\* Corresponding author:

Mohammad Balfaki, M.Sc.,

Shahid Beheshti University,

m.balfaki@alumni.sbu.ac.ir

Tel: +98 9359430494

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Tehran, Iran

E-mail:



# Revolutionary Regeneration Therapy Utilizing Dental Stem Cells and State-of-the-Art Nanotechnology Devices to Heal Injured Teeth and Tissues

Mohammad Balfaki 1\* and Erfan Salimi 2

1. Shahid Beheshti University, Tehran, Iran 2. Tib University of Baku, Baku, Azerbaijan

### Abstract

Regenerative medicine is a field of pharmacy and medicine that focuses on stem cells and other methods such as nanoscience and biotechnology to stimulate the body's natural regenerative processes and repairing damaged tissues and organs to improve function and reduce pain. In this review article, focus is on Dental Stem Cells (DSC) and other cells regeneration in human body. The appropriateness of tissue-engineered therapies relying on the multipotent regenerative abilities of DSC is accompanied by significant challenges, as growth factors and epigenetic components are crucial for preserving their multipotency while being susceptible to a range of natural and environmental factors. Current evidence highlights the positive outcomes associated with select regenerative therapies; nevertheless, to provide further support, additional data must be gathered through standardized therapies and further studies. Organoids (3D cell culture) and nano scaffolds are also being explored as potential tools for regenerative therapies. Understanding the mechanisms that determine the behavior of these cells and how they interact will enable future generation therapies. Demonstrating promise, cell therapy is an alternative approach within regenerative medicine. Developmental factors like extracellular vesicle production are thought to mediate the regenerative response through paracrine effects in cell therapy, which is widely recognized.

**Keywords:** Cell culture, Immunomodulation, Organoids, Regenerative medicine, Scaffolds, Signal pathways, Stem cell research

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## Introduction

Regenerative therapies represent a promising approach in medical biology that utilizes gene tools and technologies to possibly cure a wide range of human diseases, maladies, and disorders remotely. Additionally, we have several regenerative treatments, such as stem cell therapy 1 which uses stem cells or their products to encourage the healing mechanism of damaged, diseased, or dysfunctional tissue. Tissue engineering demonstrates numerous areas to care for cancer, storage cord blood and cell banking, gastrointestinal and gynecology, dermal, dentistry, urology, musculoskeletal, orthopedics, spine, cardiology, vascular and neurology, and the subsequent failure in organ transplantation. It also uses cells rather than donor organs <sup>2</sup>. The next method for regeneration therapies in biomedicine is gene therapy 3. By replacing diseased genes with viable duplicates and deactivating non-functioning harmful genes, the strategy involved in modifying a

person's genes for treating or curing diseases aligns seamlessly with the breakthroughs achieved in gene therapy. The next new approach in this field is Platelet-Rich Plasma (PRP) Therapy <sup>4</sup>. The therapy is based on a designed process that modifies an individual's genes to provide treatment or remedy for diseases that can synergize with gene therapy advancements through the substitution of a disease-causing gene with its healthy counterpart and the neutralization of an ineffective disease-causing gene <sup>5</sup>.

Another therapy to regenerate the human body is Extracorporeal (outside the body) Shockwave Therapy (EWST) <sup>6</sup>. A generation which makes use of surprise waves to deal with persistent painful situations of the musculoskeletal system. The process was spearheaded by the creation of EWST technology alongside the integration of fundamental physical-theoretical knowledge. An assortment of application devices such as

electrohydraulic, piezoelectric and both flat and cylindrical electromagnetic focusing mechanisms were developed along with radial ballistic devices. Early tests were conducted to evaluate their performance in various applications <sup>7</sup>. The last method of regeneration refer is Regenerative Rehabilitation <sup>8</sup>. The end objective of regenerative medicine is to create novel and efficient approaches that enable tissue regeneration and repair, thus promoting functional restoration <sup>9</sup>.

In this review, stem cell therapy for regeneration for dental disorders was studied with focus on some techniques and applications to repair and restore lost functions in tissues. The implementation of dental stem cell-based regenerative treatments involves the utilization of Dental Stem Cells (DCSs), 3D cell culture, biomaterials, signaling pathways, epigenetic direction, and immunomodulation. These components work together to advance the regeneration of dental tissues and move forward in dental care. Advanced exploration and advancement in these domains are crucial to unlocking the complete potential of DSCs in regenerative healthcare applications.

#### Stem cell research in dental tissues

DSCs are pluripotent cells that have the ability to continuously divide and produce pups that differentiate into several other cell types or tissue types. Adult solid teeth's dental pulp tissue has been isolated. Additionally, deciduous teeth are another promising area of research, offering exciting possibilities for regenerative dentistry applications <sup>10</sup>.

## Here are a few keys on DSCs

The confinement of DSCs includes extraction from different tissues within the teeth and encompassing structures:

1. The ability of DSCs like Gingival Mesenchymal Stem Cells (GMSCs), Dental Pulp Stem Cells (DPSCs), and Periodontal Ligament Stem Cells (PDLSCs) means

- they can clone and regenerate themselves which is extraordinary. This implies a method by which the stem cells isolate themselves to produce more stem cells, lifelong reproduction of the stem cell pool as well as multipotent isolation capacity and flexibility <sup>11</sup>. The most recent study demonstrated the characteristics of these cells, including their ability to exhibit immunomodulatory and stem cell-like pathways and to promote tissue regeneration or recovery <sup>11,12</sup>.
- 2. Dental stem cells (DSCs) have demonstrated significant potential in treating a variety of dental conditions, including periodontitis, dental caries, and Temporomandibular Joint (TMJ) osteoarthritis <sup>13,14</sup>. They have to be investigated for potential in tissue engineering and regenerative medication application <sup>15</sup>.
- 3. Regeneration of damaged dental tissues: Stem cell-based approaches are being explored to recover harmed or lost tissues within the verbal and maxillofacial locale, counting teeth <sup>16</sup>. As figure 1 indicates the innovative organ germination strategy is utilized in regenerative treatment to effectively transplant bioengineered dental embryos comprising epithelial and mesenchymal cells, resulting in the complete recovery of functional teeth and bio-roots *in vivo*. Analysts are investigating procedures utilizing DSCs, biomaterials, and particular 3D culture conditions to imitate the characteristic specialty of DSCs. This approach points to addressing embedded disappointment and giving elective regenerative pharmaceutical alternatives for dental wellbeing <sup>16,17</sup>.
- 4. Clinical applications: Clinical applications of DSCs have been utilized for treating dental ailments and surgical sites. They have the capability to recover harmed or lost tissues within the verbal and maxillofacial locale, counting teeth <sup>18</sup>. Clinical trials are currently underway to evaluate the suitability and safety of DSC therapy for various dental applications <sup>16</sup>. Their special

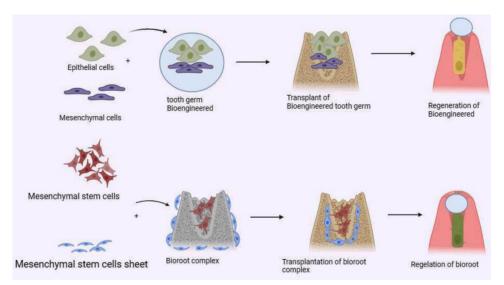


Figure 1. Utilizing an innovative organ germination strategy, regenerative treatment facilitates the transplantation of bioengineered dental embryos composed of epithelial and mesenchymal cells to restore fully functional teeth and bio-roots *in vivo*.

characteristics, restorative potential, and capacity to recover harmed tissues make them important for the treatment of dental maladies and tissue designing applications. Progressing inquiries about clinical trials are growing our understanding of DSCs and some of their applications in dental care.

Also, for regenerative treatments without any immune system reaction from the body's cells, DSCs can serve as a hopeful replacement for bone marrow-derived stem cells for inflammation. They have regenerative potential and can separate into different cell sorts included in tooth and tissue regeneration. DSCs are effectively available and helpful together, making them an appealing alternative for regenerative capacity dentistry applications. Continuous inquiry about points to optimize their regenerative capacity and create inventive procedures for dental tissue designing and regeneration process.

## Distinctive types of dental stem cells and their properties

Recent research has identified and characterized several distinct types of dental stem cells. These cells exhibit unique characteristics and hold enormous promise for the field of regenerative medicine and the mechanism of cell metabolism. It is fascinating to observe the extraordinary capabilities of DSCs like DPSCs, Stem cell Human Deciduous (SHEDs), PDLSCs, Stem Cells collected from Apical Papilla (SCAPs), and Dental Follicle Progenitor Cells (DFPCs) within the domain of regenerative medicine as they continue to be identified and characterized. These cells possess invaluable characteristics like high stemness properties, regenerative activities, and a remarkable potential for differentiating into various cell types; thus, making them exceedingly favorable for advancements in tissue engineering and regenerative medicine in vivo 19,20. Based on recent search results, there are several potential advantages associated with utilizing DSCs in regenerative medicine including non-invasiveness, high stemness properties, regenerative activities and differentiation potential <sup>20,21</sup>.

# Here are some types of DSCs and their unique characteristics listed below

Dental pulp stem cells (DPSCs): DPSCs are a promising form of DSCs that can be isolated from the tooth's pulp considering they have characteristics similar to those of stem cells, including the ability to self-renew and divide into several cell types. In addition, they demonstrate immunomodulatory properties in an *in vivo* situation <sup>22</sup>. DPSCs have a few properties and characteristics that make them profitable in different investigations and applications. Here are a few key properties of DPSCs based on the high proliferation rate <sup>23</sup>, low immunogenicity <sup>24</sup>, multipotency <sup>24</sup>, differentiation potential <sup>25</sup>, neuro-immunomodulatory properties <sup>26</sup>, and angiogenic potential <sup>27</sup>. It's critical to note that these properties may change depending on the particular thinking about or inquiry about the setting.

DPSCs proceed to be a dynamic range of inquiries about, and investigations are underway to delve into the extensive possibilities and uses in regenerative medicine and tissue engineering.

Stem cell human deciduous (SHED): SHED, occupies a special position because they can be extracted from the pulp of milk teeth. These cells exhibit clonogenicity, the ability to self-renew and the ability to differentiate into many cell types. In addition, SHED exhibits both stem cell-like properties and immunomodulatory properties <sup>28</sup>. SHED has a high proliferative rate and can separate into osteoblasts, neural cells or adipocytes and odontoblasts <sup>19</sup>. Different development variables, calculations related to vascular endothelial development and figures reflecting platelet-derived development and fundamental fibroblast development calculations, had been investigated as natural signals to control stem cells' destiny for pulp regeneration <sup>29,30</sup>.

The term (PDLSCs) designates the stem cells found in the periodontal ligament: PDLSCs are a type of dental stem cell that can be disconnected from the periodontal ligament <sup>31</sup>. The perivascular space of the periodontium serves as a habitat for this stem cell, which has the potential to transplant and rejuvenate the Periodontal tendons (PDL). By using stem cells, growth factors or the extracellular skeleton, it becomes possible to effectively repair damaged periodontal tissue through tissue regeneration <sup>31,32</sup>. Conjointly they can recognize different cell sorts included inside the course of action and back of the periodontium, which joins the gums, periodontal tendon, cementum, and alveolar bone 31,33. PDLSCs have been compared to other sorts of stem cells collected from Apical Papilla (SCAP) and deciduous teeth (SHED) and represent prime examples of diverse stem cell sources to evaluate their periodontal properties and separation potential 32. The impact of various factors, including Bone Morphogenetic Proteins (BMPs), commonly known as bone morphogens, has been explored <sup>34</sup>.

Specifically, dental follicle stem cells (DFSCs): DSCs can detach from tooth follicles. These particular cells exhibit clonogenicity and self-renewal abilities while possessing multipotent separable abilities. In addition, they exhibit both stem cell-like properties and immunomodulatory properties <sup>35,36</sup>. The presence of pluripotency markers like OCT-4, SOX-2, and NANOG in DFPCs demonstrates their self-renewal capacity and multipotent potential <sup>37</sup>. DFSCs can attain osteogenic, adaptogenic, chondrogenic, neural, and cardiomyocytic separation in a particularly initiated environment <sup>38</sup>. DFSCs outperform other DSCs by having a much higher rate of cell expansion, greater ability to form colonies, as well as a more primitive and much better anti-inflammatory effect <sup>35</sup>.

Table 1 details the classification and research progress of Dental Stem Cells specifically in relation to periapical lesions and their regeneration <sup>45,46</sup>.

Table 1. Dental stem cells, also known as Dental Pulp Stem Cells (DPSCs), are a promising source of regenerative medicine due to their unique properties
and ability to differentiate into various cell types

Cell type	Induction/targeted differentiation	Application area	Approval status	Nanotechnology device commercialization status
Dental Pulp Stem Cells (DPSCs) (39)	Osteogenic, chondrogenic	Dental tissue engineering	Research stage	Limited
Stem Cells from Human Exfoliated Deciduous Teeth (SHED) (40)	Osteogenic, neurogenic	Regenerative dentistry	Research stage	Limited
Periodontal Ligament Stem Cells (PDLSCs) (41)	Osteogenic	Periodontal regeneration	Research stage	Limited
Dental Follicle Progenitor Cells (DFPCs) (42)	Odontogenic	Tooth regeneration	Research stage	Limited
Stem Cells from Apical Papilla (SCAP) (43)	Odontogenic	Dental pulp regeneration	Research stage	Limited
Gingival Mesenchymal Stem Cells (GMSCs) (44)	Osteogenic, adipogenic	Periodontal tissue regeneration	Research stage	Limited
Human Natal Dental Pulp Stem Cells (NDP-SCs) (43)	Osteogenic	Dental tissue repair	Research stage	Limited

There are several FDA approvals that involve the use of DSCs or stem cell-derived technologies. Some examples of such notable uses follow:

AlloDerm Regenerative Tissue Matrix is an acellular dermal matrix derived from donated human skin, used in various dental and surgical procedures. It finds its application in periodontal surgery and grafting for its inherent property of tissue regeneration. Though not a stem cell product, it is mixed with DSCs to enhance the healing and regeneration process in oral applications.

Another example of FDA clinical approval is Osteogeny which is a bone grafting material that can be used in conjunction with stem cells. It finds its application in dental implant procedures for the purpose of encouraging bone regeneration. If combined with DPSCs, the osteogenic potentiality is enhanced, hence it can serve as a tool in regenerative dentistry.

The Platelet-Rich Plasma (PRP) and Platelet-Rich Fibrin (PRF) are obtained from the patient's blood and contain growth factors responsible for promoting healing. These products have been commonly used in the enhancement of a regenerative process in periodontal and implant surgeries in conjunction with DSC therapies. Though they themselves are not FDA-approved as direct stem cell therapies, their application with stem cells is immense in clinical practice <sup>47-49</sup>.

# Stem cells originating from the dental apical papilla, also known as (SCAPs)

Mesenchymal stem cells, so-called SCAPs, obtained from the apical papillae of young teeth have great potential for regenerative endodontic applications <sup>31,50</sup>. The results of a recent comparative study examining DPSC and SCAP indicate that the two cell types have different mesenchymal stem cell markers and may have different genetic backgrounds. Although no significant differences in cell viability, growth potential, or motility were observed between DPSCs and SCAP, discrepancies in the emitted profile were observed <sup>50</sup>

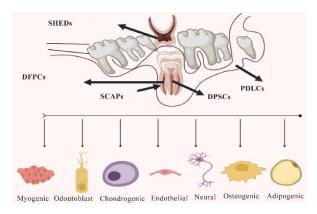


Figure 2. Dental Stem Cells (DSCs) particularly Dental Pulp Stem Cells (DPSCs) and Stem Cells from Apical Papilla (SCAPs) exhibit significant differentiation potentials that are being explored for regenerative medicine applications, especially in dental tissue engineering.

and the remarkable differentiating abilities of DSCs as shown in figure 2. A recent study comparing DPSCs and SCAPs discovered that both cell types expressed mesenchymal stem cell markers and exhibited the potential to differentiate into different lineages. Although no significant differences were observed in terms of cell functionality, proliferation, or motility between DPSC and SCAP, variations were noted in the secretion profile.

Through analysis, it was shown that the fusion of Mineral Trioxide Aggregate (MTA), Enamel Matrix Derivative (EMD), and Low-level laser therapy (LLLT) complemented by dentine generated notable consequences. EMD and LLLT seem to lead to a critical increment in SCAP practicality compared with another treatment bunch <sup>51</sup>. This study aimed to investigate the properties of human Dental Pulp Stem Cells (hDPSC)-derived, Jagged1-treated dECM (Extracellular Matrix) and its intrinsic effect on odonto-/osteogenic SCAP

separation, leading to the discovery of dECM-like fibrous organizational structures. The effects of decellularized ECM is the next important finding from recent research on SCAPs. There was a reduction in the components of calcium and phosphate following deECM <sup>52</sup>. Another investigation demonstrates the SCAP features' cytokine profiling. The key goal of this study is to examine the effects of oral bacteria and their supernatants on the cytokine profile of SCAP, with particular attention to the potential consequences for osteogenic and immunogenic characteristics <sup>53</sup>.

The objective of this study is to investigate the effects of oral bacteria and their supernatants on the SCAP cytokine profile and evaluate how they may influence the osteogenic and immunogenic potential of SCAP included in tooth arrangement and repair. They have the potential to be used in regenerative dentistry procedures such as dental pulp, periodontal tissue, and dental tissue regeneration. The potential of these DSCs in treating dental diseases and restoring oral health is being continuously explored through ongoing clinical studies.

### Organoids (3Dcel culture) used in dental stem cell research

Organoids are three-dimensional (3D) systems for cell culture that imitate the composition and capabilities of human organs. They are created from stem cells. They offer a more physiologically appropriate habitat closer to in vivo circumstances, which substantially improves over typical two-dimensional (2D) cultures. Because of their capacity to facilitate more accurate modelling of human tissues, organoids are useful in a number of scientific categories, including drug development, personalized medicine, and cancer research <sup>54</sup>. Within the final decade and recently researched, organoid innovation has shown to be a useful tool for studying tissue science and the advancement of tissue transplanted in a 3D extracellular matrix platform throughout the last ten years of research 55. Organoids offer scientists an exciting opportunity to study signaling pathways and edit genomes in a body-like environment. However, they also have several limitations that mimic real-life challenges <sup>56,57</sup>.

Nano scaffold are designed to mimic the natural extracellular matrix, providing structural support for cell attachment, growth, and differentiation that simulates the Extracellular Matrix (ECM), like Matrigel, is used to seed the distinct essential tissue test, which contains stem cells either as single cells or enclosed within cell clusters. These developments are characterized by a multidisciplinary approach that combines engineering principles with biomedical sciences, resulting in improved medical equipment 54. The scaffold is subsequently modified using a specific combination of developmental components that simulate stem cell specialization <sup>58,59</sup>. In addition, organoids can be employed to examine the interplay between stem cells and immune cells, encompassing the function of mediators/cytokines in simulated conditions 60,61. Researchers may learn more about how different organs form and function in certain areas, such as the oral and maxillofacial tract, by using organoids as experimental models. These microscopic systems also have enormous promise for disease modelling, which may help with medication discovery and provide insight into the relationships between hosts and pathogens. Additionally, they provide incredible chances for studying the complexities of organogenesis and regenerative medicine <sup>62</sup>. Furthermore, organoids exhibit the capacity to replicate an important proportion of the complexity, unique characteristics, and genetic markers found in the original tissues and organs 62. Organoids are 3D cell culture systems used for basic science, medicine, and drug development that replicate in vivo environments. DSC can be used in bone and dental tissue engineering, and they show great promise for tissue regeneration <sup>63</sup>.

Overcoming the limitations in the availability and culture-ability of epithelial stem cells from essential tissue, serial extension (passaging) of organoid societies provides an invaluable and reliable source, while minimizing any negative traits that may be present 64. The construction of epithelial organoid models is typically performed without prior segregation of the epithelial (stem) cells from the extracted tissue sample, as a result of mesenchymal cells being unable to thrive within the specific culture conditions employed and being readily discarded 65. Organoids are threedimensional (3D) cultures that have been utilized in DSC inquiries to investigate tooth-specific science <sup>66</sup>. This occurs when cells or tissue fragments, such as stem cells, are implanted into a three-dimensional scaffold within the extracellular matrix 55.

# Some preferences for utilizing organoids in DSC research are as follows

DSCs have the ability to summarize the complex structure of tissue, making them organoids, as compared to traditional 2D cell cultures, provide a more physiologically significant exhibition. They can replicate the intricate engineering and biological intuition present in healthy tissues, enabling analysts to consider tooth-specific science and improvement with greater precision <sup>66,67</sup>. They can demonstrate genetic control, allowing for the hereditary modification of organoids and allowing analysts to consider the effects of specific traits on tooth development and function. This makes it possible to investigate the inherited factors associated with oral diseases and possible beneficial targets <sup>68</sup>.

Long-term culture and development are the main parameters in DSC culture so organoids can be extended and kept up in culture for expanded periods, permitting long-term consideration and experimentation. This empowers analysts to observe the development and behaviour of DSCs over time <sup>67</sup>. Figure 3 shows following the extraction of hDPSCs from T75 flasks, they were seeded into 24 well plates for two-dimensional culture, with subsequent transfer into drop formations on 24 suspension cell culture plates.

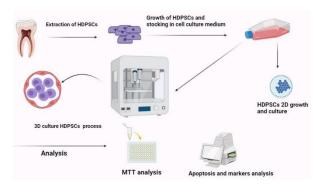


Figure 3. Extraction of a third molar involved a procedure that assured aseptic conditions during the removal of its pulp. Through collagenase digestion, the extracted pulp tissue provided suitable material for growing human Dental Pulp Stem Cells (hDPSCs) employing seeding. These cultured hDPSCs underwent passaging in T75 flasks for utilization in 3D culturing experiments and future cell stocks. Following the extraction of hDPSCs from T75 flasks, they were seeded into 24 well plates for two-dimensional culture, with subsequent transfer into drop formations on 24 suspension cell culture plates. Following a 4-day growth period, hDPSCs were collected from all scenarios including the 2D and 3D medium, and various analyses were carried out.

Patient-derived organoids can be utilized to study personal varieties in tooth improvement and illness. This permits personalized pharmaceutical approaches, where medications can be custom-made to an individual's particular hereditary cosmetics and characteristics <sup>16</sup>

# Organoids be used to study dental diseases and disorders

Organoids are three-dimensional (3D) structures that can model the architecture and function of human organs. Pluripotent stem cells, adult stem cells, or somatic cells serve as a source for them and can self-organize to reproduce the complexity of many tissues. Such self-organization allows the maintenance of key functional, structural, and biological properties similar to the original organ they represent <sup>64,69</sup>.

Organoids can be utilized to ponder dental diseases and disorders. Organoids give a 3D culture framework that permits complex intuition between cells, the stream of flagging atoms and supplements, and selforganization, which makes a difference in researchers demonstrating and getting dental physiology 70,71. Organoids can be created by using cells that have been taken from individuals with particular dental conditions, such as jaw and facial cancer and dental dysplasia <sup>72</sup>. This permits analysts to imitate the infection in a controlled research facility setting and think about its components and movement 70. Also, organoids can be utilized to analyze the morphological and useful changes related to salivary organ dysfunctions, such as pathobiology, inflammation, and regenerative pharmaceuticals 73.

Organoids have moreover been utilized to study the intestinal epithelial barrier in fiery bowel maladies, and to address respiratory diseases, they have been generated from either endogenous lung epithelial stem/pro-

genitor cells or pluripotent stem cells <sup>74</sup>. Organoids derived from human tooth tissue have been created to investigate tooth epithelial stem cell science and epithelium-mesenchyme intelligence, Consequently, this breakthrough could lead to advancements in tooth-regenerative therapy as a viable option instead of relying solely on synthetic materials for implants <sup>75</sup>. Organoids can be utilized to consider different dental diseases and disorders, including maxillofacial tumours, tooth dysplasia <sup>70</sup>, and salivary gland dysfunctions <sup>71</sup>.

### Scaffold cell culture

The utilization of biomaterials as scaffolds aids in the growth and differentiation of dental stem cell: DSC-based regenerative treatments rely heavily on the use of biomaterials <sup>76</sup>. They are utilized as scaffolds by producing nano polymers electrospinning 77 to support the development and diffraction of DSCs and cells within dental tissue have the remarkable capability to imitate the natural extracellular matrix. Figure 4 showcases a depiction in the form of a schematic diagram. This presentation highlights the seeding of DSCs on a hybrid scaffold and emphasizes their capability to differentiate, underscoring the potential for tissue regeneration in clinical applications, allowing for biodegradable and biocompatible properties that foster a conducive environment for the attachment, growth, proliferation, and differentiation of stem cells in a three-dimensional shape <sup>78,79</sup>. Various types of biocompatibility and biodegradability biomaterials have been utilized in dental stem cell-based regenerative treatments as scaffolds. such as gelatin methacrylate (GelMA) hydrogel that create covalent bonds. These naturally derived materials are biocompatible and non-toxic to cells, allowing them to interact harmoniously with each other 80. Cell culture compatible, the biodegradable copolymers PGA/PLGA exhibit unique design capabilities and property 80 and collagen-nanoHA/OPS bio composite nano scaffolds provide a supportive structure for the DSCs growth and differentiation and showed a high rate of cell proliferation. The adhesion, migration, and spatial distribution of hDFMSC within the 3D matrix

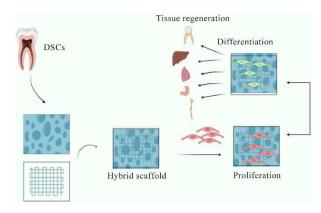


Figure 4. A depiction in the form of a schematic diagram exhibiting the seeding of DSCs on a hybrid scaffold and highlighting their differentiation capability is presented, emphasizing the potential clinical applications for tissue regeneration.

were enhanced by the increased deposition of extracellular bone matrix (osteopontin) and high ALP activity, suggesting early osteogenic differentiation. The existence of human OPN was the cause of this improvement 79,81,82

It has been discovered and experimented that using engendered scaffolds made of Poly (\(\epsilon\)-caprolactone) (PCL), Chitosan (CS), and MTA increases the expression of angiogenic markers in (DISCs). In order to promote the angiogenesis and osteogenesis of hDPSCs, scaffolds made of biopolymers from MTA/polycaprolactone (PCL) composites coated with Caffeic Acid (CA) that affected the physicochemical and biological features which were used 83. Successful results were obtained when researchers used 3D scaffolds containing MTA/polycaprolactone (PCL) composites coated with CA to stimulate angiogenic and osteogenic processes in hDPSCs 84. This may help DSCs differentiate more effectively into specific cell types, such as odontoblasts and cementoblasts 85. The possibility of using 3D porous chitosan scaffolds to treat spinal cord injuries is suggested by their capacity to induce neural development in hDPSCs 86.

Biomaterials can have a different organic direct effect on differentiation of DSCs in the human body. A few biomaterials can advance the multiplication and separation of DSCs, whereas others can inhibit their separation. By selecting suitable biomaterial coordinating the differentiation process of DSCs can lead them to develop into specific cell types 87. Mechanosensory pathways that regulate stem cell differentiation and separation to facilitate cell migration can be activated by them in DSC pulp. Gaining knowledge of these pathways can help improve the differentiation and multiplication of DSCs and thus can assist in the regeneration of tissue 85,87. Biomaterials can be falsely synthesized and designed to supply a steady structure for the development and the ability to differentiate DSCs leads to the generation of numerous quantities that are valuable in regenerative treatments <sup>79,82</sup>.

Extraction of DSCs is made possible by the distinct developmental phases in regeneration observed in diverse dental tissues such as dental pulp, apical papilla, dental follicle, tooth germ, deciduous teeth, periodontal ligament, and gingiva <sup>88,89</sup>. Their potential as an excellent bone platform material has led to the exploration of a wide variety of biomaterials, including polymers, ceramics, and composites <sup>89</sup>.

### What distinct categories of biomaterials are applied in dental tissue engineering

In dental tissue engineering, we have utilized distinctive types of biomaterials that are constructed from nano size to increase efficiency <sup>90</sup> as platforms to back the regeneration and development of dental cells and tissues in culture. Different groups can be formed based on the composition of biomaterials, properties, or applications to show the best performance in medical engineering. Here are a few of the distinctive types

of biomaterials commonly utilized in dental tissue engineering <sup>79</sup>.

For this model, we can use collagen a characteristic protein found within the extracellular matrix of different tissues, counting dental tissues. This substance stands out for its ability to be biocompatible, biodegradable, and possess remarkable cell adhesion properties. Collagen platforms can give structural back and advance cell connection, expansion, and differentiation 91

Utilized in dental stem cell culture, poly(lactic) acid (PLA) is an example of a biomaterial. PLA is an artificial polymer that demonstrates both biodegradability and biocompatibility feature. Its mechanical qualities and ability to promote cell proliferation have led to its widespread use in dental tissue regeneration <sup>91,92</sup>. One suitable biomaterial for use in DSC culture is hydrogel, which are composed of hydrophilic polymers properties in a three-dimensional configuration, have a large capacity to absorb and store water. Their features are similar to those of the common extracellular network to growth cells on it, and they offer an environment that is favorable to tissue regeneration and cell proliferation. For dental pulp tissue engineering, hydrogels such like fibrin and self-assembling peptides have been investigated in vitro 91,93. Due to the fact bioactive ceramics like hydroxyapatite and tricalcium phosphate can improve bone and dental regeneration, much research has been done on them. In addition to providing a scaffold framework, ceramics can release ions that promote tissue structure and promote cell activity 89. Considering they can be transferred minimally extensively, injectable biomaterials including hydrogels and nanofibers have gained attention in dental tissue engineering. These substances can be injected into the malformation area and used to frame as a scaffold there, supporting tissue regeneration region 94. There are several researches in the process of growing DSCs, a variety of biomaterials have been employed as scaffolds. The specific application and desired outcomes play a major role in determining the biomaterial selection in dental tissue engineering <sup>95</sup>. Analysts are still researching and developing modern biomaterials with cutting-edge properties to increase the efficacy of dental tissue regener-

For instance, a clinical trial explored the use of nanostructured materials to deliver growth factors that promote the proliferation and differentiation of DSCs *in situ*. The outcomes suggested improved healing rates and tissue integration compared to traditional methods <sup>27</sup>.

Research has shown that nanoparticles can improve the delivery and efficacy of DSC therapies by enhancing targeted drug delivery, such as growth factors responsible for tissue regeneration. Preclinical studies have proven this very effective; thus, translation into clinical trials is considered in order to evaluate its efficacy in human patients <sup>96</sup>.

Compared with conventional materials, the production costs associated with integrating nanoparticles into dental applications are frequently greater. This is because creating and working with materials at the nanoscale requires specialized techniques that might be resource-intensive and call for cutting-edge technology. Innovation and economy of scale must be maintained. Although nanotechnology has tremendous potential for improving dental treatments, both patients and professionals are quite concerned about how affordable these cutting-edge technologies will be. For these advancements to be widely used, it is imperative that they be made accessible <sup>97</sup>.

Nanoparticle-based therapeutics are intrinsically complicated because of their multicomponent nature. Designing and engineering such nanoparticles requires extra care and attention to detail, as even slight changes in composition can significantly impact the physicochemical properties and biological behavior of these particles. For example, reproducibility and consistency of the product characteristics during manufacturing are a great challenge, since most synthesis and characterization techniques applied for nanoparticles are sophisticated ones 98. The manufacturing complexities to arrive at appropriate liposomal formulations for drug delivery are multidimensional. For instance, the liposomes have to be engineered to realize defined sizes and surface properties that enhance drug loading and targeting efficacy. Variability in production processes may result in batch-to-batch inconsistencies, further complicating regulatory approval and clinical translation 99.

Quality control is crucial but difficult to ensure in the production of nanomedicine. Advanced analytical techniques are necessary for adequate nanoparticle formulation characterization. In addition, quality control measures common for conventional drugs may be inadequate, and testing methods have to be more sophisticated to evaluate not only physical but also biological properties of nanoparticles. For instance, such variability in the size and surface charge of nanoparticles may influence their therapeutic efficacy and safety profiles. Hence, the development of robust quality control protocols by using multiple orthogonal characterization techniques becomes essential for ensuring consistent product quality <sup>98,99</sup>.

# Signaling pathways culture in the regeneration of dental stem cell

Signaling pathways are significant within the laboratory's culture and the improvement in the comprehension and regulation of these pathways holds promise for enhancing the regenerative potential of dental pulp and contributing to breakthroughs in techniques for dental tissue regeneration. Signaling transduction pathways are included in regulating the regenerative potential of DSCs <sup>100</sup>. Expound modulation of signaling transduction spatially and transiently through growth factors in dish and conveyance can advance basic and

useful dental pulp regeneration <sup>100</sup>. The pivotal component in the pulp DSC signaling pathway, ascertainable at the basal region of the developing dental root, is none other than the apical papilla. This structure contains multipotent stem cells that exhibit a remarkable capacity for diverse differentiation processes 101. Another factor in the signaling pathway is cell-cell communication which plays a critical part in DSC culture regeneration. Dental Mesenchymal Stem Cells (DMSCs) have appeared to deliver a secretum, counting solvent factors and extracellular vesicles, that can initiate paracrine activity and advanced tissue regeneration 102. During the expansion phase, it has been noted that cellcell communication involving STAT3/Oct-4/Sox2 signaling enhances the pluripotency of dental pulp cells and periodontal ligament cells, making them highly favourable for dental tissue regeneration 103.

The establishment of protocols for isolation, cryopreservation, and quality control has facilitated the utilization of cryopreserved stem cells derived from dental tissue <sup>104</sup>. In addition, scientific studies have revealed that DPSCs hold the capacity to develop into useful neurons in controlled environments, showcasing their potential role in cellular treatments for neuro-degenerative disorders <sup>105</sup>.

DSCs and their secretions have shown promise for tissue regeneration, and advanced research appears to be targeting their components of cell-to-cell communication. There are, diverse signaling pathways that influence DSC differentiation in different ways (Table 2) <sup>88</sup>. Understanding these pathways can offer assistance in creating novel techniques for dental tissue regeneration. One instance is cytokines or developmental substances like Stroma cell-Derived factor (SDF), Fibroblast Growth Factor (FGF), BMP, and Vascular Endothelial Growth Factor (VEGF) by itself, Wnt can individually stimulate DSCs in terms of movement, proliferation, odontogenic differentiation, pro-angiogenesis, and pro-neurogenesis potentials 100. Another signaling pathway is the STAT3/Oct-4/Sox2 signaling pathway, through STAT3/Oct-4/Sox2 signaling, the co-culture systems demonstrated a substantial enhancement in the expression of Oct-4, Sox2, and STAT3, leading to improved dental pulp cells and periodontal ligament cells via cell-cell communication 103.

### Immunomodulating factor in DSC for regeneration

DSC culture exhibits an exceptional feature known as immunomodulation that can contribute to making strides in their viability in dental regenerative medication <sup>110</sup>. In the treatment of neurodegenerative diseases, neural injuries, cartilage defects, and bone repair, the DPSC-derived secretum demonstrates efficacy due to its ability to regulate neuroprotective, anti-inflammatory, antiapoptotic, and angiogenic processes with its assortment of biomolecules <sup>111</sup>. The inborn properties of self-renewal, immunomodulation, proangiogenic potential, and multilineage power have intrigued DSCs for their application within the regeneration of dental

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Table 2. Comparing the va	arious signaling	patnways involve	a in the regenera	ation of dental stem cells

Signaling pathway	Key functions	Components	Impact on dental stem cells
Wnt signaling pathway (106)	Migration Proliferation Odontogenic differentiation Pro-angiogenesis Pro-neurogenesis	Wnt proteins	Stimulates dental stem cells individually, enhancing regenerative potential
STAT3/Oct-4/Sox2 pathway (107)	Enhances pluripotency Promotes cell-cell communication	STAT3 Oct-4 Sox2	Increases expression of key pluripotency factors in dental pulp and periodontal ligament cells
Cytokines and growth ractors (42)	Differentiation and regeneration	SDF FGF BMP VEGF	Influences various aspects of dental stem cell behavior and differentiation
Cell-cell communication (108)	Paracrine signaling	Secretome from DMSCs	Facilitates tissue regeneration and enhances regenerative capabilities
Apical papilla (109)	Contains multipotent stem cells	Stem cells in the apical region	Key component in signaling pathways for dental pulp stem cell differentiation

tissues <sup>112</sup>. DPSCs have been employed in the dental field in a novel approach to treat oviduct wounds through DPSCs-based therapy through immunomodulation and angiogenesis *in vivo* <sup>113</sup>. While inflammation is an essential component of wound healing, an excess of it may cause damage to tissues and hinder the healing process. Immunomodulating factors are instrumental in regulating inflammation by altering macrophage behaviours and suppressing the release of proinflammatory cytokines <sup>114</sup>. It has been revealed that the microbiota that inhabits the human body could be the modulator of MSCs, including DSCs, as well <sup>115</sup>.

Understanding the interactions between microbiota and DSCs is crucial for making significant advancements in the field of Regenerative Medicine 115. Later investigations have looked for ways to optimize DSC functionality including the ability to work in vitro and in vivo, preparation of culture conditions, exposure to mechanical and physical stimuli, preconditioning by cytokines and growth variables, and genetic modification of dental stem cells 112,116. Using growth factors such as SDF, FGF, BMP, VEGF and Wnt, DSCs can efficiently regenerate lost pulp through mechanisms such as increased migratory capacity and increased proliferation and differentiation potential. The extracts obtained from DPSCs contain a diverse array of biomolecules. These extracts have shown effectiveness in treating neurodegenerative diseases, nerve damage, cartilage defects, and bone repair by modulating neuroprotective, anti-inflammatory, anti-apoptotic, and angiogenic mechanisms 100.

# Immunomodulatory mechanisms in dental stem cells: a pathway to enhanced regeneration

Nanomaterials for DSC regeneration provide a longterm opportunity as well as a challenge. On one hand, their use can improve regenerative outcomes, but on the other hand, their interactions with the immune system and possible effects after long exposure have to be taken into consideration. Further studies will be necessary to relate these interactions and enable nanotechnology to be safely translated into dental clinical practice. According to the British Journal of Cancer in 2022, the long-term follow-up has been higher in clinical trials; according to them, many toxicities take years to show up. Presently, this stands as a wide perspective for therapies that involve either the stem cells or the nanotechnology treatment, thereby establishing that in all these therapies; patients need to be followed up over time due to the probability of capturing complications arising much later 117. Although preliminary results are encouraging, there is a pressing need for more longterm studies that assess the interactions of these therapies with biological systems over an extended period, concerning complications such as chronic inflammation or tumor formation 118. For example, engineered nanoparticles might be perceived as harmful by the immune system, causing an unintended immune response that could compromise their therapeutic efficacy. The biomolecular corona concept refers to how, upon entrance into the body, proteins adsorb onto nanoparticles, which may alter their biological identity and immune responses. This interaction can either enhance or suppress immune recognition and response. Understanding this phenomenon becomes important for designing safer nanomaterials for dental applications 119.

# Cytokines and growth factors are gearing up to enhance MSCs' abilities

Cytokines, in addition to developmental variables, can influence tissues through paracrine effects on MSCs. Cytokines can interact between development components by their receptors on the surface and dynamic signal cell survival. Migration and differentiation still take place despite the repression of T cell and B cell activation, as well as dendritic cell differentiation, while simultaneously impairing the cytolytic po-

tential of Natural Killer (NK) cells and enhancing regulatory T cell differentiation 120. In any case, the immunomodulatory impact of MSCs isn't continuously accomplished but requires incitement by provocative variables, cells can be influenced by various substances, including IFN-γ, TNF-α, and IL-1β <sup>121</sup>. cytokine preparation is recommended to be a supplemental atomic procedure to cultivate the helpful potential of MSCs and contribute to building up an affable microenvironment for dental tissue repair. Specifically, dental MSC-derived secretum/conditioned medium, which contains bioactive particles Exhibiting attributes like anti-inflammatory, antiapoptotic, angiogenic, osteogenic, and neurogenic effects, these arbiters propose an exceptional cell-free regenerative and therapeutic strategy that surpasses parent cells in various aspects 112,122.

The research findings indicate that cytokines and growth factors can play a role in improving the effectiveness of mesenchymal stem cells (MSCs) in regenerative medicine: below shows the ability of cytokine and growth factors:

- 1. Pulling MSCs towards the injured area: The engineered MSCs utilize the CXC chemokine receptor 4 (CXCR4)-stromal cell-derived factor  $1\alpha$  (SDF1 $\alpha$ ) axis to migrate towards the site of injury, promoting vasculogenesis through the secretion of cytokines and growth factors by paracrine secretion and improved neuroprotective approaches to optimize supply, foster recovery of axon connectivity, and boost behavioural ability  $^{123}$ .
- 2. Extraordinary cytokine profiles: The therapeutic potential of stem cells from SHED and DPSCs may be influenced by their unique cytokine profiles <sup>19</sup>.
- 3. The significance of MSCs preconditioning: Adapting MSCs before therapeutic use by exposing them *ex vivo* to hypoxia, inflammatory stimulus, or alternative factors/conditions is a custom strategy designed to enable MSCs survival in hostile surroundings and amplify their role in regulating local immune responses <sup>124</sup>.
- 4. The paracrine effect is observed: The cytokine production capability of MSCs facilitates their role in regulating the immune microenvironment and stimulating the repair process of endothelial cells, fibroblasts, and tissue precursor cells in wounds such as the cytokine interleukin-6 (IL-6), the chemokine CCL2, the angiogenic factor VEGF, and the FGF <sup>125</sup>.

### **Discussion**

Regenerative therapy is a specialization of pharmacy and medicine. It mainly focuses on the employment of stem cells, besides other modalities like nanoscience and biotechnology, to stimulate intrinsic regenerative procedures of the body, hence restoring functionalities to damaged tissues or organs. It is also used in improving function and pain following injury. In this review article, we have focused on DSCs for the regeneration of dental cells and other cells in humans. The appropriateness of tissue-engineered therapies relying on the

multipotent regenerative abilities of this cells is accompanied by significant challenges, as growth factors and epigenetic components are crucial for preserving their multipotency, while being susceptible to a range of natural and environmental factors. Current evidence highlights the positive outcomes associated with select regenerative therapies; nevertheless, to provide further support, additional data must be gathered through standardized therapies and further studies. Organoids (3D cell culture) and nano scaffolds are also being explored as potential tools for regenerative therapies.

Strategies based on nanotechnology are being investigated to improve the safety and effectiveness of stem cell treatments for dental regeneration. Although they are still in early stages, there are several crucial clinical uses. To assess their safety, effectiveness, and clinical transferability for everyday dental uses, further extensive research and clinical trials are required. Widespread clinical acceptance will depend on overcoming obstacles with regard to immunological rejection, regulatory permission, and the source of stem cells. Future generations of respiratory therapeutics will be made possible by an understanding of the mechanisms governing these cells' behavior and interactions with their habitats. Cell therapy is an alternate strategy in regenerative medicine that shows potential. It is well known that developmental factors such as cytokine parameters and extracellular vesicle formation are expected to mediate the regenerative response in cell therapy through paracrine actions. Although clinical trials have shown stem cell therapy to be safe, it has the potential to change into illness. In contrast, stem cells and nanotechnology have great potential to improve dental regeneration and treatment; there are substantial ethical and legal barriers that must be addressed before these technologies can be fully utilized. The area of regenerative dentistry is quickly developing, and when new tools and methods appear, it is critical to implement safety measures to guarantee their responsible and ethical application.

However, considering the promising results from animal models and early-phase human trials, dental applications of these approaches are still lacking profound long-term data concerning efficacy and safety. Possible risks of tumorigenesis and immune responses from the manipulation of stem cells demand a lengthy follow-up period in clinical studies to confirm such modalities.

Ethical concerns on the source of stem cells are significant in DSC therapy, whereby one has to distinguish between embryonic and adult stem cells. This discussion encompasses the moral implications of sourcing these cells, as well as the strategies implemented to address these challenges. For instance, the use of embryonic stem cells has raised deep ethical concerns mainly because human embryos must be destroyed to acquire these cells. Critics maintain that this act is a violation of the moral status accorded to em-

bryos, which a number of people argue should be accorded protection as a form of human life. This is a contention that evokes heated debates on personhood and the ethical use of embryos for research and therapy. Obtaining informed consent from the donors is essential but complex, since many people attach emotions and morals to reproductive materials. In contrast, adult stem cells, including those from dental pulp and other tissues, generally pose fewer ethical issues. These could normally be retrieved with less moral complaint, as no embryo destruction is involved in the process.

The most common sources for adult stem cells are bone marrow, adipose tissue, and umbilical cord blood. These have become widely accepted sources both in research study and clinically, as they do not raise the same ethical dilemmas as embryonic stem cells.

The collection and use of adult stem cells are governed by established regulations that ensure respect for informed consent and donor rights, thereby reducing the risk of exploitation or coercion. Setting standardized guidelines on stem cell research may ensure that ethical issues are considered within the framework of the study. The guidelines should cover best practices in sourcing, handling, and utilization of the stem cells, hence encouraging responsible research. Public transparency should be improved through public information and education on stem cell research. These would include open forums, workshops, and clearly available resources that explain the science supporting stem cell therapies, the ethical issues involved, and the potential benefits. Engaging various stakeholders will help build trust and understanding. Efforts must be directed to make any progress with stem cell technologies available to diverse populations. This would relate to disparities in access to healthcare and the affordability of treatments created through stem cell research. Strategies may involve policy advocacy for equitable healthcare distribution and support for underserved communities.

# Conclusion

DSCs present a viable path for regenerative medicine and regeneration. These cells are notable for having intriguing characteristics such as multipotent separable ability, clonogenicity, and self-renewal that make them appropriate for tissue regeneration. Organoids are three-dimensional (3D) communities that have been utilized to reflect tooth-specific science and regenerative medicine in the field of DSCs research. They offer a valuable resource for researching prospective tooth regeneration, learning about tooth homeostasis, and researching human dental stem cell science. Nano scaffolds, cell treatment, signaling pathways, and epigenetic and immunomodulating approaches are efforts made to explore and improve the regenerative potential of DSCs. Numerous inquiries and articles have discussed the potential use of DSCs in tooth repair, reclamation, and recovery through tissue engineering and regenerative pharmaceutical applications. DSCs have the potential to repair and regenerate dental pulp, periodontal tissue and other oral structures. DSCs have to be examined for their great potential for non-dental applications, such as the advancement of stem cell-based solutions for heart, brain and bone repair. We also found that components such as epigenetics and immunological impacts as well as development variables in tissue engineering play a principal part in the development and differentiation of these cells. To conclude, DSCs emerge as a promising strategy for advancing regenerative medicine in both dentistry and its broader applications. The rising demand for regenerated materials fuels ongoing clinical trials.

#### **Conflict of Interest**

Authors declare no conflict of interest.

#### References

- Egger A, Tomic-Canic M, Tosti A. Advances in Stem Cell-Based Therapy for Hair Loss. CellR4 Repair, Replace Regen Reprogram 2020;8:e2894.
- Jenkins TL, Little D. Synthetic scaffolds for musculoskeletal tissue engineering: cellular responses to fiber parameters. NPJ Regen Med 2019;4(1):15.
- 3. Lee S, Jin G, Jang JH. Electrospun nanofibers as versatile interfaces for efficient gene delivery. J Biol Eng 2014;8(1):30.
- Foltan R, Petr J, Streblov J, Guha A. Orthodontic treatment enhanced using PRP: An alternative to Orthognathic surgery? J Dent Oral Disord Ther 2019;7(1):1-8.
- Ramaswamy Reddy S, Reddy R, Babu Nc, Ashok G. Stem-cell therapy and platelet-rich plasma in regenerative medicines: A review on pros and cons of the technologies. J Oral Maxillofac Pathol 2018;22(3):367-74.
- Campbell JD, Milenkovic U, Usta MF, Albersen M, Bivalacqua TJ. The good, bad, and the ugly of regenerative therapies for erectile dysfunction. Transl Androl Urol 2020;9(S2):S252-S261.
- 7. Auersperg V, Trieb K. Extracorporeal shock wave therapy: an update. EFORT Open Rev 2020;5(10):584-92.
- Moritz CT, Ambrosio F. Regenerative Rehabilitation: Combining Stem Cell Therapies and Activity-Dependent Stimulation. Pediatr Phys Ther Off Publ Sect Pediatr Am Phys Ther Assoc 2017;29 Suppl 3(Suppl 3 IV STEP 2016 CONFERENCE PROCEEDINGS):S10-S15.
- Perez-Terzic C, Childers MK. Regenerative Rehabilitation. Am J Phys Med Rehabil 2014;93(11 Suppl 3):S73-S78.
- Perczel-Kovách K, Hegedűs O, Földes A, et al. STRO-1 positive cell expansion during osteogenic differentiation: A comparative study of three mesenchymal stem cell types of dental origin. Arch Oral Biol 2021;122:104995.
- Chalisserry EP, Nam SY, Park SH, Anil S. Therapeutic potential of dental stem cells. J Tissue Eng 2017;8: 204173141770253.
- 12. Leyendecker Junior A, Gomes Pinheiro CC, Lazzaretti

- Fernandes T, Franco Bueno D. The use of human dental pulp stem cells for in vivo bone tissue engineering: A systematic review. J Tissue Eng 2018;9: 204173141775276.
- Shilpa P, Sultana N, Kaul R, Bhat S. Stem cells: Boon to dentistry and medicine. Dent Res J (Isfahan) 2013;10(2): 149-54.
- Smojver I, Katalinić I, Bjelica R, et al. Mesenchymal Stem Cells Based Treatment in Dental Medicine: A Narrative Review. Int J Mol Sci 2022;23(3):1662.
- Peterson A, Nair LS. Hair Follicle Stem Cells for Tissue Regeneration. Tissue Eng Part B Rev 2022;28(4):695-706.
- Baena AR, Casasco A, Monti M. Hypes and Hopes of Stem Cell Therapies in Dentistry: a Review. Stem Cell Rev Rep 2022;18(4):1294-308.
- 17. Chen Y, Huang H, Li G, Yu J, Fang F, Qiu W. Dentalderived mesenchymal stem cell sheets: a prospective tissue engineering for regenerative medicine. Stem Cell Res Ther 2022;13(1):38.
- Botelho J, Cavacas MA, Machado V, Mendes JJ. Dental stem cells: recent progresses in tissue engineering and regenerative medicine. Ann Med 2017;49(8):644-51.
- Kang CM, Shin MK, Jeon M, Lee YH, Song JS, Lee JH. Distinctive cytokine profiles of stem cells from human exfoliated deciduous teeth and dental pulp stem cells. J Dent Sci 2022;17(1):276-83.
- Bates K, Gallicchio VS. Dental Stem Cell Banking and Applications of Dental Stem Cells for Regenerative Medicine. InNovel Perspectives of Stem Cell Manufacturing and Therapies 2021 Jan 19. IntechOpen.
- Alhazzazi T, Alghamdi F. Clinical applications of dental stem cells in modern regenerative medicine: A systematic review with updates. Niger J Clin Pract 2021;24 (4):457.
- Tsutsui T. Dental Pulp Stem Cells: Advances to Applications. Stem Cells Cloning 2020;13:33-42.
- Abdullah MF, Abdullah SF, Omar NS, Mahmood Z, Fazliah Mohd Noor SN, Kannan TP, et al. Proliferation rate of stem cells derived from human dental pulp and identification of differentially expressed genes. Cell Biol Int 2014;38(5):582-90.
- Ahangar P, Mills SJ, Cowin AJ. Mesenchymal Stem Cell Secretome as an Emerging Cell-Free Alternative for Improving Wound Repair. Int J Mol Sci 2020;21(19): 7038.
- 25. Park JH, Gillispie GJ, Copus JS, Zhang W, Atala A, Yoo JJ, et al. The effect of BMP-mimetic peptide tethering bioinks on the differentiation of dental pulp stem cells (DPSCs) in 3D bioprinted dental constructs. Biofabrication 2020;12(3):035029.
- Gnanasegaran N, Govindasamy V, Mani V, Abu Kasim NH. Neuroimmunomodulatory properties of DPSCs in an in vitro model of Parkinson's disease. IUBMB Life 2017;69(9):689-99.
- 27. Zhang Y, Liu J, Zou T, Qi Y, Yi B, Dissanayaka WL, et al. DPSCs treated by TGF-β1 regulate angiogenic sprouting of three-dimensionally co-cultured HUVECs and DPSCs through VEGF-Ang-Tie2 signaling. Stem

- Cell Res Ther 2021;12(1):281.
- 28. Karamzadeh R, Eslaminejad MB. Dental-related stem cells and their potential in regenerative medicine. InRegenerative Medicine and Tissue Engineering 2013 May 22. IntechOpen.
- 29. Zhang M, Jiang F, Zhang X, Wang S, Jin Y, Zhang W, et al. The Effects of Platelet-Derived Growth Factor-BB on Human Dental Pulp Stem Cells Mediated Dentin-Pulp Complex Regeneration. Stem Cells Transl Med 2017;6 (12):2126-34.
- Yang JW, Zhang YF, Sun ZY, Song GT, Chen Z. Dental pulp tissue engineering with bFGF-incorporated silk fibroin scaffolds. J Biomater Appl 2015;30(2):221-9.
- Lymperi S, Ligoudistianou C, Taraslia V, Kontakiotis E, Anastasiadou E. Dental Stem Cells and their Applications in Dental Tissue Engineering. Open Dent J 2013;7 (1):76-81.
- Chen K, Xiong H, Huang Y, Liu C. Comparative analysis of in vitro periodontal characteristics of stem cells from apical papilla (SCAP) and periodontal ligament stem cells (PDLSCs). Arch Oral Biol 2013;58(8):997-1006.
- Iwayama T, Sakashita H, Takedachi M, Murakami S. Periodontal tissue stem cells and mesenchymal stem cells in the periodontal ligament. Jpn Dent Sci Rev 2022;58: 172-8.
- 34. Zhu W, Liang M. Periodontal Ligament Stem Cells: Current Status, Concerns, and Future Prospects. Stem Cells Int 2015;2015:972313.
- 35. Yang C, Du XY, Luo W. Clinical application prospects and transformation value of dental follicle stem cells in oral and neurological diseases. World J Stem Cells 2023; 15(4):136-49.
- 36. Wei X, Guo S, Liu Q, Liu L, Huo F, Wu Y, et al. Dental Follicle Stem Cells Promote Periodontal Regeneration through Periostin-Mediated Macrophage Infiltration and Reprogramming in an Inflammatory Microenvironment. Int J Mol Sci 2023;24(7):6353.
- 37. Bi R, Lyu P, Song Y, Li P, Song D, Cui C, et al. Function of Dental Follicle Progenitor/Stem Cells and Their Potential in Regenerative Medicine: From Mechanisms to Applications. Biomolecules 2021;11(7):997.
- 38. Zhang J, Ding H, Liu X, Sheng Y, Liu X, Jiang C. Dental Follicle Stem Cells: Tissue Engineering and Immunomodulation. Stem Cells Dev 2019;28(15):986-94.
- Kim JH, Irfan M, Hossain MA, George A, Chung S. BDNF/TrkB Is a Crucial Regulator in the Inflammation-Mediated Odontoblastic Differentiation of Dental Pulp Stem Cells. Cells 2023;12(14):1851.
- Mahdani FY, Khansa SP, Pongsumae D, Yastuti WT, Wardana MA, Surboyo MD, et al. The Viability of Osteoblasts against SHED Metabolites and EGCG for Biomaterial Osteogenesis. Research Journal of Pharmacy and Technology. 2024 Aug 1;17(8):4015-21.
- 41. Ye G, Li C, Xiang X, Chen C, Zhang R, Yang X, et al. Bone Morphogenetic Protein-9 Induces PDLSCs Osteogenic Differentiation through the ERK and p38 Signal Pathways. Int J Med Sci 2014;11(10):1065-72.

- 42. Zhou L, Zhao S, Xing X. Effects of different signaling pathways on odontogenic differentiation of dental pulp stem cells: a review. Front Physiol 2023;14:1272764.
- 43. Yakimanskiy AA, Kaskevich KI, Zhukova E V, Berezin IA, Litvinova LS, Chulkova TG, et al. Synthesis, Photo-and Electroluminescence of New Polyfluorene Copolymers Containing Dicyanostilbene and 9,10-Dicyanophenanthrene in the Main Chain. Materials (Basel) 2023;16 (16):5592.
- 44. Abedian Z, Jenabian N, Moghadamnia AA, Zabihi E, Pourbagher R, Hossein-Nataj H, et al. A comparative study on immunophenotypic characterization and osteogenic differentiation of human mesenchymal stromal cells derived from periodontal ligament and gingiva. J Periodontol 2020;91(9):1194-202.
- 45. Fernandes M, Ataide I. Nonsurgical management of periapical lesions. J Conserv Dent 2010;13(4):240-5.
- Song WP, Jin LY, Zhu MD, Wang H, Xia DS. Clinical trials using dental stem cells: 2022 update. World J Stem Cells 2023;15(3):31-51.
- 47. Usha GV. Perception about Future Prospects of Dental Stem Cell Therapy among House Surgeons and Postgraduate Students: A Cross-Sectional Survey. International Journal of Research Publication and Reviews 2023;4(9):378-82.
- Santelia M, Gallicchio VS. The Use Of Dental Stem Cell Therapy In The Treatment Of Gum Deterioration. J Stem Cell Res 2023;4(2):1-7.
- 49. Choi H, Park KH, Jung N, Shim JS, Moon HS, Kim HJ, et al. In Vivo Study for Clinical Application of Dental Stem Cell Therapy Incorporated with Dental Titanium Implants. Materials (Basel) 2021;14(2):381.
- Smeda M, Galler KM, Woelflick M, Rosendahl A, Moehle C, Lenhardt B, et al. Molecular Biological Comparison of Dental Pulp- and Apical Papilla-Derived Stem Cells. Int J Mol Sci 2022;23(5):2615.
- Zafari J, Jouni FJ, Nikzad F, Esmailnasab S, Javan ZA, Karkehabadi H. Combination of Dental-Capping Agents with Low Level Laser Therapy Promotes Proliferation of Stem Cells from Apical Papilla. Photobiomodul Photomed Laser Surg 2023;41(1):3-9.
- 52. Phothichailert S, Nowwarote N, Fournier BPJ, Trachoo V, Roytrakul S, Namangkalakul W, et al. Effects of decellularized extracellular matrix derived from Jagged1-treated human dental pulp stem cells on biological responses of stem cells isolated from apical papilla. Front Cell Dev Biol 2022;10:948812.
- 53. Zymovets V, Razghonova Y, Rakhimova O, Aripaka K, Manoharan L, Kelk P, et al. Combined Transcriptomic and Protein Array Cytokine Profiling of Human Stem Cells from Dental Apical Papilla Modulated by Oral Bacteria. Int J Mol Sci 2022;23(9):5098.
- 54. de Dios-Figueroa GT, Aguilera-Marquez JDR, Camacho-Villegas TA, Lugo-Fabres PH. 3D Cell Culture Models in COVID-19 Times: A Review of 3D Technologies to Understand and Accelerate Therapeutic Drug Discovery. Biomedicines 2021;9(6):602.
- 55. Cox B, Laporte E, Vennekens A, Kobayashi H, Nys C, Van Zundert I, et al. Organoids from pituitary as a novel

- research model toward pituitary stem cell exploration. J Endocrinol 2019;240(2):287-308.
- 56. Yang S, Hu H, Kung H, Zou R, Dai Y, Hu Y, et al. Organoids: The current status and biomedical applications. MedComm 2023;4(3):e274.
- Suarez-Martinez E, Suazo-Sanchez I, Celis-Romero M, Carnero A. 3D and organoid culture in research: physiology, hereditary genetic diseases and cancer. Cell Biosci 2022;12(1):39.
- 58. Boretto M, Cox B, Noben M, Hendriks N, Fassbender A, Roose H, et al. Development of organoids from mouse and human endometrium showing endometrial epithelium physiology and long-term expandability. Development 2017;144(10):1775-86.
- Sato T, Vries RG, Snippert HJ, van de Wetering M, Barker N, Stange DE, et al. Single Lgr5 stem cells build crypt-villus structures in vitro without a mesenchymal niche. Nature 2009;459(7244):262-5.
- 60. Chenchula S, Kumar S, Babu S. Comparative Efficacy of 3Dimensional (3D) Cell Culture Organoids Vs 2Dimensional (2D) Cell Cultures Vs Experimental Animal Models In Disease modeling, Drug development, And Drug Toxicity Testing. Int J Curr Res Rev 2019;11(24): 11-17.
- Wang Z, Yu T, Hou Y, Zhou W, Ding Y, Nie H. Mesenchymal Stem Cell Therapy for ALI/ARDS: Therapeutic Potential and Challenges. Curr Pharm Des 2022;28(27): 2234-40.
- 62. Singh B, Mal G, Verma V, Tiwari R, Khan MI, Mohapatra RK, et al. Stem cell therapies and benefaction of somatic cell nuclear transfer cloning in COVID-19 era. Stem Cell Res Ther. 2021;12(1):283.
- 63. Kim W, Gwon Y, Park S, Kim H, Kim J. Therapeutic strategies of three-dimensional stem cell spheroids and organoids for tissue repair and regeneration. Bioact Mater 2023;19(March 2022):50-74.
- 64. Zhao Z, Chen X, Dowbaj AM, Sljukic A, Bratlie K, Lin L, et al. Organoids. Nat Rev Methods Prim 2022;2(1):94.
- Schutgens F, Clevers H. Human Organoids: Tools for Understanding Biology and Treating Diseases. Annu Rev Pathol Mech Dis 2020;15(1):211-34.
- 66. Hermans F, Hemeryck L, Bueds C, Torres Pereiro M, Hasevoets S, Kobayashi H, et al. Organoids from mouse molar and incisor as new tools to study tooth-specific biology and development. Stem Cell Reports 2023;18(5): 1166-81.
- Nikonorova VG, Chrishtop VV, Mironov VA, Prilepskii AY. Advantages and Potential Benefits of Using Organoids in Nanotoxicology. Cells 2023;12(4):610.
- 68. Jeong SY, Lee S, Choi WH, Jee JH, Kim HR, Yoo J. Fabrication of Dentin-Pulp-Like Organoids Using Dental-Pulp Stem Cells. Cells 2020;9(3):642.
- 69. de Jongh D, Massey EK; VANGUARD consortium; Bunnik EM. Organoids: a systematic review of ethical issues. Stem Cell Res Ther 2022;13(1):337.
- 70. Gao X, Wu Y, Liao L, Tian W. Oral Organoids: Progress and Challenges. J Dent Res 2021;100(5):454-63.
- 71. Wang Y, Sun Y. Engineered organoids in oral and

- maxillofacial regeneration. iScience 2023;26(1):105757.
- Rossi M, Alviano F, Righi S, Sabattini E, Agostinelli C. Three-dimensional models: a novel approach for lymphoma research. J Cancer Res Clin Oncol 2022;148(4): 753-65.
- Yoshimoto S, Yoshizumi J, Anzai H, Morishita K, Okamura K, Hiraki A, et al. Inhibition of Alk signaling promotes the induction of human salivary-gland-derived organoids. Dis Model Mech 2020 Sep 28;13(9): dmm045054.
- Li Y, Wu Q, Sun X, Shen J, Chen H. Organoids as a Powerful Model for Respiratory Diseases. Stem Cells Int 2020;2020:5847876.
- Hemeryck L, Lambrichts I, Bronckaers A, Vankelecom H. Establishing Organoids from Human Tooth as a Powerful Tool Toward Mechanistic Research and Regenerative Therapy. J Vis Exp 2022;(182).
- Lukomskyj AO, Rao N, Yan L, Pye JS, Li H, Wang B, et al. Stem Cell-Based Tissue Engineering for the Treatment of Burn Wounds: A Systematic Review of Preclinical Studies. Stem Cell Rev Reports 2022;18(6): 1926-55.
- 77. Semnani D, Naghashzargar E, Hadjianfar M, Dehghan Manshadi F, Mohammadi S, et al. Evaluation of PCL/chitosan electrospun nanofibers for liver tissue engineering. Int J Polym Mater Polym Biomater 2017;66(3): 149-57.
- Sevari SP, Ansari S, Moshaverinia A. A narrative overview of utilizing biomaterials to recapitulate the salient regenerative features of dental-derived mesenchymal stem cells. Int J Oral Sci 2021;13(1):22.
- Bupphathong S, Quiroz C, Huang W, Chung PF, Tao HY, Lin CH. Gelatin Methacrylate Hydrogel for Tissue Engineering Applications—A Review on Material Modifications. Pharmaceuticals (Basel) 2022;15(2):171.
- Costa AC, Alves PM, Monteiro FJ, Salgado C. Interactions between Dental MSCs and Biomimetic Composite Scaffold during Bone Remodeling Followed by In Vivo Real-Time Bioimaging. Int J Mol Sci 2023;24(3):1827.
- 81. Salgado CL, Barrias CC, Monteiro FJM. Clarifying the Tooth-Derived Stem Cells Behavior in a 3D Biomimetic Scaffold for Bone Tissue Engineering Applications. Front Bioeng Biotechnol 2020;8(June):1-15.
- 82. Tien N, Lee JJ, Lee AKX, Lin YH, Chen JX, Kuo TY, et al. Additive Manufacturing of Caffeic Acid-Inspired Mineral Trioxide Aggregate/Poly-ε-Caprolactone Scaffold for Regulating Vascular Induction and Osteogenic Regeneration of Dental Pulp Stem Cells. Cells 2021;10 (11):2911.
- 83. Alipour M, Aghazadeh Z, Hassanpour M, Ghorbani M, Salehi R, Aghazadeh M. MTA-Enriched Polymeric Scaffolds Enhanced the Expression of Angiogenic Markers in Human Dental Pulp Stem Cells. Petrenko Y, ed. Stem Cells Int 2022;2022:1-9.
- 84. Granz CL, Gorji A. Dental stem cells: The role of biomaterials and scaffolds in developing novel therapeutic strategies. World J Stem Cells 2020;12(9):897-921.
- 85. Zheng K, Feng G, Zhang J, Xing J, Huang D, Lian M, et

- al. Basic fibroblast growth factor promotes human dental pulp stem cells cultured in 3D porous chitosan scaffolds to neural differentiation. Int J Neurosci 2021;131(7):625-33.
- 86. Aguilar-Ayala FJ, Aguilar-Pérez FJ, Nic-Can GI, Rojas-Herrera R, Chuc-Gamboa G, Aguilar-Pérez D, et al. A Molecular View on Biomaterials and Dental Stem Cells Interactions: Literature Review. Appl Sci 2022;12(12): 5815.
- 87. Staniowski T, Zawadzka-Knefel A, Skośkiewicz-Malinowska K. Therapeutic Potential of Dental Pulp Stem Cells According to Different Transplant Types. Molecules 2021;26(24):7423.
- 88. Ercal P, Pekozer GG. A Current Overview of Scaffold-Based Bone Regeneration Strategies with Dental Stem Cells. Adv Exp Med Biol 2020;1288:61-85.
- 89. Mirshekari M, Ghomi AB, Mehravaran A. Smart terbinafine recent nano-advances in delivery of terbinafine. Nanomedicine J 2021;8(4):241-54.
- 90. Galler KM, Brandl FP, Kirchhof S, Widbiller M, Eidt A, Buchalla W, et al. Suitability of Different Natural and Synthetic Biomaterials for Dental Pulp Tissue Engineering. Tissue Eng Part A 2018;24(3-4):234-44.
- 91. Castañeda-Rodríguez S, González-Torres M, Ribas-Aparicio RM, Del Prado-Audelo ML, Leyva-Gómez G, Gürer ES, et al. Recent advances in modified poly (lactic acid) as tissue engineering materials. J Biol Eng 2023;17 (1):21.
- 92. Dou XQ, Feng CL. Amino Acids and Peptide-Based Supramolecular Hydrogels for Three-Dimensional Cell Culture. Adv Mater 2017;29(16):1604062.
- Haugen HJ, Basu P, Sukul M, Mano JF, Reseland JE. Injectable Biomaterials for Dental Tissue Regeneration. Int J Mol Sci 2020;21(10):3442.
- 94. Farag MM. Recent trends on biomaterials for tissue regeneration applications: review. J Mater Sci 2023;58(2): 527-58.
- 95. Li X, Wang Y, Huang D, Jiang Z, He Z, Luo M, et al. Nanomaterials Modulating the Fate of Dental-Derived Mesenchymal Stem Cells Involved in Oral Tissue Reconstruction: A Systematic Review. Int J Nanomedicine 2023 Sep 21;18:5377-5406.
- 96. Hakim LK, Yari A, Nikparto N, Mehraban SH, Cheperli S, Asadi A, et al. The current applications of nano and biomaterials in drug delivery of dental implant. BMC Oral Health 2024;24(1):126.
- 97. Desai N. Challenges in Development of Nanoparticle-Based Therapeutics. AAPS J 2012;14(2):282-95.
- 98. Joseph T, Kar Mahapatra D, Esmaeili A, Piszczyk Ł, Hasanin MS, Kattali M, et al. Nanoparticles: Taking a Unique Position in Medicine. Nanomaterials 2023;13(3): 574.
- 99. Liang C, Liao L, Tian W. Stem Cell-based Dental Pulp Regeneration: Insights From Signaling Pathways. Stem Cell Rev Reports 2021;17(4):1251-63.
- 100. Driesen RB, Gervois P, Vangansewinkel T, Lambrichts I. Unraveling the Role of the Apical Papilla During Dental Root Maturation. Front Cell Dev Biol 2021;9:

665600.

- 101. Chouaib B, Cuisinier F, Collart-Dutilleul PY. Dental stem cell-conditioned medium for tissue regeneration: Optimization of production and storage. World J Stem Cells 2022;14(4):287-302.
- 102. Peng Z, Liu L, Zhang W, Wei X. Pluripotency of Dental Pulp Cells and Periodontal Ligament Cells Was Enhanced through Cell-Cell Communication via STAT3/ Oct-4/Sox2 Signaling. Yuan Q, ed. Stem Cells Int 2021; 2021:8898506.
- 103. Khaseb S, Orooji M, Pour MG, Safavi SM, Eghbal MJ, Rezai Rad M. Dental stem cell banking: Techniques and protocols. Cell Biol Int 2021;45(9):1851-65.
- 104. Gervois P, Struys T, Hilkens P, Bronckaers A, Ratajczak J, Politis C, et al. Neurogenic Maturation of Human Dental Pulp Stem Cells Following Neurosphere Generation Induces Morphological and Electrophysiological Characteristics of Functional Neurons. Stem Cells Dev 2015;24(3):296-311.
- 105. Amir M, Jeevithan L, Barkat M, et al. Advances in Regenerative Dentistry: A Systematic Review of Harnessing Wnt/ $\beta$ -Catenin in Dentin-Pulp Regeneration. Cells 2024;13(13):1153.
- 106. Galoczova M, Coates P, Vojtesek B. STAT3, stem cells, cancer stem cells and p63. Cell Mol Biol Lett 2018;23 (1):12.
- 107. Kumar A, Kumar V, Rattan V, Jha V, Bhattacharyya S. Secretome Cues Modulate the Neurogenic Potential of Bone Marrow and Dental Stem Cells. Mol Neurobiol 2017;54(6):4672-82.
- 108. Liu Q, Gao Y, He J. Stem Cells from the Apical Papilla (SCAPs): Past, Present, Prospects, and Challenges. Biomedicines 2023;11(7):2047.
- 109. Aydin S, Şahin F. Stem Cells Derived from Dental Tissues. Adv Exp Med Biol 2019;1144:123-32.
- 110. Kichenbrand C, Velot E, Menu P, Moby V. Dental Pulp Stem Cell-Derived Conditioned Medium: An Attractive Alternative for Regenerative Therapy. Tissue Eng Part B Rev 2019;25(1):78-88.
- 111. Zhang SY, Ren JY, Yang B. Priming strategies for controlling stem cell fate: Applications and challenges in dental tissue regeneration. World J Stem Cells 2021; 13(11):1625-46.
- 112. Luo L, Xing Z, Liao X, Luo Y, Ai Y, He Y, et al. Dental pulp stem cells-based therapy for the oviduct injury via immunomodulation and angiogenesis in vivo. Cell Prolif 2022;55(10):e13293.
- 113. Xie C, Luo M, Chen M, Wang M, Qu X, Lei B.

- Bioactive Poly(octanediol-citrate-polyglycol) Accelerates Skin Regeneration through M2 Polarization Immunomodulating and Early Angiogenesis. Adv Healthc Mater 2022;11(10):e2101931.
- 114. Tu H, Xiao E, Liu O. Taking Microbiota into Consideration in Mesenchymal Stem Cell Research. J Dent Res 2022;101(8):880-6.
- 115. Andrukhov O, Behm C, Blufstein A, Rausch-Fan X. Immunomodulatory properties of dental tissue-derived mesenchymal stem cells: Implication in disease and tissue regeneration. World J Stem Cells. 2019;11(9): 604-17.
- 116. Cuzick J. The importance of long-term follow up of participants in clinical trials. Br J Cancer 2023;128(3): 432-8.
- 117. Yamada Y, Nakamura-Yamada S, Konoki R, Baba S. Promising advances in clinical trials of dental tissue-derived cell-based regenerative medicine. Stem Cell Res Ther 2020;11(1):175.
- 118. Palmieri V, Caracciolo G. Tuning the immune system by nanoparticle–biomolecular corona. Nanoscale Adv 2022;4(16):3300-8.
- 119. Song N, Scholtemeijer M, Shah K. Mesenchymal Stem Cell Immunomodulation: Mechanisms and Therapeutic Potential. Trends Pharmacol Sci 2020;41(9):653-64.
- 120. Ren G, Zhang L, Zhao X, Xu G, Zhang Y, Roberts AI, et al. Mesenchymal Stem Cell-Mediated Immunosuppression Occurs via Concerted Action of Chemokines and Nitric Oxide. Cell Stem Cell 2008;2(2):141-50.
- 121. El Moshy S, Radwan IA, Rady D, Abbass MMS, El-Rashidy AA, Sadek KM, et al. Dental Stem Cell-Derived Secretome/Conditioned Medium: The Future for Regenerative Therapeutic Applications. Stem Cells Int 2020;2020:7593402.
- 122. Wang Z, Wang Y, Wang Z, Gutkind JS, Wang Z, Wang F, et al. Engineered Mesenchymal Stem Cells with Enhanced Tropism and Paracrine Secretion of Cytokines and Growth Factors to Treat Traumatic Brain Injury. Stem Cells 2015;33(2):456-67.
- 123. Saparov A, Ogay V, Nurgozhin T, Jumabay M, Chen WCW. Preconditioning of Human Mesenchymal Stem Cells to Enhance Their Regulation of the Immune Response. Stem Cells Int 2016;2016:3924858.
- 124. Liu C, Xu Y, Lu Y, Du P, Li X, Wang C, et al. Mesenchymal stromal cells pretreated with proinflammatory cytokines enhance skin wound healing via IL-6-dependent M2 polarization. Stem Cell Res Ther 2022;13 (1):414.