

# Is There Any Association Between the *MEF2A* Gene Changes and Coronary Artery Disease?

Soodeh Omid<sup>1</sup>, Serwa Ghasemi<sup>2</sup>, Samira Kalayinia<sup>3</sup>

<sup>1</sup> Department of Genetic, Faculty of Advanced Medical Technologies, Golestan University of Medical Sciences, Gorgan, Iran

<sup>2</sup> Department of Biology, School of Basic Sciences, Islamic Azad University Research Tehran Branch, Tehran, Iran

<sup>3</sup> Cardiogenetic Research Center, Rajaie Cardiovascular Medical and Research Center, Iran University of Medical Sciences, Tehran, Iran

Received: 11 Mar. 2020; Accepted: 18 Jul. 2020

**Abstract-** Coronary artery disease (CAD) is a common multifactorial disease with a high rate of morbidity and mortality worldwide. The *MEF2A* gene transcription factor belongs to the myocyte enhancer factor-2 (MEF2) family and is involved in critical processes such as calcium-dependent signaling pathways and cardiac development. Although the variants of the *MEF2A* gene were studied in different CAD and myocardial infarction (MI) populations, the reality of this gene association with CAD is still unclear. This study reports the first *in silico* investigation on *MEF2A* variants. All reported variants in CAD/MI patients were collected from eleven countries. Their pathogenicity and variant position conservation were surveyed by online prediction tools, including Mutation-Taster, Polyphen-2, PROVEAN, SIFT, CADD, and GERP. *In silico* analysis did not confirm the pathogenic effect of 21-bp deletion, which was introduced as a monogenic cause of CAD. c.704C>A (p.S235Y), c.812C>G (p.P271R), c.836C>T (p.P279L) and c.848G>A (p.G283D) missenses, c.1315C>T (p.R439X) nonsense, and seven out-of-frame deletions were predicted as disease-causing variants. Although some variants of the *MEF2A* gene affect protein structure, the *MEF2A* variation studies in CAD/MI patients and *in silico* analysis do not approve the association and pathogenicity of *MEF2A* variants in the familial/sporadic CAD.

© 2020 Tehran University of Medical Sciences. All rights reserved.

*Acta Med Iran* 2020;58(8):366-375.

**Keywords:** Coronary artery disease; Myocyte enhancer factor-2 (*MEF2A*); *In silico* analysis

## Introduction

According to recent World Health Organization (WHO) reports, cardiovascular diseases (CVD) with 17.9 million deaths per year, is the primary cause of death worldwide. Coronary artery disease (CAD) is one of the CVDs with defects in heart vessels (1). CAD is the cause of one-third of deaths in people older than 35 years (2). Various modifiable risk factors can result in this condition, including hypertension, high blood cholesterol levels, smoking, diabetes, overweight or obesity, lack of physical activity, unhealthy diet, stress, and conventional risk factors such as age, gender, family history, and race (3). Several genes have been considered to have a role in the monogenic form of CAD, comprising *LDLR*, *ApoB-100*, *ARH* and *PCSK9*, *ApoA1*, *ABCA1*, and *LCAT*, *LPL*, *ApoC-II*, and *ABCG5/8* genes which are involved in lipid metabolism. Moreover, some other genes that have no direct effects on plasma lipid levels include *LRP6*,

*CYP27A1*, *ST6GALNAC5*, and *MEF2A* (4).

The myocyte enhancer factor-2 (MEF2) transcription factors family with four proteins, *i.e.*, MEF2-A, -B, -C, and -D, mediates calcium-dependent signaling pathways in the various process such as division, differentiation, and cell death (5). The *MEF2A* gene, which is located in the 99,565,417 to 99,716,466 bp interval on chromosome 15q26.3, has thirteen transcripts, nine of which are protein-coding. The longest transcript with 5824 bp and 11 coding exons encodes a protein with 499 amino acids (Ensemble accession number: ENSG00000068305). MEF2A, in cooperation with other MEF2 transcription factors, regulates the differentiation of cardiac muscle cells (6), whereas MEF2A plays a critical role in the post-natal heart without supporting of other MEF2 isoforms. As Naya *et al.*, indicated, mice with a deficiency in the *MEF2A* gene died in the first week of life (7). The first report of the role of *MEF2A* in CAD was from a family with thirteen patients. In this family, seven amino acid

**Corresponding Author:** S. Kalayinia

Cardiogenetic Research Center, Rajaie Cardiovascular Medical and Research Center, Iran University of Medical Sciences, Tehran, Iran  
Tel: +98 21 23923148, Fax: +98 21 22663213, E-mail address: samira.kalayi@yahoo.com

deletions in exon 11 resulted in CAD with an autosomal dominant pattern (8). In another study, the sequencing of the *MEF2A* gene in 300 premature CAD patients and a healthy group revealed only one missense variant in one of the CAD patients and the specific seven amino acid deletions in one of 300 unaffected individuals. Further screening of 1500 additional individuals without CAD by this team indicated two cases with seven amino acid deletions and no association with CAD in their family analysis (9). Given the contradictory results of studies in different populations, the conformity of this transcription factor role in CAD/MI requires more investigation. This study is the first to analyze by *in silico* analysis all *MEF2A* reported variants, including substitutions, deletions, and insertions.

## Materials and Methods

### Collection of *MEF2A* gene variations

Due to what has been concluded in some publications about the *MEF2A* role in CAD etiology, we selected this gene for more investigation of its variations. “*MEF2A*” and “*MEF2A* variations” terms were used for our search. Variations which had been reported in different countries were collected from PubMed (<https://www.ncbi.nlm.nih.gov/pubmed/>), Google Scholar, and Google in general.

### Investigation of variants frequency

The frequency of all collected *MEF2A* variants was achieved from the 1000 Genomes Project (10) (IGSR; <http://www.internationalgenome.org/>), and Exome Aggregation Consortium (11) (ExAC; <http://exac.broadinstitute.org/>).

### Variants evaluation

All reported variants in CAD/MI patients were analyzed to determine any possible effects of them, by computational prediction tools including Mutation Taster (<http://www.mutationtaster.org/>) (12), Polymorphism Phenotyping (PolyPhen-2; <http://genetics.bwh.harvard.edu/pph/>) (13), Sorting Intolerant From Tolerant (SIFT; <http://sift.bii.a-star.edu.sg/>) (14), Protein Variation Effect Analyzer (PROVEAN; <http://provean.jcvi.org/index.php>) (15), Combined Annotation Dependent Depletion (CADD; <https://cadd.gs.washington.edu/>) (16), Rare Exome Variant Ensemble Learner (REVEL) (17); and the conservation scores of variant position was calculated by Genomic Evolutionary Rate Profiling (GERP; <http://mendel.stanford.edu/SidowLab/downloads/gerp/>)

online (18).

**Mutation Taster** has the highest accuracy in comparison of other tools (SIFT, PROVEAN, PolyPhen-2) in effect prediction of the insertion/deletion, intronic, and intron-exon splice site changes (12).

**PolyPhen-2** predicts the effects of amino acid changes on protein stability and function based on structural properties and conservation profiles. The PolyPhen-2 score ranges from 0.0 to 1.0; variants with scores in the range between 0.95-1 are damaging (13).

**SIFT** classifies substitutions as intolerant and tolerant changes in genomic and protein levels based on sequence homology with similar sequences. Moreover, SIFT updated for insertions/deletions effect prediction recently. The SIFT score ranges from 0.0 to 1.0; substitutions with scores <0.05 have been considered deleterious (14).

**PROVEAN** measures the similarity of the query protein sequence to its homologous protein sequence with and without variation. PROVEAN provides a prediction of variants in genomic and protein levels for single/multiple substitutions and in-frame insertions/deletions. The default score threshold is set at -2.5; substitutions with probabilities <0.05 are deleterious (15).

**CADD** reports a single measure as the C-score (ranging from 1 to 99) due to multiple annotations. CADD is a useful tool for the prediction of single-nucleotide and short insertions/deletion variants (16).

**REVEL** is a method for predicting of missense variant pathogenicity. This method integrates scores from 13 prediction tools. REVEL scores range from 0 to 1, and higher scores are predicted more likely pathogenic (17).

**GERP** calculates a conservation score for any nucleotide positions by using the multiple alignments of 35 mammal's genome sequences. It ranges from -12.3 to 6.17, i.e., 6.17 is the most conservation score (18).

It is designed to predict the functional consequences of not only amino acid substitutions but also intronic and synonymous alterations, short insertion and/or deletion (indel) mutations, and variants spanning intron-exon borders

It is designed to predict the functional consequences of not only amino acid substitutions but also intronic and synonymous alterations, short insertion and/or deletion (indel) mutations, and variants spanning intron-exon borders

It is designed to predict the functional consequences of not only amino acid substitutions but also intronic and synonymous alterations, short insertion and/or deletion (indel) mutations, and variants spanning intron-exon

borde.

### MEF2A gene-gene interactions

Interactions of the *MEF2A* gene were analyzed by the Multiple Association Network Integration Algorithm (GeneMANIA; <https://genemania.org/>). GeneMANIA finds related genes to the query gene based on various biological databases, and its output is categorized according to Protein-protein interaction (Physical Interaction), similar gene expression levels (co-expression), same domains in the protein structure (shared protein domains), the association of genes when perturbed genes affect each other (genetic interaction), genes products are participating in the same pathway (pathway), gene products are found in the same cellular location or tissue (co-localization), prediction of functional association between gene products especially through orthology (predicted) and other relationships such as phenotype correlations, disease information, and chemical genomics data (other) (19).

## Results

### Variants reports in various populations

Investigation of different populations from twelve countries, including American, Scandinavian, and Japanese, Chinese, Spanish, Italian, Iranian, Sicilian, Saudi, Turkish, Irish, and German CAD and/or MI patients indicate twenty-four variations in the coding region of the *MEF2A* gene. The greatest diversity of substitution variants, i.e., eight different ones, have been observed in Chinese and Italian populations (Table 1). The contribution of missense, synonym, and nonsense variants are eleven, eight, and one substitution, respectively, as well as there, are one deletion and insertion variants (Table 2). In the coding sequence of this gene, two polymorphic regions were identified as the poly-glutamine and poly-proline regions (Figure 1).

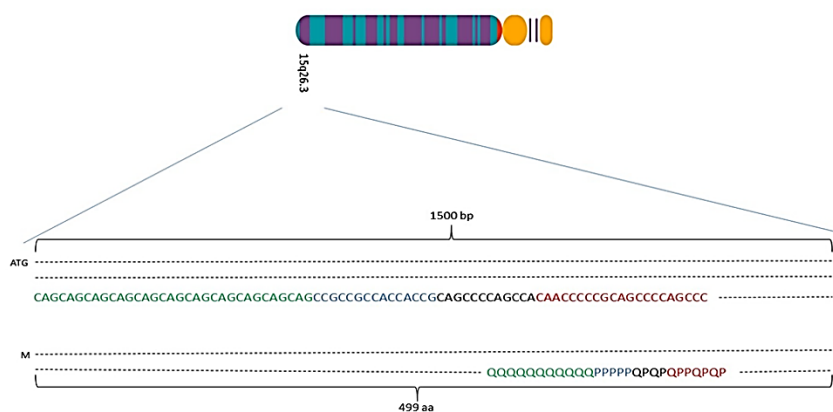
**Table 1. Distribution of reported variants in various populations.**

No.	Mutation	Protein change	America	Scandinavia	Japan	China	Spain	Italy	Iran	Cecil	Saudi	Turkey	Ireland	Germany	Disease
1	c.704C>A	p.S235Y						10 (29)							MI
2	c.736A>G	p.S246G						10 (29)							MI
3	c.788A>G	p.N263S	207 (30)					3 (29)							CAD/ MI
4	c.812C>G	p.P271R						10 (29)							MI
5	c.835C>G	p.P279A						11 (29)							MI
6	c.836C>T	p.P279L	207 (30)				483 (23)	3 (29)							CAD / MI
7	c.848G>A	p.G283D	207 (30)												CAD / MI
8	c.860C>G	p.P287R						1 (29)							MI
9	c.867T>C	p.N289N			379 (21)	257 (20)			52 (25)						CAD/MI
10	c.873G>A	p.Q291Q							52 (25)						CAD
11	c.1055C>T	p.S352L	300 (9)												CAD
12	c.1224A>G	p.P408P									1186 (35)				CAD
13	c.1227G>A	p.S409S					483 (23)				1186 (35)				CAD / MI
14	c.1266_1267insCCGCAGCAG	p.422PQQins				257 (20)									CAD
15	c.1268C>A	p.P423Q				156 (37)									CAD
16	c.1275A>G	p.P425P				1045 (36)									CAD
17	c.1279C>T	p.P427S				1045 (36) 257 (20) 111 (38)									CAD

Cont Table 1.

18	c.1281G>A	p.P427P			379 (21)	1045 (36) 257 (20) 111 (38) 210 (39)								CAD/ MI
19	c.1294_1314delCAACCCCCGCAGCCCCAGCCC	p.432-438QPPQPQ del	300 (9)	13 (8)	379 (21)	257 (20) 1045 (36)	483 (23)	5 (29)	300 (45) 52 (25)	1186 (35)				CAD / MI
20	c.1315C>T	p.R439X			379 (21)									MI
21	c.1329G>T	p.G443G			379 (21)	257 (20) 1045 (36) 111 (38) 156 (37) 210 (39)	483 (23)		300 (45) 52 (25)	1186 (35)				CAD/ MI
22	c.1416A>G	p.P472P							300 (45) 52 (25)	1186 (35)				CAD
The number of different mutations in each country			5	1	5	8	3	8	4	1	4	-	-	-
The number of people were studied in each country			520	13	379	1779	483	2008	352	1079	1186	69 (24)	1494 (26)	>1700 & 23 family (27)

All MEF2A coding variants are reported according to NCBI nucleotide (NM\_005587.3) and protein (NP\_005578.2) sequences.  
CAD: coronary artery disease; MI: myocardial infarction



**Figure 1.** Nucleotide and amino acid sequence of two polymorphic regions, including poly-glutamine, poly-proline, and 21-bp deletion, according to NCBI nucleotide (NM\_005587.3) and protein (NP\_005578.2) sequences.

## MEF2A variants pathogenicity

### Missense variants

Among missense variants, eight substitutions were predicted as the disease-causing, seven damages, five deleterious, and eight had an effect on protein function by Mutation Taster, PolyPhen-2, PROVEAN, and SIFT, respectively. The c.704C>A (p.S235Y), c.812C>G (p.P271R), c.836C>T (p.P279L) and c.848G>A (p.G283D) variants were considered as the damaging changes in protein structure by all prediction tools. CADD scores of these four variants are reported as such 28.8, 29.2, 31, and 32, respectively. In addition, the

conservation score of S235Y has been predicted 5.55, and the last three variants, i.e., p.P271R, p.P279L, and p.G283D, 5.85 by GERP (Table 2).

### Nonsense variant

A nonsense variant in the MEF2A gene, c.1315C>T (p.R439X), changes the amino acid to stop codon, was predicted as a pathogenic mutation by Mutation taster and SIFT. It had a CADD score 44, which is the highest pathogenicity score among all MEF2A reported variants, and a conservation GERP score of 5.05 (Table 2).

Table 2. *In silico* analysis of reported MEF2A variants

No.	Mutation	Protein change	dbSNP	Mutation type	HGMD	Mutation taster	Polyphen_2	PROVEAN	SIFT	GERP	CADD	REVEL	ClinVar	EXAC Het/Hom	1000G Het/Hom
1	c.704C>A	p.S235Y	rs751251460	M	-	DC	PD	De	AFP	5.55	28.8	0.56	CAD,MI	1/0	0/0
2	c.736A>G	p.S246G	rs755896449	M	-	P	B	N	T	5.55	22.6	0.21	-	3/0	0/0
3	c.788A>G	p.N263S	rs121918530	M	CM043299	DC	B	N	T	-9.35	8.9	0.13	-	110/0	2/0
4	c.812C>G	p.P271R	rs776085239	M	-	DC	PD	De	AFP	5.85	29.2	0.45	CAD,MI	1/0	0/0
5	c.835C>G	p.P279A	-	M	-	DC	B	De	AFP	4.94	21.8	0.08	CAD,MI	3/0	0/0
6	c.836C>T	p.P279L	rs121918529	M	CM043300	DC	PD	De	AFP	5.85	31	0.28	-	85/0	4/0
7	c.848G>A	p.G283D	rs121918531	M	CM043301	DC	PD	De	AFP	5.85	32	0.47	-	2/0	0/0
8	c.860C>G	p.P287R	rs751751585	M	-	DC	PD	N	AFP	5.85	25.4	0.36	-	1/0	0/0
9	c.867T>C	p.N289N	rs325408	S	-	P	-	N	T	-	11.1	-	-	-	851/1108
10	c.873G>A	p.Q291Q	rs325407	S	-	P	-	N	T	-	8.6	-	-	-	315/2063
11	c.1055C>T	p.S352L	-	M	-	DC	PD	N	T	5.65	24.6	0.15	-	5/0	-
12	c.1224A>G	p.P408P	rs144461661	S	-	DC	-	N	T	-	14.7	-	-	4/0	4/0
13	c.1227G>A	p.S409S	rs3730059	S	-	DC	-	N	T	-	9.6	-	-	335/9	53/1
14	c.1266_1267insCCGCAGCAG	p.422PQ Q ins	-	I	-	P	-	N	N	-	12.1	-	-	-	-
15	c.1268C>A	p.P423Q	-	M	-	P	B	N	AFP	-5.5	0.4	0.07	-	20/0	-
16	c.1275A>G	p.P425P	-	S	-	P	-	N	T	-	0.2	-	-	1/0	-
17	c.1279C>T	p.P427S	-	M	-	P	PD	N	AFP	3.65	8.6	0.11	CAD	1/0	-
18	c.1281G>A	p.P427P	rs367780642	S	-	DC	-	N	T	-	0.02	-	-	21/0	30/0
19	c.1294_1314delCAACCCCCAGCCCCAGCCC	p.432-438QPP QPQPdel	-	D	CG035245	P	-	N	N	-	21.4	-	-	-	-
20	c.1315C>T	p.R439X	-	N	CM056644	DC	-	NA	D	5.05	44	-	-	-	-
21	c.1329G>T	p.G443G	rs325400	S	-	P	-	N	T	-	16.4	-	-	5252/0	997/736
22	c.1416A>G	p.P472P	rs34851361	S	-	P	-	N	T	-	6.4	-	-	2124/0	139/9
23	CCG/CCA del	4P or 5P	-	D	CD068109	-	-	-	-	-	-	-	CAD, MI	-	-
24	(CAG) <sub>n</sub>	Poly Q	rs3138597	RV	CE077839	-	-	-	-	-	-	-	-	-	-

All MEF2A coding variants are reported according to NCBI nucleotide (NM\_005587.3) and protein (NP\_005578.2) sequences. Variants were not classified into disease-causing and benign by CADD, GERP, and REVEL programs; the cutoff of 20, 5, and 0.5 was used for CADD, GERP, and REVEL scores respectively so that variants with CADD score  $\geq 20$  were grouped as harmful variants, GERP score  $\geq 5$  were grouped as conserved and REVEL score above 0.5 were grouped as likely disease-causing variants. M: Missense; S: Synonymous; D: Deletion; I: Insertion; N: Nonsense; RV: Repeat variations; DC: Disease-causing; P: Polymorphism; PD: Probably damaging; B: Benign; De: Deleterious; N: Neutral; NA: Not available; AFP: Affect protein function; T: Tolerated

### Synonymous variants

Although three of eight synonymous variants were predicted as the disease-causing by Mutation Taster, but all of them were Neutral and Tolerated by the last tools, i.e., PROVEAN and SIFT. The CADD scores of all synonymous variants were  $< 20$  (Table 2).

### Deletion and insertion variants

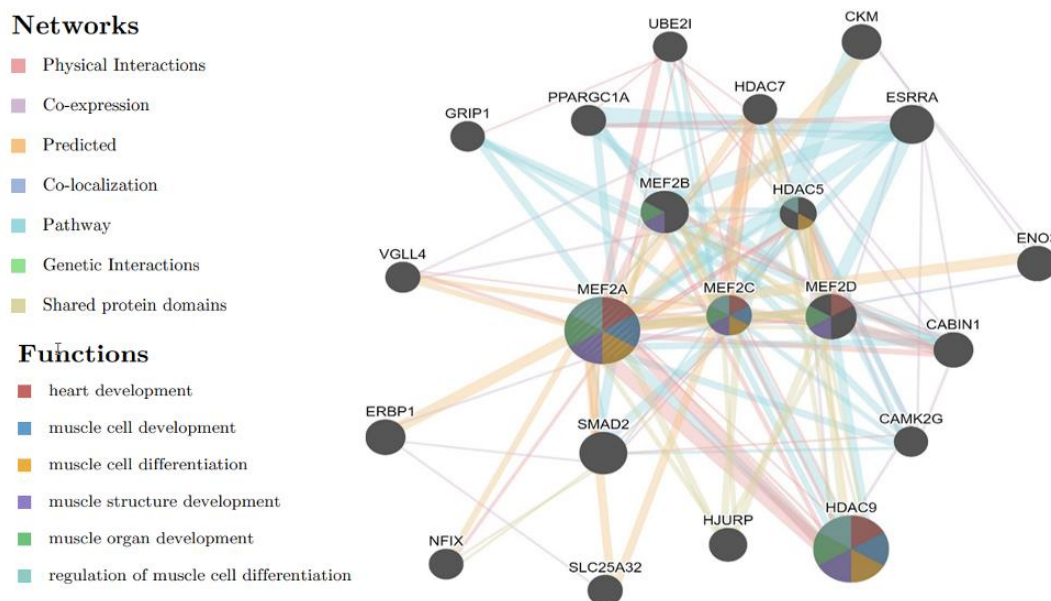
In the coding region of the MEF2A gene, one 21-bp deletion was reported for the first time as a CAD causal mutation by autosomal dominant pattern (8). Moreover, one insertion variant, including 9-bp nucleotides, was concluded in the Chinese population (20). These variants were predicted Polymorphism and Neutral by Mutation

Taster, PROVEAN, and SIFT. They had CADD scores 21.4 and 12.1, respectively. 7-amino acid deletion and 3-amino acid insertion were other variants that were located in the polymorphic areas, including 4-15 (Q)<sub>n</sub> and subsequently, 4-5 (P)<sub>n</sub> (Table 2).

There were nine out-of-frame deletions in which seven of them were frame-shift, disease-causing, and damaging by Mutation-Taster and SIFT. In addition, the CADD scores of them were >20 (Table 3).

### Gene-gene interactions

*MEF2A* gene has been present in the complex network with nineteen genes. In this network, most of the *MEF2A* gene interactions were by *MEF2D*, *MEF2C*, *MEF2B*, *SMAD2*, *HDAC9* genes (Table 4). Among them, *MEF2A*, *MEF2C*, *MEF2D*, *MEF2B*, *HDAC9*, *HDAC5* involve in muscle/heart development and differentiation (Figure 2).



**Figure 2.** Gene’s interactions and their functions are predicted by GeneMANIA based on the *MEF2A* query. Networks are represented as colored lines between genes, and functions are represented as colored circles for each gene

**Table 3. Out-of-frame deletions were reported in the *MEF2A* gene.**

No.	Mutation	Protein change	Mutation type	HGMD	Mutation taster	Poly-phen_2	PROVEAN	SIFT	CADD	Phys. location
1	c.1265_1266delAG	frameshift	Deletion	CD101281	Disease causing	-	NA	Damaging	24.2	chr15:100252741_100252742delAG
2	c.1268_1268delC	frameshift	Deletion	CD101282	Disease causing	-	NA	Damaging	22.7	chr15:100252744_100252744delC
3	c.1269_1269delG	frameshift	Deletion	CD101283	Disease causing	-	NA	Damaging	23.9	chr15:100252745_100252745delG
4	c.1270_1271delCC	frameshift	Deletion	CD101284	Disease causing	-	NA	Damaging	21.4	chr15:100252746_100252747delCC
5	c.1272_1272delG	frameshift	Deletion	CD101285	Disease causing	-	NA	Damaging	23.8	chr15:100252748_100252748delG
6	c.1273_1274delCC	frameshift	Deletion	CD101286	Disease causing	-	NA	Damaging	21.3	chr15:100252749_100252750delCC
7	c.1275_1275delA	frameshift	Deletion	CD101287	Disease causing	-	NA	Damaging	23.9	chr15:100252751_100252751delA

All *MEF2A* coding variants are reported according to NCBI nucleotide (NM\_005587.3) and protein (NP\_005578.2) sequences. Variants were not classified into disease-causing and benign by the CADD program; the cutoff of 20 was used for CADD scores, so variants with score  $\geq 20$  were grouped as harmful variants.

Table 4. Classification of related genes with MEF2A according to GeneMANIA network categories

Physical interactions	Co-expression	Predicted	Co-localization	Pathway	Genetic interaction	Shared protein domains
HDAC9	MEF2C	MEF2D	MEF2D	HDAC9		MEF2D
MEF2D		MEF2B		SMAD2		MEF2B
MEF2B		SMAD2		ESRRA		MEF2C
SMAD2		MEF2C		CABIN1		HJURP
MEF2C		SERBP1		HDAC5		
CABIN1		ENO3		PPARGC1A		
HDAC5		CKM		GRIP1		
VGLL4		SLC25A32		CAMK2G		
NFIX		VGLL4				
HDAC7		NFIX				
UBE2I		HDAC7				

CABIN1; Calcineurin Binding Protein 1, CAMK2G; Calcium/Calmodulin Dependent Protein Kinase II Gamma, CKM; Creatine Kinase, M-Type, ENO3; Enolase 3, ESRRA; Estrogen Related Receptor Alpha, GRIP1; Glutamate Receptor Interacting Protein 1, HDAC5; Histone Deacetylase 5, HDAC7; Histone Deacetylase 7, HDAC9; Histone Deacetylase 9, HJURP; Holliday Junction Recognition Protein, MEF2B; Myocyte Enhancer Factor 2B, MEF2C; Myocyte Enhancer Factor 2C, MEF2D; Myocyte Enhancer Factor 2D, NFIX; Nuclear Factor I X, PPARGC1A; PPARG Coactivator 1 Alpha, SERBP1; SERPINE1 MRNA Binding Protein 1, SLC25A32; Solute Carrier Family 25 Member 32, SMAD2; SMAD Family Member 2, UBE2I; Ubiquitin Conjugating Enzyme E2 I, VGLL4; Vestigial Like Family Member 4

## Discussion

CAD is a multifactorial disease in which genetics plays a main role with approximately 50-60% heritability. Some genes have been identified as CAD monogenic causes, such as genes involved in high LDL, TG, and low HDL and genes with no effect on plasma lipid levels, such as the *MEF2A* gene (4).

The *MEF2A* gene was introduced as an associated gene with CAD by Wang *et al.*, in 2003, in a large family with an autosomal dominant pattern. In this study, they reported the 15q26 region, which contains 93 genes as an associated locus with CAD. Because of the *MEF2A* expression in blood vessels during early embryogenesis of mouse, Wang *et al.*, analyzed only the *MEF2A* gene and identified 21-bp deletion as a causal mutation of CAD (8). Although 21-bp deletion was introduced as a pathogenic mutation for CAD due to the conformational change in the *MEF2A* structure, its association with CAD was not confirmed in other populations (21-28). Functional studies by Guella *et al.*, also showed that the 21-bp deletion did not alter the nuclear localization and transactivating properties of the *MEF2A* protein (29). Though the CADD score of this deletion was >20, its pathogenicity was not corroborated by *in silico* analysis. It is concluded that the assumption of a single-gene inheritance pattern for 21-bp deletion results from a high probability of common diseases such as CAD in individuals in a big family (9).

In 2004, the sequencing and single-strand

conformation polymorphism (SSCP) analysis of all *MEF2A* exons in 207 CAD/MI patients and 191 control by Bhagavatula *et al.*, found three missense mutations (N263S, P279L, and G283D) in only four patients (30). These three variants were reported by the ClinVar (<https://www.ncbi.nlm.nih.gov/clinvar/>) free database, which interprets relationships among variants and phenotypes (31). P279L and G283D were pathogenic mutations for the CAD/MI condition; however, N263S was a likely benign variant. Moreover, according to *in silico* analysis, P279L and G283D were pathogenic changes in the *MEF2A* gene. Although P279L and N263S were reported in Spanish and Italian CAD/MI patients (23,29), these two variants did not impair the transactivation activity of the protein (29). S235Y and P271R were other variants that were reported in Italian CAD/MI patients, and *in silico* analysis confirmed their pathogenicity, similar to other variants identified in this population that did not change the transactivation activity of *MEF2A* by transcription activation assays (29). R439X, a nonsense variant with a pathogenic effect according to our *in silico* analysis, was reported in a 70-year-old Japanese patient with MI who had two risk factors, diabetes mellitus, and smoking, but no family history of ischemic heart disease (21). Polymorphic variations of the polyQ and polyP regions of *MEF2A* were reported in various populations, some as pathogenic (32-34), and others with no association with CAD/MI (20-27,29,35-39). Deletion of one proline and changes in glutamine length in CAD patients were reported as

pathogenic variations in the Chinese population by Yuan Hong *et al.*, but they did not detect any other variations in CAD patients (32). Moreover, glutamine repeat extension, especially (CAG)<sub>9</sub> variant, was reported as a highly frequent variant associated with CAD in the Chinese population (33). In a 2016 study, a 6-bp deletion “CAGCCG” in polyQ and polyP regions was introduced for the first time in a large Chinese family with some suspected CAD/MI individuals. Interestingly, this deletion was detected in all family members including patients and healthy individuals. More investigation of the 6-bp deletion in unrelated CAD and control groups did not confirm this deletion in sporadic CAD patients (34). Several deletions were reported in a Saudi population with frame-shift consensus. Except two of these deletions, i.e., c.1265\_1266delAG and c.1270\_1271delCC, others were not associated with CAD, although they were predicted to be disease-causing variants due to >20 CADD scores (35).

*MEF2A* is one of the essential players in the transcriptional factor network, including *GATA4*, *NKX2.5*, and *Srf* genes. This network has critical roles in cardiac transcriptome regulation in cooperation with histones modifications (40). *MEF2A* interacts with *HDAC5* and *HDAC9* in the cardiac development pathway (41). Despite the significant function of the *MEF2A* gene in heart development, the results of *MEF2A* studies and *in silico* analysis did not confirm its association with familial and sporadic CAD. Furthermore, the 15q26 region was reported as a risk region for CAD patients by genome-wide association studies (GWAS) (42,43); however, the *MEF2A* gene was not introduced as an associated gene with CAD in this region by GWAS (44). Given that CAD is a common and multifactorial disease and the difficulty in pathogenic variants determination, the current researchers believe the role of *MEF2A* in CAD/MI can be further surveyed by additional functional studies.

The *in silico* analysis of *MEF2A* variations indicated that some variants have pathogenic effects on protein structures; however, the evaluation of case-control studies in *MEF2A* association with CAD/MI and *in silico* analysis did not detect an association between this gene and familial/sporadic CAD.

## Acknowledgments

This research was provided by Rajaie Cardiovascular, Medical and Research Center (RCMRC), Tehran, Iran, Golestan University of Medical Science (GUMS), Gorgan, Iran, and Islamic Azad University Research

Tehran Branch, Tehran, Iran.

## References

1. Organization. WH. Cardiovascular diseases (CVDs). 2016.
2. Sanchis-Gomar F, Perez-Quilis C, Leischik R, Lucia A. Epidemiology of coronary heart disease and acute coronary syndrome. *Ann Transl Med* 2016;4:256.
3. Hajar R. Risk factors for coronary artery disease: historical perspectives. *Heart Views* 2017;18:109-14.
4. Dai X, Wiernek S, Evans JP, Runge M. Genetics of coronary artery disease and myocardial infarction. *World J Cardiol* 2016;8:1-23.
5. McKinsey TA, Zhang CL, Olson EN. MEF2: a calcium-dependent regulator of cell division, differentiation and death. *Trends Biochem Sci* 2002;27:40-7.
6. Pon JR, Marra MA. MEF2 transcription factors: developmental regulators and emerging cancer genes. *Oncotarget* 2016;7:2297-312.
7. Naya FJ, Black BL, Wu H, Bassel-Duby R, Richardson JA, Hill JA, et al. Mitochondrial deficiency and cardiac sudden death in mice lacking the MEF2A transcription factor. *Nat Med* 2002;8:1303-9.
8. Wang L, Fan C, Topol SE, Topol EJ, Wang QJS. Mutation of MEF2A in an inherited disorder with features of coronary artery disease. *Science* 2003;302:1578-81.
9. Altshuler D, Hirschhorn JN, Joci. MEF2A sequence variants and coronary artery disease: a change of heart? *J Clin Invest* 2005;115:831-3.
10. 1000 Genomes Project Consortium; Auton A, Brooks LD, Durbin RM, Garrison EP, Kang HM, et al. A global reference for human genetic variation. *Nature* 2015;526:68-74.
11. Lek M, Karczewski KJ, Minikel EV, Samocha KE, Banks E, Fennell T, et al. Analysis of protein-coding genetic variation in 60,706 humans. *Nature* 2016;536:285-91.
12. Schwarz JM, Cooper DN, Schuelke M, Seelow D. MutationTaster2: mutation prediction for the deep-sequencing age. *Nat Methods* 2014;11:361-2.
13. Adzhubei I, Jordan DM, Sunyaev SR. Predicting functional effect of human missense mutations using PolyPhen-2. *Curr Protoc Hum Genet* 2013;7:7-20.
14. Sim N-L, Kumar P, Hu J, Henikoff S, Schneider G, Ng PC. SIFT web server: predicting effects of amino acid substitutions on proteins. *Nucleic Acids Res* 2012;40:W452-7.
15. Choi Y, Chan AP. PROVEAN web server: a tool to predict the functional effect of amino acid substitutions and indels. *Bioinformatics* 2015;31:2745-7.
16. Kircher M, Witten DM, Jain P, O’Roak BJ, Cooper GM, Shendure J. A general framework for estimating the



## MEF2A variants in coronary artery disease

- relative pathogenicity of human genetic variants. *Nat Genet* 2014;46:310-5.
17. Ioannidis NM, Rothstein JH, Pejaver V, Middha S, McDonnell SK, Baheti S, et al. REVEL: an ensemble method for predicting the pathogenicity of rare missense variants. *Am J Hum Genet* 2016;99:877-85.
  18. Davydov EV, Goode DL, Sirota M, Cooper GM, Sidow A, Batzoglou S. Identifying a high fraction of the human genome to be under selective constraint using GERP++. *PLoS Comput Biol* 2010;6:e1001025.
  19. Warde-Farley D, Donaldson SL, Comes O, Zuberi K, Badrawi R, Chao P, et al. The GeneMANIA prediction server: biological network integration for gene prioritization and predicting gene function. *Nucleic Acids Res* 2010;38:W214-20.
  20. Dai DP, Zhou XY, Xiao Y, Xu F, Sun FC, Ji FS, et al. Structural changes in exon 11 of MEF2A are not related to sporadic coronary artery disease in Han Chinese population. *Eur J Clin Invest* 2010;40:669-77.
  21. Kajimoto K, Shioji K, Tago N, Tomoike H, Nonogi H, Goto Y, et al. Assessment of MEF2A mutations in myocardial infarction in Japanese patients. *Circ J* 2005;69:1192-5.
  22. Weng L, Kavaslar N, Ustaszewska A, Doelle H, Schackwitz W, Hébert S, et al. Lack of MEF2A mutations in coronary artery disease. *J Clin Invest* 2005;115:1016-20.
  23. González P, García-Castro M, Reguero JR, Batalla A, Ordoñez AG, Palop RL, et al. The Pro279Leu variant in the transcription factor MEF2A is associated with myocardial infarction. *J Med Genet* 2006;43:167-9.
  24. Gulec S, Ruchan Akar A, Akar N. MEF2A sequence variants in Turkish population. 2008;14:465-7.
  25. Rahatloo KI, Davaran S, Elahi E. Lack of association between the MEF2A gene and coronary artery disease in Iranian families. *Iran J Basic Med Sci* 2013;16:950-4.
  26. Horan PG, Allen AR, Hughes AE, Patterson CC, Spence M, McGlinchey PG, et al. Lack of MEF2A  $\Delta$ 7aa mutation in Irish families with early onset ischaemic heart disease, a family based study. *BMC Med Gen* 2006;7:65.
  27. Lieb W, Mayer B, Konig IR, Borwitzky I, Gotz A, Kain S, et al. Lack of association between the MEF2A gene and myocardial infarction. *Circulation* 2008;117:185-91.
  28. Maiolino G, Colonna S, Zanchetta M, Pedon L, Seccia TM, Cesari M, et al. Exon 11 deletion in the myocyte enhancer factor (MEF) 2A and early onset coronary artery disease gene in a Sicilian family. *Eur J Cardiovasc Prev Rehabil* 2011;18:557-60.
  29. Guella I, Rimoldi V, Asselta R, Ardissino D, Francolini M, Martinelli N, et al. Association and functional analyses of MEF2A as a susceptibility gene for premature myocardial infarction and coronary artery disease. *Circ Cardiovasc Genet* 2009;2:165-72.
  30. Bhagavatula MK, Fan C, Shen G-Q, Cassano J, Plow EF, Topol EJ, et al. Transcription factor MEF2A mutations in patients with coronary artery disease. *Hum Mol Genet* 2004;13:3181-8.
  31. Landrum MJ, Lee JM, Benson M, Brown G, Chao C, Chitipiralla S, et al. ClinVar: public archive of interpretations of clinically relevant variants. *Nucleic Acids Res* 2015;44:D862-8.
  32. Yuan H, Lü H-W, Hu J, Chen S-H, Yang G-P, Huang ZJ. MEF2A gene and susceptibility to coronary artery disease in the Chinese people. *Zhong Nan Da Xue Xue Bao Yi Xue Ban* 2006;31:453-7.
  33. Han Y, Yang Y, Zhang X, Yan C, Xi S, Kang J. Relationship of the CAG repeat polymorphism of the MEF2A gene and coronary artery disease in a Chinese population. *Clin Chem Lab Med* 2007;45(8):987-92.
  34. Xu DL, Tian HL, Cai WL, Zheng J, Gao M, Zhang MX, et al. Novel 6-bp deletion in MEF2A linked to premature coronary artery disease in a large Chinese family. *Mol Med Rep* 2016;14:649-54.
  35. Elhawari S, Al-Boudari O, Muiya P, Khalak H, Andres E, Al-Shahid M, et al. A study of the role of the myocyte-specific enhancer factor-2A gene in coronary artery disease. *Atherosclerosis* 2010;209:152-4.
  36. Liu Y, Niu W, Wu Z, Su X, Chen Q, Lu L, et al. Variants in exon 11 of MEF2A gene and coronary artery disease: evidence from a case-control study, systematic review, and meta-analysis. *PLOSE ONE* 2012;7:e31406.
  37. Li J, Yang J, Li W, Du R, Gui L, Tian L, et al. Study on novel mutations of MEF2A gene in Chinese patients with coronary artery disease. *Zhonghua Yi Xue Yi Chuan Xue Za Zhi* 2006;23:265-8.
  38. Dai Y, Zhang S, Wu W. Analysis of MEF2A mutations in a Chinese population with premature coronary artery disease. *Genet Test Mol Biomarkers* 2013;17:352-5.
  39. Li J, Chen H, Yang J, Li W, Du R, Tian L. MEF2A gene mutations and susceptibility to coronary artery disease in the Chinese population. *Genet Mol Res* 2014;13:8396-402.
  40. Schlesinger J, Schueler M, Grunert M, Fischer JJ, Zhang Q, Krueger T, et al. The cardiac transcription network modulated by Gata4, Mef2a, Nkx2. 5, Srf, histone modifications, and microRNAs. *PLoS Genet* 2011;7:e1001313.
  41. Chang S, McKinsey TA, Zhang CL, Richardson JA, Hill JA, Olson EN. Histone deacetylases 5 and 9 govern responsiveness of the heart to a subset of stress signals and play redundant roles in heart development. *Mol Cell Biol* 2004;24:8467-76.
  42. van der Harst P, Verweij NJCr. Identification of 64 novel genetic loci provides an expanded view on the genetic

- architecture of coronary artery disease. *Circ Res* 2018;122:433-43.
43. Nikpay M, Goel A, Won H-H, Hall LM, Willenborg C, Kanoni S, et al. A comprehensive 1000 Genomes-based genome-wide association meta-analysis of coronary artery disease. *Nat Genet* 2015;47:1121-30.
  44. Omidi S, Ebrahimzadeh F, Kalayinia S. 9P21. 3 locus; an important region in coronary artery disease: a panel approach to investigation of the etiology of coronary artery disease. *Int J Cardiovasc Pract* 2019;4:21-35.
  45. Foroughmand AM, Shahbazi Z, Galehdari H, Borujeni MP, Dinarvand P, Golabgirkhademi K. Association of MEF2A gene polymorphisms with coronary artery disease. *Iran Red Crescent Med J* 2014;16: e13533.