Zhang X, Dong L, Zhao J, Zhang CZhu C. Acetylbritannilactone Modulates MicroRNA-155-Mediated Inflammatory Response in Ischemic Cerebral Tissues. Mol Med. 2015;21(1):197-209.
- 358. Wang WX, Prajapati P, Vekaria HJ, Spry M, Cloud AL, Sullivan PG, et al. Temporal changes in inflammatory mitochondria-enriched microR-NAs following traumatic brain injury and effects of miR-146a nanoparticle delivery. Neural Regen Res. 2021;16(3):514-22.
- 359. Raikwar SP, Thangavel R, Ahmed ME, Selvakumar GP, Kempuraj D, Wu K, et al. Real-Time Noninvasive Bioluminescence, Ultrasound and Photoacoustic Imaging in NFκB-RE-Luc Transgenic Mice Reveal Glia Maturation Factor-Mediated Immediate and Sustained Spatio-Temporal Activation of NFκB Signaling Post-Traumatic Brain Injury in a Gender-Specific Manner. Cell Mol Neurobiol. 2021;41(8):1687-706.
- 360. Paez-Colasante X, Figueroa-Romero C, Sakowski SA, Goutman SAFeldman EL. Amyotrophic lateral sclerosis: mechanisms and therapeutics in the epigenomic era. Nat Rev Neurol. 2015;11(5):266-79.
- 361. Guedes JR, Custódia CM, Silva RJ, de Almeida LP, Pedroso de Lima MCCardoso AL. Early miR-155 upregulation contributes to neuroinflammation in Alzheimer's disease triple transgenic mouse model. Hum Mol Genet. 2014;23(23):6286-301.
- 362. Pagano A, Castelnuovo M, Tortelli F, Ferrari R, Dieci GCancedda R. New small nuclear RNA gene-like transcriptional units as sources of regulatory transcripts. PLoS Genet. 2007;3(2):e1.
- 363. Li D, Zhang J, Li X, Chen Y, Yu FLiu Q. Insights into lncRNAs in Alzheimer's disease mechanisms. RNA Biol. 2021;18(7):1037-47.
- 364. Gavazzo P, Vella S, Marchetti C, Nizzari M, Cancedda RPagano A. Acquisition of neuron-like electrophysiological properties in neuroblastoma cells by controlled expression of NDM29 ncRNA. J Neurochem. 2011;119(5):989-1001.

- 365. Castelnuovo M, Massone S, Tasso R, Fiorino G, Gatti M, Robello M, et al. An Alu-like RNA promotes cell differentiation and reduces malignancy of human neuroblastoma cells. FASEB J. 2010;24(10):4033-46.
- 366. Massone S, Ciarlo E, Vella S, Nizzari M, Florio T, Russo C, et al. NDM29, a RNA polymerase III-dependent non coding RNA, promotes amyloidogenic processing of APP and amyloid β secretion. Biochim Biophys Acta. 2012;1823(7):1170-7.
- 367. Hansen TB, Wiklund ED, Bramsen JB, Villadsen SB, Statham AL, Clark SJ, et al. miRNA-dependent gene silencing involving Ago2-mediated cleavage of a circular antisense RNA. EMBO J. 2011;30(21):4414-22.
- 368. Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, Damgaard CK, et al. Natural RNA circles function as efficient microRNA sponges. Nature. 2013;495(7441):384-8.
- 369. Chen J, Yang J, Fei X, Wang XWang K. CircRNA ciRS-7: a Novel Oncogene in Multiple Cancers. Int J Biol Sci. 2021;17(1):379-89.
- 370. Zhao Y, Alexandrov PN, Jaber VLukiw WJ. Deficiency in the Ubiquitin Conjugating Enzyme UBE2A in Alzheimer's Disease (AD) is Linked to Deficits in a Natural Circular miRNA-7 Sponge (circRNA; ciRS-7). Genes. 2016;7(12):116.
- 371. Davis DH, Muniz Terrera G, Keage H, Rahkonen T, Oinas M, Matthews FE, et al. Delirium is a strong risk factor for dementia in the oldest-old: a population-based cohort study. Brain. 2012;135(9):2809-16.
- 372. Jamieson GA, Maitland NJ, Wilcock GK, Yates CMItzhaki RF. Herpes simplex virus type 1 DNA is present in specific regions of brain from aged people with and without senile dementia of the Alzheimer type. J Pathol. 1992;167(4):365-68.
- 373. Honjo K, van Reekum RVerhoeff NP. Alzheimer's disease and infection: do infectious agents contribute to progression of Alzheimer's disease? Alzheimers Dement. 2009;5(4):348-60.
- 374. Piacentini R, Civitelli L, Ripoli C, Marcocci ME, De Chiara G, Garaci E, et al. HSV-1 promotes Ca2+-mediated APP phosphorylation and Aβ accumulation in rat cortical neurons. Neurobiol Aging. 2011;32(12):2323.e13-23.e26.
- 375. Itzhaki RF, Lin W-R, Shang D, Wilcock GK, Faragher BJamieson GA. Herpes simplex virus type 1 in brain and risk of Alzheimer's disease. Lancet. 1997;349(9047):241-44.
- 376. Carbone I, Lazzarotto T, Ianni M, Porcellini E, Forti P, Masliah E, et al. Herpes virus in Alzheimer's disease: relation to progression of the disease. Neurobiol Aging. 2014;35(1):122-29.

- 377. Bu XL, Yao XQ, Jiao SS, Zeng F, Liu YH, Xiang Y, et al. A study on the association between infectious burden and Alzheimer's disease. Eur J Neurol. 2015;22(12):1519-25.
- 378. Tohidpour A, Morgun AV, Boitsova EB, Malinovskaya NA, Martynova GP, Khilazheva ED, et al. Neuroinflammation and infection: molecular mechanisms associated with dysfunction of neurovascular unit. Front Cell Infect Microbiol. 2017;7:276.
- 379. Alisky JM. The coming problem of HIV-associated Alzheimer's disease. Med Hypotheses. 2007;69(5):1140-43.
- 380. Li L, Mao S, Wang J, Ding XZen JY. Viral infection and neurological disorders—potential role of extracellular nucleotides in neuroinflammation. ExRNA. 2019;1(1):26.
- 381. Sun X-W, Liu C-MTeng Z-Q. Commentary: multiscale analysis of independent Alzheimer's cohorts finds disruption of molecular, genetic, and clinical networks by human herpesvirus. Front Mol Neurosci. 2018;11:340.
- 382. Allnutt MA, Johnson K, Bennett DA, Connor SM, Troncoso JC, Pletnikova O, et al. Human herpesvirus 6 detection in Alzheimer's disease cases and controls across multiple cohorts. Neuron. 2020;105(6):1027-35.e2.
- 383. Maheshwari PEslick GD. Bacterial infection and Alzheimer's disease: a meta-analysis. J Alzheimers Dis. 2015;43(3):957-66.
- 384. McManus RMHeneka MT. Role of neuroinflammation in neurodegeneration: new insights. Alzheimers Res Ther. 2017;9(1):14.
- 385. Gérard HC, Dreses-Werringloer U, Wildt KS, Deka S, Oszust C, Balin BJ, et al. Chlamydophila (Chlamydia) pneumoniae in the Alzheimer's brain. FEMS Immunol Med Microbiol. 2006;48(3):355-66.
- 386. Roubaud-Baudron C, Krolak-Salmon P, Quadrio I, Mégraud FSalles N. Impact of chronic Helicobacter pylori infection on Alzheimer's disease: preliminary results. Neurobiol Aging. 2012;33(5):1009.e11-09.e19.
- 387. Beydoun MA, Beydoun HA, Shroff MR, Kitner-Triolo MHZonderman AB. Helicobacter pylori seropositivity and cognitive performance among US adults: evidence from a large national survey. Psychosom Med. 2013;75(5):486.
- 388. Kamer AR, Pirraglia E, Tsui W, Rusinek H, Vallabhajosula S, Mosconi L, et al. Periodontal disease associates with higher brain amyloid load in normal elderly. Neurobiol Aging. 2015;36(2):627-33.
- 389. Riviere GR, Riviere KHSmith KS. Molecular and immunological evidence of oral Treponema

in the human brain and their association with Alzheimer's disease. Oral Microbiol Immunol. 2002;17(2):113-8.

- 390. Miklossy J, Kis A, Radenovic A, Miller L, Forro L, Martins R, et al. Beta-amyloid deposition and Alzheimer's type changes induced by Borrelia spirochetes. Neurobiol Aging. 2006;27(2):228-36.
- 391. Kanagasingam S, Chukkapalli SS, Welbury RSinghrao SK. Porphyromonas gingivalis is a Strong Risk Factor for Alzheimer's Disease. J Alzheimers Dis Rep. 2020;4(1):501-11.
- 392. Dominy SS, Lynch C, Ermini F, Benedyk M, Marczyk A, Konradi A, et al. Porphyromonas gingivalis in Alzheimer's disease brains: Evidence for disease causation and treatment with small-molecule inhibitors. Sci Adv. 2019;5(1).
- 393. Confettura AD, Cuboni E, Ammar MR, Jia S, Gomes GM, Yuanxiang P, et al. Neddylation-dependent protein degradation is a nexus between synaptic insulin resistance, neuroinflammation and Alzheimer's disease. Transl Neurodegener. 2022;11(1):2.
- 394. Van Dyken PLacoste B. Impact of metabolic syndrome on neuroinflammation and the blood-brain barrier. Front Neurosci. 2018:930.
- 395. Gomez G, Beason-Held LL, Bilgel M, An Y, Wong DF, Studenski S, et al. Metabolic syndrome and amyloid accumulation in the aging brain. J Alzheimers Dis. 2018;65(2):629-39.
- 396. Calsolaro VEdison P. Neuroinflammation in Alzheimer's disease: current evidence and future directions. Alzheimers Dement. 2016;12(6):719-32.
- 397. Talbot K, Wang H-Y, Kazi H, Han L-Y, Bakshi KP, Stucky A, et al. Demonstrated brain insulin resistance in Alzheimer's disease patients is associated with IGF-1 resistance, IRS-1 dysregulation, and cognitive decline. J Clin Invest. 2012;122(4):1316-38.
- 398. Mullins RJ, Mustapic M, Goetzl EJKapogiannis D. Exosomal biomarkers of brain insulin resistance associated with regional atrophy in Alzheimer's disease. Hum Brain Mapp. 2017;38(4):1933-40.
- 399. Moloney AM, Griffin RJ, Timmons S, O'Connor R, Ravid RO'Neill C. Defects in IGF-1 receptor, insulin receptor and IRS-1/2 in Alzheimer's disease indicate possible resistance to IGF-1 and insulin signalling. Neurobiol Aging. 2010;31(2):224-43.
- 400. Lee HJ, Seo HI, Cha HY, Yang YJ, Kwon SHYang SJ. Diabetes and Alzheimer's disease: mechanisms and nutritional aspects. Clin Nutr Res. 2018;7(4):229.
- 401. Neth BJCraft S. Insulin Resistance and Alzheimer's Disease: Bioenergetic Linkages. Front Aging Neurosci. 2017;9:345.

- 402. Mehla J, Chauhan BCChauhan NB. Experimental induction of type 2 diabetes in aging-accelerated mice triggered Alzheimer-like pathology and memory deficits. J Alzheimers Dis. 2014;39(1):145-62.
- 403. Vandal M, White PJ, Tremblay C, St-Amour I, Chevrier G, Emond V, et al. Insulin reverses the high-fat diet-induced increase in brain A β and improves memory in an animal model of Alzheimer disease. Diabetes. 2014;63(12):4291-301.
- 404. Currais A, Prior M, Lo D, Jolivalt C, Schubert DMaher P. Diabetes exacerbates amyloid and neurovascular pathology in aging-accelerated mice. Aging Cell. 2012;11(6):1017-26.
- 405. Cao D, Lu H, Lewis TLLi L. Intake of sucrose-sweetened water induces insulin resistance and exacerbates memory deficits and amyloidosis in a transgenic mouse model of Alzheimer disease. J Biol Chem. 2007;282(50):36275-82.
- 406. Kim I, Lee J, Hong HJ, Jung ES, Ku YH, Jeong IK, et al. A relationship between Alzheimer's disease and type 2 diabetes mellitus through the measurement of serum amyloid-beta autoantibodies. J Alzheimers Dis. 2010;19(4):1371-6.
- 407. Yang Y, Wu Y, Zhang SSong W. High glucose promotes Aβ production by inhibiting APP degradation. PLoS One. 2013;8(7):e69824.
- 408. Rorbach-Dolata APiwowar A. Neurometabolic evidence supporting the hypothesis of increased incidence of type 3 diabetes mellitus in the 21st century. Biomed Res Int. 2019;2019.
- 409. Rojas M, Chávez-Castillo M, Bautista J, Ortega Á, Nava M, Salazar J, et al. Alzheimer's disease and type 2 diabetes mellitus: Pathophysiologic and pharmacotherapeutics links. World J Diabetes. 2021;12(6):745.
- 410. Monteiro RAzevedo I. Chronic inflammation in obesity and the metabolic syndrome. Mediators Inflamm. 2010;2010.
- 411. Hotamisligil GS. Inflammation and metabolic disorders. Nature. 2006;444(7121):860-67.
- 412. Tilg HMoschen AR. Adipocytokines: mediators linking adipose tissue, inflammation and immunity. Nat Rev Immunol. 2006;6(10):772-83.
- 413. Zarkesh-Esfahani H, Pockley G, Metcalfe RA, Bidlingmaier M, Wu Z, Ajami A, et al. Highdose leptin activates human leukocytes via receptor expression on monocytes. J Immunol. 2001;167(8):4593-99.
- 414. Fantuzzi G. Adipose tissue, adipokines, and inflammation. J Allergy Clin Immunol. 2005;115(5):911-19.
- 415. Coppack SW. Pro-inflammatory cytokines and adipose tissue. Proc Nutr Soc. 2001;60(3):349-56.

- 416. Pinteaux E, Inoue W, Schmidt L, Molina-Holgado F, Rothwell NJLuheshi GN. Leptin induces interleukin-1β release from rat microglial cells through a caspase 1 independent mechanism. J Neurochem. 2007;102(3):826-33.
- 417. Lam QLK, Zheng B-J, Jin D-Y, Cao XLu L. Leptin induces CD40 expression through the activation of Akt in murine dendritic cells. Elsevier. 2007.
- 418. Haczeyni F, Bell-Anderson KSFarrell GC. Causes and mechanisms of adipocyte enlargement and adipose expansion. Obes Rev. 2018;19(3):406-20.
- 419. Patel HPatel V. Inflammation and metabolic syndrome-an overview. Curr Res Nutr Food Sci J. 2015;3(3):263-68.
- 420. Aguilar DFernandez ML. Hypercholesterolemia induces adipose dysfunction in conditions of obesity and nonobesity. Adv Nutr. 2014;5(5):497-502.
- 421. Lin Y, Rajala MW, Berger JP, Moller DE, Barzilai NScherer PE. Hyperglycemia-induced production of acute phase reactants in adipose tissue. J Biol Chem. 2001;276(45):42077-83.
- 422. Mondal A, Saha P, Bose D, Chatterjee S, Seth RK, Xiao S, et al. Environmental Microcystin exposure in underlying NAFLD-induced exacerbation of neuroinflammation, blood-brain barrier dysfunction, and neurodegeneration are NLRP3 and S100B dependent. Toxicology. 2021;461:152901.
- 423. Picone P, Di Carlo MNuzzo D. Obesity and Alzheimer's disease: Molecular bases. Eur J Neurosci. 2020;52(8):3944-50.
- 424. McLarnon JG. A Leaky Blood–Brain Barrier to Fibrinogen Contributes to Oxidative Damage in Alzheimer's Disease. Antioxidants. 2021;11(1):102.
- 425. Shin Y, Choi SH, Kim E, Bylykbashi E, Kim JA, Chung S, et al. Blood-brain barrier dysfunction in a 3D in vitro model of Alzheimer's disease. Adv Sci (Weinh). 2019;6(20):1900962.
- 426. Wang C, Shou Y, Pan J, Du Y, Liu CWang H. The relationship between cholesterol level and Alzheimer's disease-associated APP proteolysis/Aβ metabolism. Nutr Neurosci. 2019;22(7):453-63.
- 427. Roca-Agujetas V, de Dios C, Abadin XColell A. Upregulation of brain cholesterol levels inhibits mitophagy in Alzheimer disease. Autophagy. 2021;17(6):1555-57.
- 428. Pagliassotti MJ, Kim PY, Estrada AL, Stewart CMGentile CL. Endoplasmic reticulum stress in obesity and obesity-related disorders: An expanded view. Metabolism. 2016;65(9):1238-46.
- 429. Salminen A, Kaarniranta KKauppinen A. ER stress activates immunosuppressive network: implications for aging and Alzheimer's disease. J Mol Med. 2020;98(5):633-50.
- 430. Uddin MS, Tewari D, Sharma G, Kabir MT,

Barreto GE, Bin-Jumah MN, et al. Molecular Mechanisms of ER Stress and UPR in the Pathogenesis of Alzheimer's Disease. Mol Neurobiol. 2020;57:2902-19.

- 431. Kern PA, Saghizadeh M, Ong JM, Bosch RJ, Deem RSimsolo RB. The expression of tumor necrosis factor in human adipose tissue. Regulation by obesity, weight loss, and relationship to lipoprotein lipase. J Clin Invest. 1995;95(5):2111-19.
- 432. Tumminia A, Vinciguerra F, Parisi MFrittitta L. Type 2 Diabetes Mellitus and Alzheimer's Disease: Role of Insulin Signalling and Therapeutic Implications. Int J Mol Sci. 2018;19(11):3306.
- 433. Valerio A, Cardile A, Cozzi V, Bracale R, Tedesco L, Pisconti A, et al. TNF-α downregulates eNOS expression and mitochondrial biogenesis in fat and muscle of obese rodents. J Clin Invest. 2006;116(10):2791-98.
- 434. Bairamian D, Sha S, Rolhion N, Sokol H, Dorothée G, Lemere CA, et al. Microbiota in neuroinflammation and synaptic dysfunction: a focus on Alzheimer's disease. Mol Neurodegener. 2022;17(1):19.
- 435. Lavelle ASokol H. Gut microbiota-derived metabolites as key actors in inflammatory bowel disease. Nat Rev Gastroenterol Hepatol. 2020;17(4):223-37.
- 436. McCarville JL, Chen GY, Cuevas VD, Troha KAyres JS. Microbiota metabolites in health and disease. Annu Rev Immunol. 2020;38:147-70.
- 437. Okin DMedzhitov R. Evolution of inflammatory diseases. Curr Biol. 2012;22(17):R733-R40.
- 438. Vainchtein IDMolofsky AV. Astrocytes and microglia: in sickness and in health. Trends Neurosci. 2020;43(3):144-54.
- 439. Harach T, Marungruang N, Duthilleul N, Cheatham V, Mc Coy K, Frisoni G, et al. Reduction of Abeta amyloid pathology in APPPS1 transgenic mice in the absence of gut microbiota. Sci Rep. 2017;7(1):1-15.
- 440. Chen Y, Fang L, Chen S, Zhou H, Fan Y, Lin L, et al. Gut microbiome alterations precede cerebral amyloidosis and microglial pathology in a mouse model of Alzheimer's disease. Biomed Res Int. 2020;2020.
- 441. Kowalski KMulak A. Brain-gut-microbiota axis in Alzheimer's disease. J Neurogastroenterol Motil. 2019;25(1):48.
- 442. Vogt NM, Kerby RL, Dill-McFarland KA, Harding SJ, Merluzzi AP, Johnson SC, et al. Gut microbiome alterations in Alzheimer's disease. Sci Rep. 2017;7(1):13537.
- 443. Du Y, Wu H-T, Qin X-Y, Cao C, Liu Y, Cao Z-Z, et al. Postmortem brain, cerebrospinal fluid, and

blood neurotrophic factor levels in Alzheimer's disease: a systematic review and meta-analysis. J Mol Neurosci. 2018;65(3):289-300.

- 444. Gareau MG, Wine E, Rodrigues DM, Cho JH, Whary MT, Philpott DJ, et al. Bacterial infection causes stress-induced memory dysfunction in mice. Gut. 2011;60(3):307-17.
- 445. Liang S, Wang T, Hu X, Luo J, Li W, Wu X, et al. Administration of Lactobacillus helveticus NS8 improves behavioral, cognitive, and biochemical aberrations caused by chronic restraint stress. Neuroscience. 2015;310:561-77.
- 446. Quan NBanks WA. Brain-immune communication pathways. Brain Behav Immun. 2007;21(6):727-35.
- 447. Huston JM. The vagus nerve and the inflammatory reflex: wandering on a new treatment paradigm for systemic inflammation and sepsis. Surg Infect (Larchmt). 2012;13(4):187-93.
- 448. Cummings J, Lee G, Zhong K, Fonseca JTaghva K. Alzheimer's disease drug development pipeline: 2021. Alzheimers Dement (N Y). 2021;7(1):e12179.
- 449. Yang L, Liu Y, Wang Y, Li JLiu N. Azeliragon ameliorates Alzheimer's disease via the Janus tyrosine kinase and signal transducer and activator of transcription signaling pathway. Clinics (Sao Paulo). 2021;76.
- 450. Sabbagh MNDecourt B. COR388 (atuzaginstat): an investigational gingipain inhibitor for the treatment of Alzheimer disease. Expert Opin Investig Drugs. 2022;31(10):987-93.
- 451. Padda ISParmar M. Aducanumab. StatPearls [Internet]. StatPearls Publishing; 2021.
- 452. Hampel H, Williams C, Etcheto A, Goodsaid F, Parmentier F, Sallantin J, et al. A precision medicine framework using artificial intelligence for the identification and confirmation of genomic biomarkers of response to an Alzheimer's disease therapy: analysis of the blarcamesine (ANAV-EX2-73) Phase 2a clinical study. Alzheimers Dement (N Y). 2020;6(1):e12013.
- 453. Herrmann N, Ruthirakuhan M, Gallagher D, Verhoeff NPL, Kiss A, Black SE, et al. Randomized placebo-controlled trial of nabilone for agitation in Alzheimer's disease. Am J Geriatr Psychiatry. 2019;27(11):1161-73.
- 454. Hoang K, Watt H, Golemme M, Perry RJ, Ritchie C, Wilson D, et al. Noradrenergic Add-on Therapy with Extended-Release Guanfacine in Alzheimer's Disease (NorAD): Study protocol for a randomised clinical trial and COVID-19 amendments. Trials. 2022;23(1):1-13.
- 455. Liao Z, Cheng L, Li X, Zhang M, Wang SHuo R.

Meta-analysis of ginkgo biloba preparation for the treatment of Alzheimer's disease. Clin Neuropharmacol. 2020;43(4):93-99.

- 456. Grossberg GT, Kohegyi E, Mergel V, Josiassen MK, Meulien D, Hobart M, et al. Efficacy and safety of brexpiprazole for the treatment of agitation in Alzheimer's dementia: two 12-week, randomized, double-blind, placebo-controlled trials. Am J Geriatr Psychiatry. 2020;28(4):383-400.
- 457. Khoury R, Marx C, Mirgati S, Velury D, Chakkamparambil BGrossberg GT. AVP-786 as a promising treatment option for Alzheimer's Disease including agitation. Expert Opin Pharmacother. 2021;22(7):783-95.
- 458. Vandenberghe R, Riviere ME, Caputo A, Sovago J, Maguire RP, Farlow M, et al. Active Aβ immunotherapy CAD106 in Alzheimer's disease: A phase 2b study. Alzheimers Dement (N Y). 2017;3(1):10-22.
- 459. Wiessner C, Wiederhold K-H, Tissot AC, Frey P, Danner S, Jacobson LH, et al. The second-generation active A β immunotherapy CAD106 reduces amyloid accumulation in APP transgenic mice while minimizing potential side effects. J Neurosci. 2011;31(25):9323-31.
- 460. Winblad B, Andreasen N, Minthon L, Floesser A, Imbert G, Dumortier T, et al. Safety, tolerability, and antibody response of active Aβ immunotherapy with CAD106 in patients with Alzheimer's disease: randomised, double-blind, placebo-controlled, first-in-human study. Lancet Neurol. 2012;11(7):597-604.
- 461. Giancane G, Minoia F, Davì S, Bracciolini G, Consolaro ARavelli A. IL-1 inhibition in systemic juvenile idiopathic arthritis. Front Pharmacol. 2016;7:467.
- 462. Qi Y, Klyubin I, Cuello ACRowan MJ. NLRP3-dependent synaptic plasticity deficit in an Alzheimer's disease amyloidosis model in vivo. Neurobiol Dis. 2018;114:24-30.
- 463. Kitazawa M, Cheng D, Tsukamoto MR, Koike MA, Wes PD, Vasilevko V, et al. Blocking IL-1 signaling rescues cognition, attenuates tau pathology, and restores neuronal β-catenin pathway function in an Alzheimer's disease model. J Immunol. 2011;187(12):6539-49.
- 464. Jiang H, He H, Chen Y, Huang W, Cheng J, Ye J, et al. Identification of a selective and direct NLRP3 inhibitor to treat inflammatory disorders. J Exp Med. 2017;214(11):3219-38.
- 465. Fulp J, He L, Toldo S, Jiang Y, Boice A, Guo C, et al. Structural insights of benzenesulfonamide analogues as NLRP3 inflammasome inhibitors: design, synthesis, and biological characterization.

J Med Chem. 2018;61(12):5412-23.

- 466. Munoz L, Ralay Ranaivo H, Roy SM, Hu W, Craft JM, McNamara LK, et al. A novel p38 alpha MAPK inhibitor suppresses brain proinflammatory cytokine up-regulation and attenuates synaptic dysfunction and behavioral deficits in an Alzheimer's disease mouse model. J Neuroinflammation. 2007;4:21.
- 467. Yanagisawa D, Ibrahim NF, Taguchi H, Morikawa S, Hirao K, Shirai N, et al. Curcumin derivative with the substitution at C-4 position, but not curcumin, is effective against amyloid pathology in APP/PS1 mice. Neurobiol Aging. 2015;36(1):201-10.
- 468. Zhang S, Gao L, Liu X, Lu T, Xie CJia J. Resveratrol Attenuates Microglial Activation via SIRT1-SOCS1 Pathway. Evid Based Complement Alternat Med. 2017;2017:8791832.
- 469. Hou Y, Wei Y, Lautrup S, Yang B, Wang Y, Cordonnier S, et al. NAD(+) supplementation reduces neuroinflammation and cell senescence in a transgenic mouse model of Alzheimer's disease via cGAS-STING. Proc Natl Acad Sci U S A. 2021;118(37).
- 470. Cai Y, Liu J, Wang B, Sun MYang H. Microglia in the Neuroinflammatory Pathogenesis of Alzheimer's Disease and Related Therapeutic Targets. Front Immunol. 2022;13:856376.
- 471. Lee YS, Gupta DP, Park SH, Yang HJSong GJ. Anti-Inflammatory Effects of Dimethyl Fumarate in Microglia via an Autophagy Dependent Pathway. Front Pharmacol. 2021;12:612981.
- 472. Tang RH, Qi RQLiu HY. Interleukin-4 affects microglial autophagic flux. Neural Regen Res. 2019;14(9):1594-602.
- 473. Frias DP, Gomes RLN, Yoshizaki K, Carvalho-Oliveira R, Matsuda M, Junqueira MS, et al. Nrf2 positively regulates autophagy antioxidant response in human bronchial epithelial cells exposed to diesel exhaust particles. Sci Rep. 2020;10(1):3704.
- 474. Zhang W, Feng CJiang H. Novel target for treating Alzheimer's Diseases: Crosstalk between the Nrf2 pathway and autophagy. Ageing Res Rev. 2021;65:101207.
- 475. Zhou X, Chu X, Xin D, Li T, Bai X, Qiu J, et al. L-Cysteine-Derived H(2)S Promotes Microglia M2 Polarization via Activation of the AMPK Pathway in Hypoxia-Ischemic Neonatal Mice. Front Mol Neurosci. 2019;12:58.
- 476. Li C, Zhang C, Zhou H, Feng Y, Tang F, Hoi MPM, et al. Inhibitory Effects of Betulinic Acid on LPS-Induced Neuroinflammation Involve M2 Microglial Polarization via CaMKKβ-Dependent AMPK Activation. Front Mol Neurosci.

2018;11:98.

- 477. Ji J, Xue TF, Guo XD, Yang J, Guo RB, Wang J, et al. Antagonizing peroxisome proliferator-activated receptor γ facilitates M1-to-M2 shift of microglia by enhancing autophagy via the LKB1-AMPK signaling pathway. Aging Cell. 2018;17(4):e12774.
- 478. Cui W, Sun C, Ma Y, Wang S, Wang XZhang Y. Inhibition of TLR4 Induces M2 Microglial Polarization and Provides Neuroprotection via the NLRP3 Inflammasome in Alzheimer's Disease. Front Neurosci. 2020;14:444.